

# Gross and Histopathological Lesion Induced on the Gills Ofbarbus Fish by Monogenea Trematods Habitating in Western Part of Lake Tana, Amhara Region, Ethiopia

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## Research Article

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# Abstract

**Back ground:** Gill parasitic is common on cultured and wild fish. Many of these species have long been recognized to have the potential to affect the growth, fecundity and survival of hosts. the objective of the study were to determine the prevalence and identify, gross and histopathological lesion of gills caused by *Dactylogyrus* and *Gryodactylus* spp. parasitic infection on naturally infected barbus fish.

**Result:** A cross sectional study was conducted from in Lake Tana, amhara region. Three hundred eight four gill specimens were collected from barbus fish and of these, 22 (5.7%) of fish were infected with monogenean parasites. Higher prevalence of *Dactylogyrus* spp. (4.86%) was recorded than *Gryodactylus* sp (1.04). Large size fishes ( $\geq 20$ cm) were more susceptible to *Dactylogyrus* spp and *Gryodactylus* spp. However, highest prevalence of *Dactylogyrus* sp. was found in large fish size (6.85%). *Gryodactylus* sp. was not recorded in all small size fish. Descriptive statistics such as percentages was used to describe the nature and the characteristics of the disease. At gross examination of gills, hyperaemia and swollen, excess mucus secretion, paleness, congestion of branchial blood vessel were observed. Gills specimens of infected fish were fixed with 10% formalin, for further identification and stained with the haematoxylin-eosin. Histopathological changes included hyperplasia, congestion and mucous cell proliferation of the gill epithelium and damaged primary and secondary lamellae, the uplifting of respiratory epithelial wall and damaged pillar cells.

**Conclusion:** Gross and histopathological changes induced by the *Dactylogyrus* spp and *Gryodactylus* spp of parasites adversely affected the proper functioning of the gills of the host fish which can lead to detrimental effect on the health status of the fish and may result in huge economic losses through increased mortality.

## Background

Gill parasites are common on cultured and wild fish. Many of these species have long been recognized to have the potential to affect the growth, fecundity and survival of hosts [1] Moreover, extensive tissue damage resulting from the feeding and attachment of these parasites have been reported in several species of fish [2]. Among the comment fish parasite, *Dactylogyrus* sp. was mostly observed in gills. *Dactylogyrus* sp have a series of hooks that enable them to attach while feeding. Most species are host- and site-specific, requiring only one host to complete an entire life cycle. Fish parasites result in huge economic losses as they increase mortality and also increase farm inputs via increased treatment expenses and cause reduction in growth rate due to the parasitic disease outbreak [3].

Also there was no research work conducted on gross and histopathological analysis on lesions caused by monogenea parasite of barbus fish at Lake Tana. Therefore, the objectives of this study were: General objective: The general aim of the study was to determine the pathological effect of gill trematode infection and the prevalence of gill *Dactylogyrus* spp and *Gryodactylus* spp of barbus fish at western part of the Lake Tana, Amahara region. Specific objectives: To determine the prevalence of gill

*Dactylogyrus* spp and *Gyrodactylus* spp parasites in barbus fish in western part of LakeTana, to identify the type of *Dactylogyrus* spp and *Gyrodactylus* spp parasites affecting gills on barbus fish in western part of Lake Tana and to determine the gross and histopathological lesions caused by the identified parasite of gills in barbus fish at western part of Lake Tana.

## Results

A total number of 384 *laebo barbus* fish were observed in Lake Tana. Out of the 384 fish examined, 22 (5.72) were infested with monogenean parasites where the prevalence of *Dactylogyrus* sp. was 4.86 % while that of *Gyrodactylus* sp. was 1.04% (Table 1).

Table 1

Prevalence of *Gyrodactylus* sp and *Dactylogyrus* sp in barbus fish in the study site.

Parasite species	No. examined	No. Infected	$\chi^2$	P-value
<i>Gyrodactylus</i>	384	4	9.17	0.0025
<i>Dactylogyrus</i>	384	18		
Total	384	22		

Highest prevalence of *Dactylogyrus* sp. was found in large sized fish group (6.85 %) and lowest in small sized fish group (2.29%). While highest prevalence of *Gyrodactylus* sp. was observed in large fish group (1.717%) and lowest in medium sized fish group (0.81%). *Gyrodactylus* sp. infestation was not recorded in small sized fish group.

Table 2

Length wise prevalence (%) of *Dactylogyrus* and *Gyrodactylus* sp. in the study area

Body length(cm)	No. examined	<i>Dactylogyrus</i> sp	<i>Gyrodactilus</i> sp.
Small (1-10)	87	2(2.38)	-
Medium (10.5-20)	122	4(3.27)	1(0.81)
Large (>20)	175	12(6.85)	3(1.71)
Total	384	18	4(1.04)

## Gross lesion examination

Gross examination of gill tissues caused by monogenean parasite revealed pale gills (22 infected fish), excess mucus production (8 infected fish), hyperemic (2 infected fish) and swollen gill tissue (18 infected

fish), hemorrhage (1 infected fish), congestion of (1 infected fish) was observed.

## Histopathological lesions

Microscopic pathological changes were observed on the gills of parasitized fish, and they were comprised of fusion, curling and necrotic change the gill filaments. The proliferation of bronchial tips, atrophy, complete loss of gill lamellar epithelium, hyperplasia and fusion of secondary gill lamellae were recorded. Uplifting of primary and secondary gill lamellae epithelium was also observed. Leucocytes infiltration, telangiectasia in secondary lamellae, congestion on the gill filaments and mucous cells proliferation were also observed during histopathological examination.

