

Targeting CD33 for Acute Myeloid Leukemia Therapy

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Abstract

Objective

The aim of this study was to analyze the level of CD33 expression in patients with newly diagnosed AML and determine its correlation with clinical characteristics. Methods: Samples were collected for analysis from AML patients at diagnosis. We evaluated the level of CD33 expression by flow cytometry analysis of bone marrow. Chi-square or t- tests were used to assess the association of categorical variables between the high and low CD33 expression groups. Spearman's rho was applied when appropriate. Survival curves were generated by the Kaplan–Meier method.

Results

In this study we evaluated the level of CD33 expression in de novo patients diagnosed from November 2013 until January 2019. The mean value of 73.4% was used as the cutoff for the two groups. Statistical analysis revealed that 53 of the 86 (61.2%) AML patients were above the mean. Although there was no statistical significance between CD33 expression level and gene mutation, FLT3 mutation ($P=0.002$) and NPM1 mutation ($P=0.001$) were more likely to be seen in the high CD33 group. The overall survival (OS) was worse in the high CD33 group (39.0 m vs. 16.7 m, $\chi^2 = 7.385$, $P = 0.007$). Multivariate analysis showed that the high expression of CD33 was an unfavorable prognostic factor. Of the 86 patients, CD33-high was closely related to the patients with normal karyotype ($\chi^2 = 4.891$, $P = 0.027$), high white blood cell count (WBC, $t = 2.804$, $P = 0.007$), and a high ratio of primitive cells ($t = 2.851$, $P = 0.005$).

Conclusions

These findings provide a strong rationale for targeting CD33 in combination with chemotherapy, which can be considered a promising therapeutic strategy for AML.

Introduction

In adult patients, acute myeloid leukemia (AML) is a common hematological malignancy. Most of the patients have a poor prognosis. Conventional chemotherapy for AML, including induction and consolidation treatment, is only partially effective. Patients often require bone marrow transplantation and multiple rounds of consolidation therapy. Even with regular chemotherapy, the overall 5-year survival remains below 30% for all patients and is lower for older patients. Most importantly, many patients are refractory to conventional chemotherapy [1–2]. Thus, we need to develop new methods to improve survival in this fatal form of leukemia. Recently, immunotherapy has been recognized as a new treatment strategy for hematologic malignancies [3–4]. CAR-T immunotherapy has shown excellent results in ALL (acute lymphocytic leukemia) and lymphoma, and it is possible that patients with acute myeloid leukemia (AML) can also benefit from this treatment strategy [5].

Unlike ALL, AML surface antigens are highly heterogeneous. We need to identify antigens that are specifically expressed in AML as markers for immunotherapy. CD123, CD44, CD174 and CD33 are expressed in hematopoietic stem and progenitor cells, increasing off-target toxicity and killing hematopoietic stem and progenitor cells in immunotherapy [3,6–7].

Most efforts of developing monoclonal antibodies or antibody-drug conjugates (ADCs) for AML have focused on targeting CD33 (cluster of differentiation antigen 33). Leukemic blasts and myeloid leukemia-initiating cells express CD33. CD33 does not appear on the surface of primitive stem cells or multipotent progenitor cells. These factors make it a favorable target for immunotherapy of AML[1,8–10]. There have been a number of reports confirming that CD33 is a feasible target for immunotherapy of AML. Due to the approval of anti-CD33 Mylotarg® (GO, gemtuzumab ozogamicin) in 2000, GO was the first anticancer ADC on the market[8,10–12]. In our study, we investigated the correlation between the level of CD33 expression in patients with newly diagnosed AML and the prognosis of patients. All data were obtained from 86 newly diagnosed AML patients. Of course, all processes met the ethical standards, and patient consent was obtained. This study provides more persuasive evidence for the immunotherapy of AML.

Patients And Method

Patients

Between November 2013 and January 2019 in our institution (Department of Hematology, First Affiliated Hospital of Henan University of Science and Technology), 86 patients with an initial diagnosis and complete information were enrolled in the study group. All patients were diagnosed, evaluated and treated according to National Comprehensive Cancer Network (NCCN) guidelines[13]. All patients' records were evaluated retrospectively for the level of CD33 expression in de novo AML patients. Patients were grouped according to expression levels above and under the mean, that is, into high and low level of CD33 expression groups. The association between patient clinical information and CD33 expression was analyzed. Detailed baseline characteristics are shown in Table 1.

Table 1
Patients' characteristics at initial diagnosis.

Variables	Low-CD33(n = 33)	High-CD33(n = 53)	P
Sex(N,M/F)	20/13	34/19	0.742
Age[Y,M(range)]	44(11–83)	47(16–86)	0.457
WBC[10 ⁹ /L,M(range)]	16.30(0.8–145.2)	54.66(0.9–471.6)	0.007
PLT[10 ⁹ /L,M (range)]	58.71(7.0–309.0)	58.70(3.0–275.0)	1.000
primitive cell(%)	46.64(16.7–81.5)	60.60(7.5–98.8)	0.005
FAB subtype(N,%)	0(0)	1 (1.9)	0.063
M0	0(0)	4 (7.5)	0.027
M1	27(81.8)	23(43.4)	
M2	5(15.2)	16(30.2)	
M4	1(3.0)	9 (17.0)	
M5			
Karyotype			
Normal	10(30.3)	29(54.7)	0.114
Other	23(69.7)	24(45.3)	0.775
Gene	25(75.8)	47(88.7)	0.783
Mutated	8(24.2)	6(11.3)	
Non-Mutated	17(51.5)	29(54.7)	
Induction chemotherapy	16(48.8)	24(45.3)	
CR(N,%)	15(45.5)	21(39.6)	
NR(N,%)	18(54.5)	31(58.5)	
Relapse or not(N,%)		1 (1.9)	
Yes			
No			
NA			
WBC,the count of white blood cell; PLT,platelets ; CR,complete remission ; NR,not remission ; YES, referred to relapse; NO, referred to not relapse ; NA,not available.			

Treatments

Induction chemotherapy used standard DA (daunorubicin + cytarabine), IA (nordomycin + cytarabine), MA (mitoxantrone + cytarabine) or CAG with or without the D (arabin cytidine + aclamycin + granulocyte colony stimulating factor with or without decitabine) regimen. Consolidation chemotherapy was given after complete remission (CR), and the regimen referred to induction chemotherapy and usually included cytarabine. For patients with moderate or poor prognosis, allogeneic hematopoietic stem cell transplantation should be performed with appropriate, medium and large dose regimens.

Methods

A multicolor immunolabeling method was used to determine the expression ratio of CD33. The proportion of CD33 expression was determined by FACS analysis (FACS Navios, Beckman Coulter). as described[14–15]. Bone marrow chromosome karyotype analysis was performed by the R-banding method. Real-time quantitative PCR (RT–PCR) assay was used to detect fusion genes. Gene mutations were targeted for sequencing by the Illumina sequencing platform. The COSMIC, dbSNP, PolyPhen, SIFT and other databases and software for bioinformatics analysis were applied to determine pathogenic gene mutations [16].

Efficacy, prognosis evaluation and follow-up

In the retrospective analysis, the risk stratification and the definition of CR, NR, OS, etc. were obtained from the NCCN guidelines of AML[13]. Follow-up was conducted by consulting medical records or via telephone. The follow-up deadline was June 31, 2019.

The probability of OS was calculated according to Kaplan–Meier analysis and compared by the log-rank test using SPSS 20.0. A $P < 0.05$ was considered significant.