## Discussions

In present study, the histopathological effect caused by monogenea is mainly due to their morphological structures and specialized mode of attachment with host tissues. Infestation of the fish with *Gryodactylus* and *Dactylogyrus*, results severe damage to gills lamellae may be similar to lesions that were reported by [4].

In this study the Parasites attach gill epithelium and destruction of the cells or by congesting blood from the ruptured blood vessels. Gill epithelial cells are mechanically injured by the hooks of identified parasites. The blood expelled from this would be fed by the parasite and activity of some parasites can lead to bacterial or fungal secondary infection and causes mass mortality in cultured and wild situation and consequently fish respiration will be impaired, which results in reduced feeding, weight loss and general deterioration of health. Decreasing the body weight and condition factor, sever changes in osmoregulation or respiratory dysfunction and finally death may be observed in infection with *Gryodactylus* and *Dactylogyrus* parasite these study agree with [5].

The histopathological lesion which is observed in this study include lamellar fusion, hyperplasia, and aneurysm. Some alternations in blood vessel may occur when fish suffer from severe types of stress. In this case, damaged pillar cells can result in an increased blood flow inside the lamellae and cause blood congestion or even an aneurysm [6]. In the present study of monogenea infection, gill inflammation, swollen, hyperemic, hemorrhage, and excessive mucus production were recorded in infected barbus fish gill tissue. Leads to, blockage of blood vessels leading to respiratory and osmoregulatory failure according to [7].

In the present study, necrosis lamellae were observed in barbus fish infected with *Gryodactylus* and *Dactylogyrus* spp of monogenean parasites. The mechanism by which the destruction of the branchial vessels may happen by the parasite, where the blood pressure is low and no extensive hemorrhages are caused and the very short clotting time of blood brings about rapid occlusions of the vessel then thrombus is formed resulting in ischemia, which in turn leads to necrosis, these result similar with [8] and [9]. In the present study, reduced chloride cells were seen. The infection of identified parasite in fish

gills has an impact on the host's ability to regulate its ion balance. These findings agree with reports of [10] and [11]. These parasites reduced the number of chloride cells which are the main site of ion absorption and secretion [12].

In the present study hemorrhage was observed. *Gryodactylus* and *Dactylogyrus* parasitic damage the gills by feeding on the delicate tissue of the gill lamellae or on the blood circulating within the lamellae, leading to a loss of respiratory surface area, extensive gill damage and severe hemorrhage, with inflammation. Blood vessels in the gill filaments are blocked and this leads to atrophy of gill tips. These findings agree with [13] and [14].

## Conclusions

The major features of *Gryodactylus* and *Dactylogyrus* parasite spp. infections on the gill filaments of the fish in this study mostly include destruction of the gill filaments and lamellae, exerted by feeding of the parasite and the result in hyperplasia and thickening of the epithelial cells reducing the surface area for effective respiration. Gill damage could result in loss of gill surface area for respiration, which would lead to suffocation particularly at high water temperatures. The histoarchitectural change in gills hinders the oxygen intakes and in the long term causes huge mortality in aquaculture practices. According to the result of the gross and histopathological changes induced by the infections of parasites would adversely affect the proper functioning of the gills of infected fish. Further study should be warranted on the prevalence and effect of other parasites.

## Materials And Methods

### Study population

The study animals were fish of barbus (*Labeo barbus*) that was collected from Lake Tana, Amahara region, Ethiopia. Sampled fishes were selected by simple random sampling technique. Large, medium and small sized fishes were included in my study. Large, medium and small sized fishes were selected by simple random sampling techniques associations that are found in western part of Lake Tana. The study population, small (1 to 10 cm), medium (10.5 to 20cm) and large size fishes ( $\geq 20$  cm) were categorized according to [15].

### Sample collection techniques, gross and histopathological examination

A total of 384 randomly selected fish, *Labeo barbus*, were sampled and examined. The length, date and site of collection of host specimens were recorded. Gross lesion and health conditions of each specimen were recorded and monogenean parasite (*Gryodactylus* and *Dactylogyrus* spp.) of gills were collected with forceps. Postmortem examination of gills of the investigated fishes was examined according to

[16]. Damaged fish tissues (gills) were taken from the parasite attachment area of infested fishes and were cut out in fresh condition fixed in 10% buffered neutral formalin. In addition, monogeneans parasites of gills were collected and preserved in 10% buffered neutral formalin. Then, the necessary information was labeled on the sampling bottles. Finally, the samples were transported to Faculty of Veterinary Medicine Pathology and parasitology laboratory, University of Gondar, for identification of parasites and histopathological examination of tissue specimens. After using the fixative, the tissues were washed in running water for 24 hr to remove the fixative entirely. The tissues were then processed using a 18 hr automatic tissue processor. The tissue processor contains 12 beakers (8 glass beakers contain alcohol, 2 glass beakers contain xylene and 2 glass beakers containing paraffin wax).

The tissues after being processed are embedded using an automatic embedding centre. Embedding is a process of submerging a tissue in a metal plastic disposable embedding mould containing molten paraffin wax, which became solidified when it was cold. This formed a support medium for the tissue during sectioning. The blocked tissues were sectioned at 4-5 microns on a rotary microtome (LEITZ 2535, Germany), and floated into pre-coated slides and were placed in a clean grease-free slide which was then placed on a hot plate for 30 min in order for the section to adhere to the slides. The staining method used was the H&E staining method. Standard histological procedure was followed as described by [17]. This method was used in order to demonstrate the general structure of the tissues. These were then dewaxed in xylene. The processed sections were later taken to water by using descending grades of alcohol, that is, from absolute alcohol, 95% alcohol, 70% alcohol, 50%, for 5 min each and rinse with tap water for 10 min. It was stained in haematoxylin for 6 min, then put in running water for 20 min.

This was counter-stained in 1% Eosin for 15 min and, then dehydrated using ascending grades of alcohol (95% alcohol and absolute alcohol). These were cleared in xylene, mounted using D.P.X (a mountant) and viewed under light microscope [18]. Lesions were described and scored as none for no lesion, mild, or severe depends on the type of lesion [19]. Preserved in 10% formalin and labeled with all necessary information for further identification. The collected parasites placed on glass microscope slides with a drop of 10% formalin and slightly compressed between a slide and a cover slip prior to being examined under stereomicroscope (4x, 10x, 40x) and observe morphological features of the parasite. The monogenean parasites were identified microscopically using the identification guideline of [20].

## Data analysis

The data were entered and managed in Microsoft Excel. All the data analysis was done by SPSS software version 12. Descriptive statistics were applied for the analysis of the data obtained. Descriptive statistics such as percentages were used to describe the nature and the characteristics of the data. The prevalence of monogenean parasite (*Gyrodactylus* and *Dactylogyrus* spp.) was analyzed using Chi-square test and  $<0.05$  was considered as significant.

## Declarations

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# Author's contribution

Samrawit Melkamu-Mekonnon contributed for data collection, sample analysis, data summarization manuscript preparation Abiyot Workeale Alemu, for manuscript preparation, data analysis; statistical analysis, table and figure preparation as a contribution.

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# Availability of data and materials

All data supporting these research findings are included within the manuscript. The databases (without personally identifiable information) are available from the corresponding author upon request.

# Consent for publication

Not applicable.

# Competing interest

The authors declare that they have no competing interests.

# Ethics approval and consent to participate

The study reviewed by Animal Ethics and Experimentation review process of the Research and Community Services Council Office of the College of Veterinary Medicine, Gondar University, Ethiopia. Samples collected after getting verbal consent from willing owners. Considering the less invasive nature of the sampling procedure, verbal consent was enough and approved by the ethics review committee.

# Author's detailed

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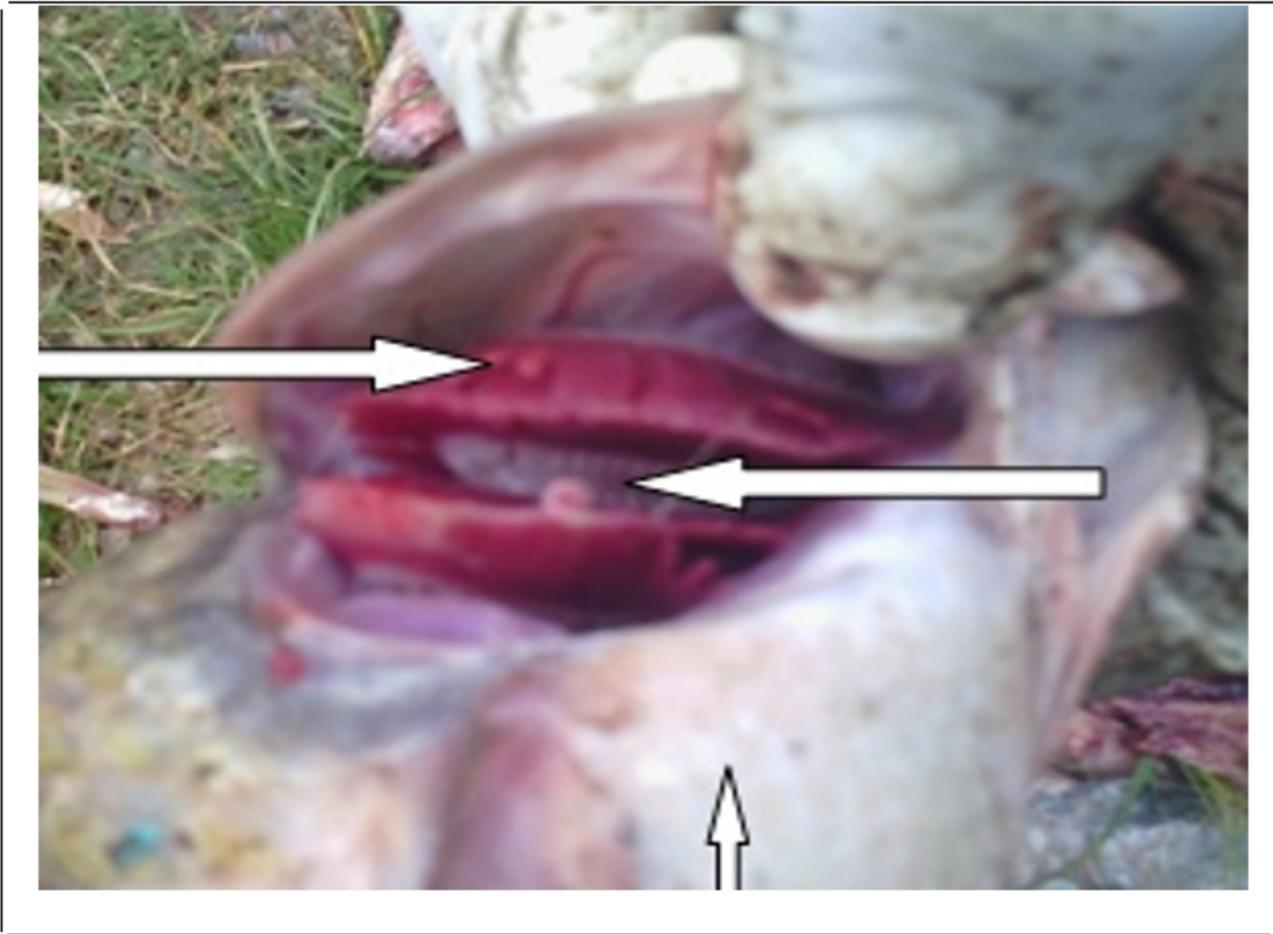
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## Figures



**Figure 1**

Paleness of gill tissue



**Figure 2**

Monogenean parasite attached to the gills and congested



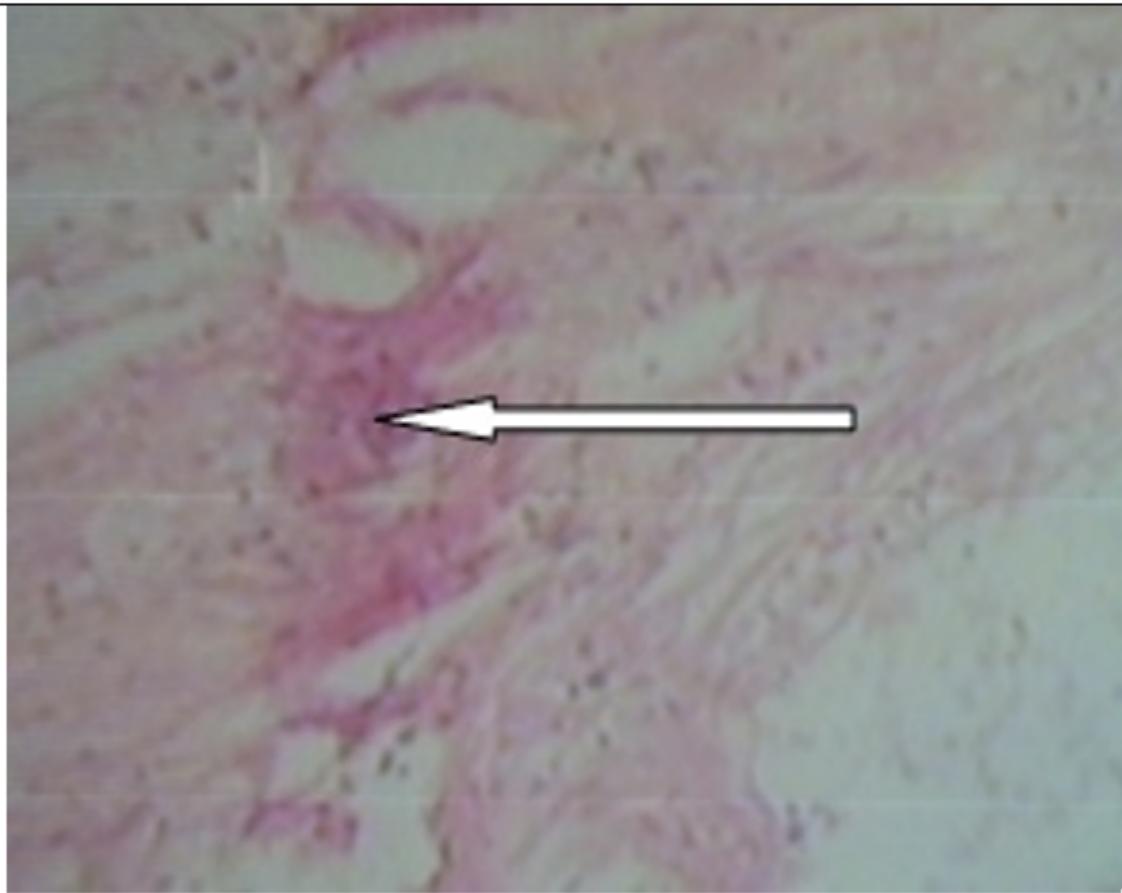
**Figure 3**

Monogenea parasite attached to between the gill filaments



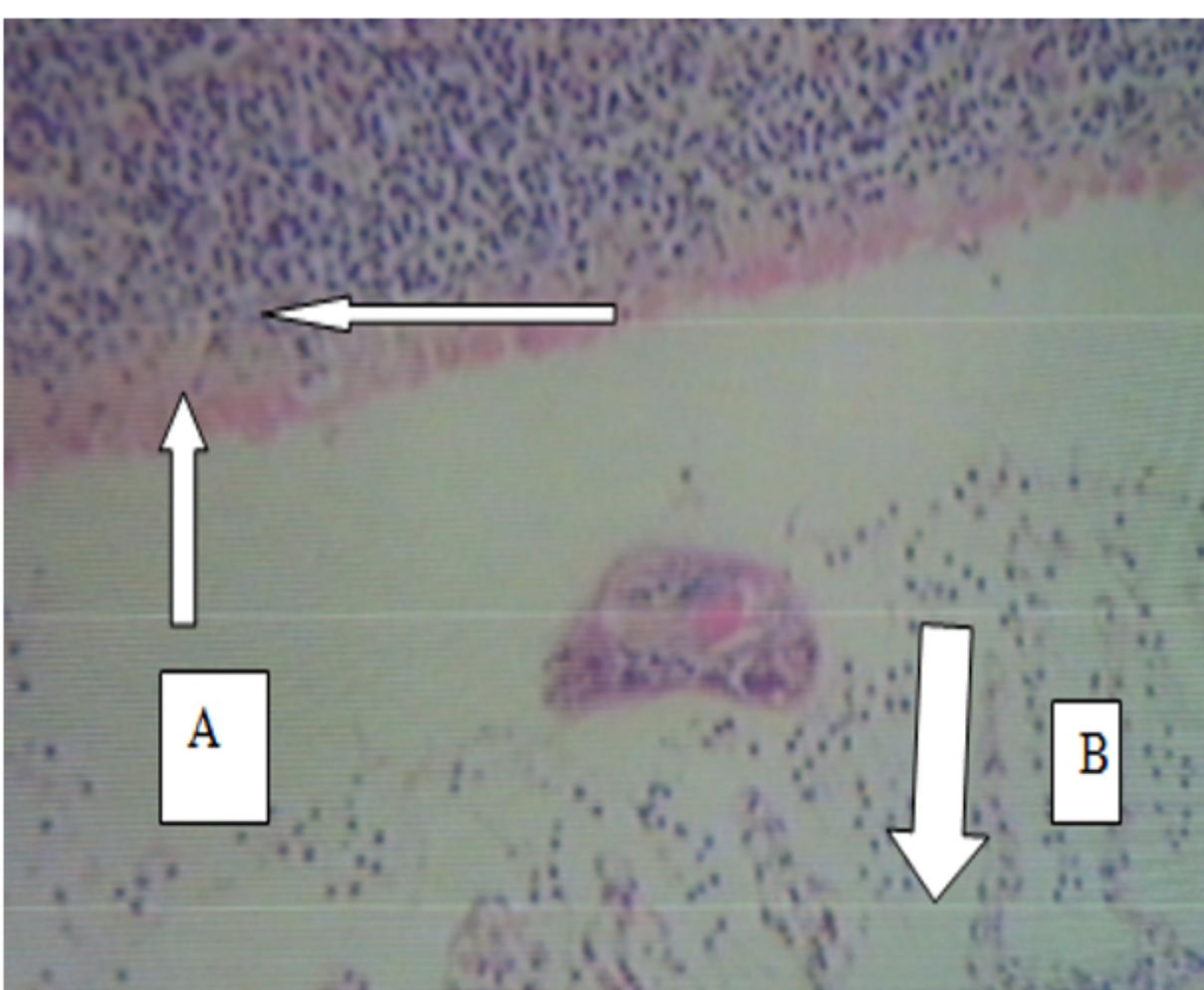
**Figure 4**

Normal structure of gills.



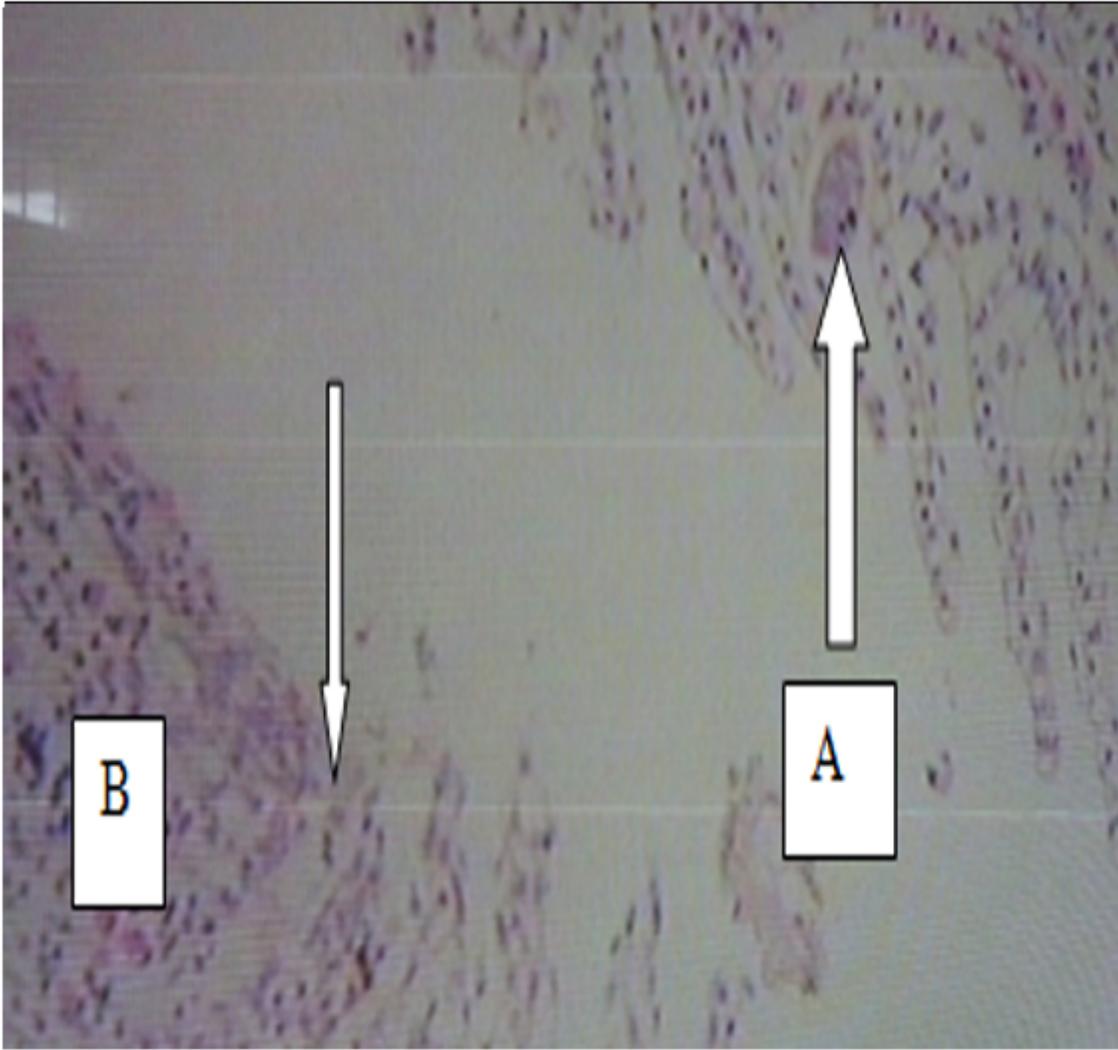
**Figure 5**

Hemorrhage of blood vessels



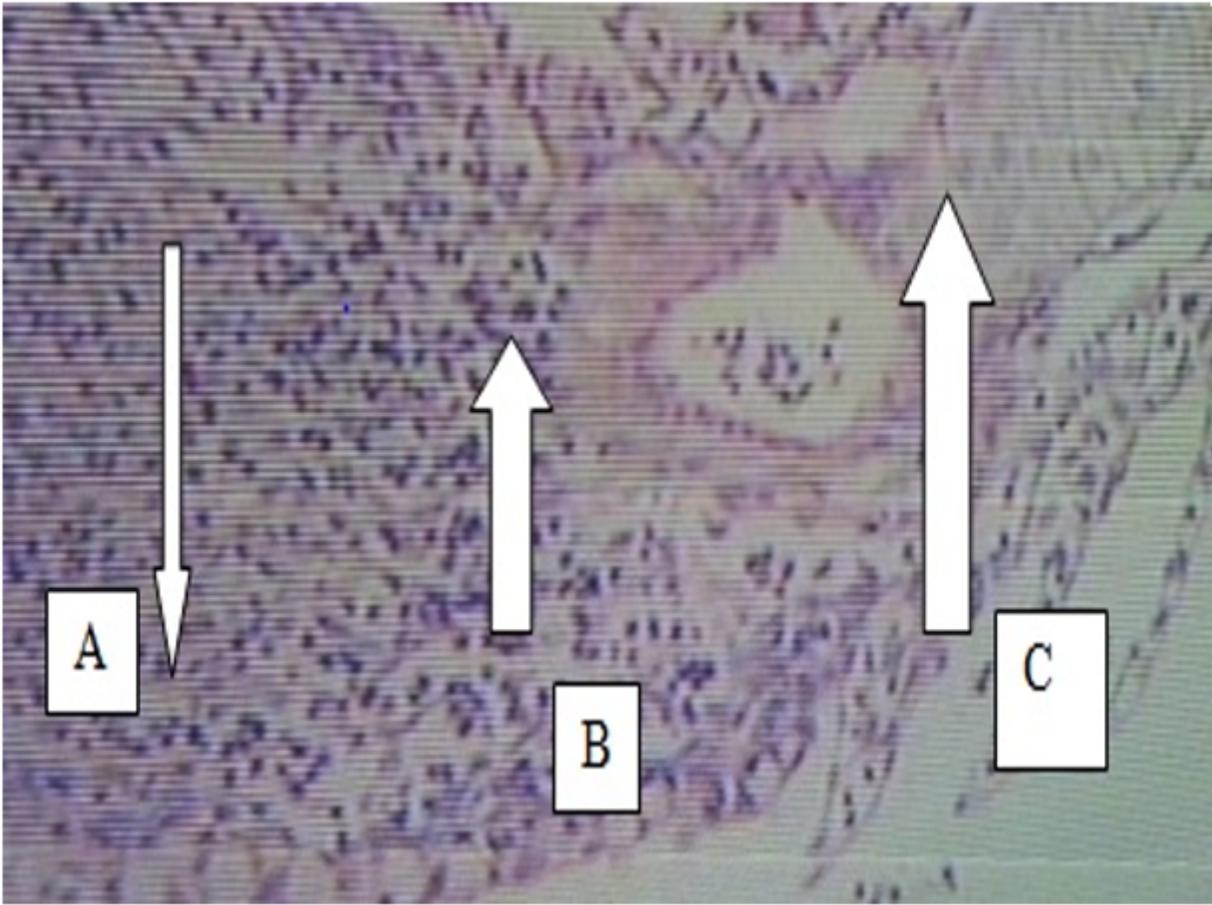
**Figure 6**

Monogenea parasite attached to the secondary gill lamellae(A) and chloride cell damage(B) (arrow



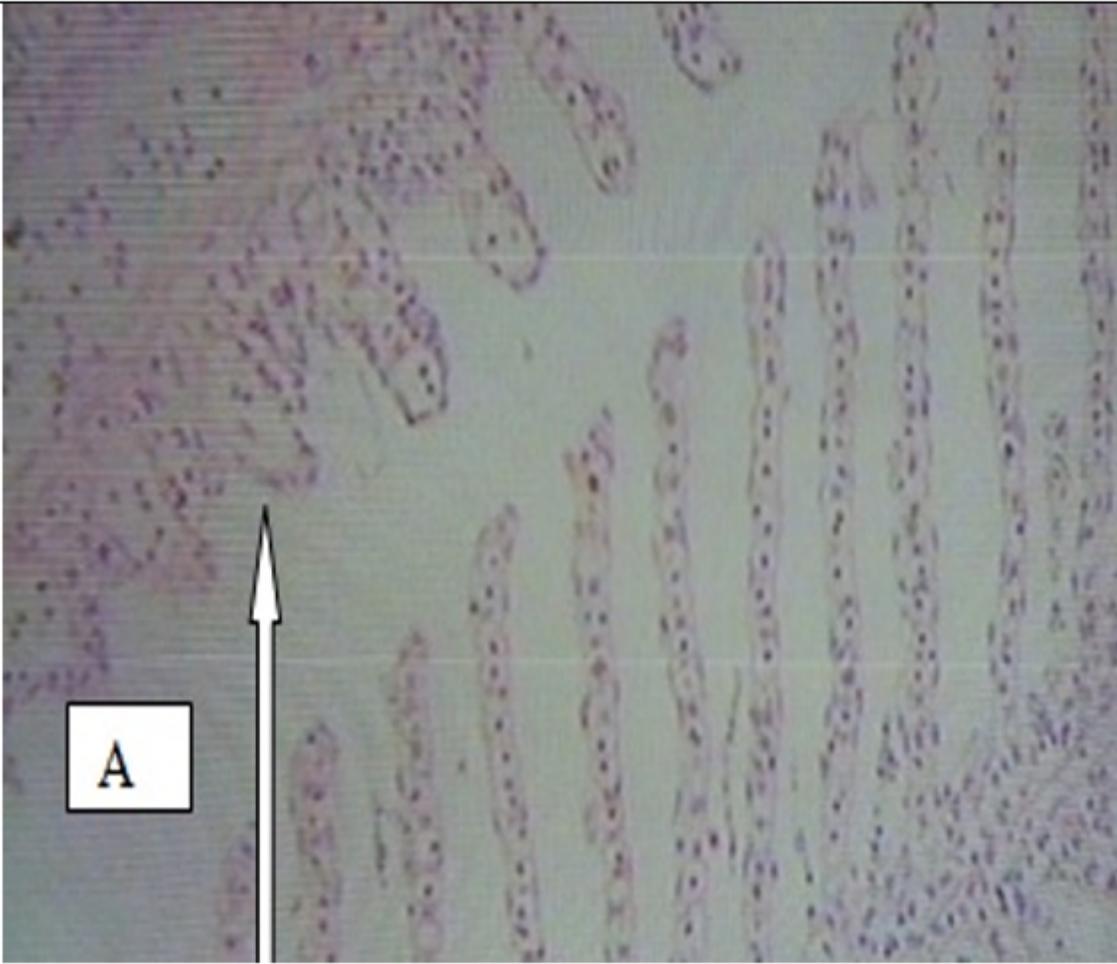
**Figure 7**

Monogenea parasite attached to inter lamellar space (a) and fusion of adjacent lamellae(B) (arrow)



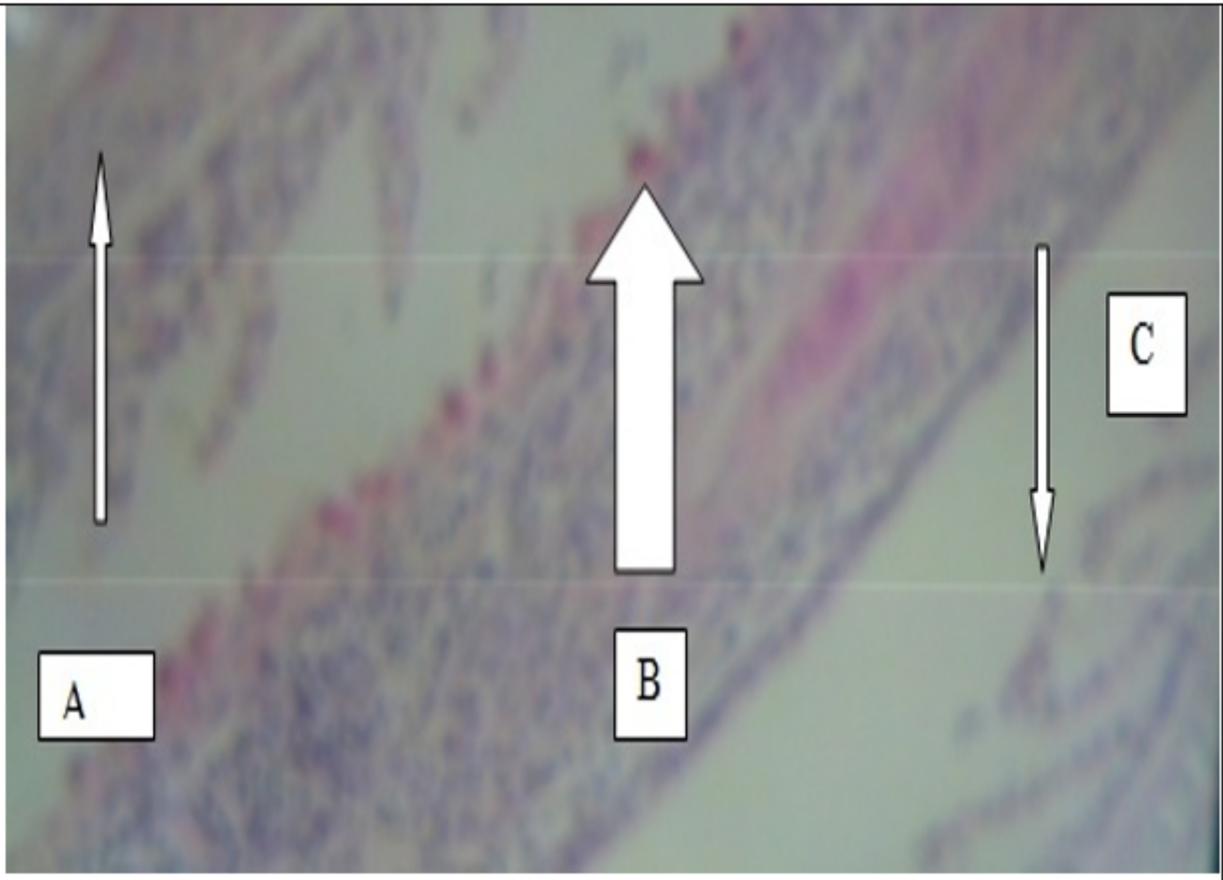
**Figure 8**

Increase number of goblet cell (A) leucocytes infiltration (B) congestion of blood vessel (C) (arrows)



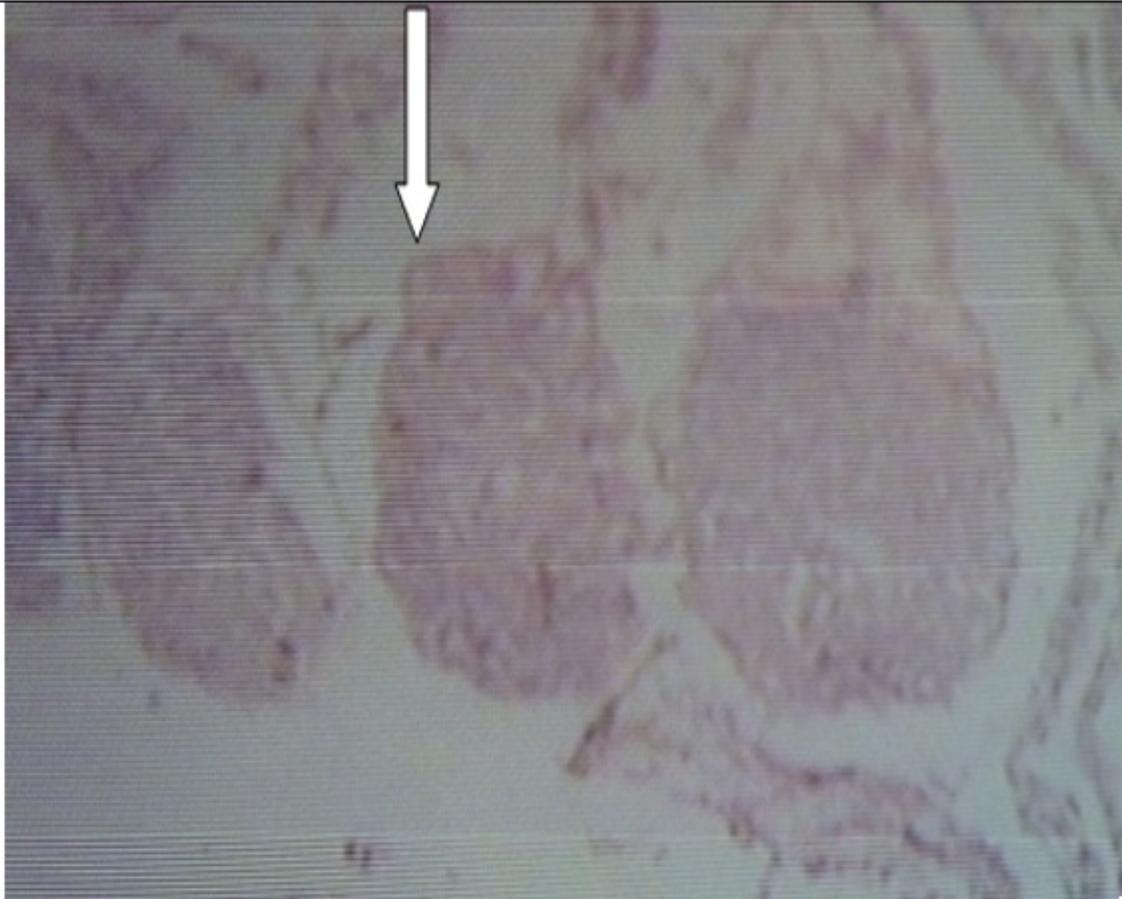
**Figure 9**

Atrophy of secondary lamellae (A) (arrow)



**Figure 10**

necrosis (A)complete loss of secondary lamellae(B)and decrease chloride cells number(C) (arrows)



**Figure 11**

Telangiectasia condition at the tip of secondary lamellae (arrow)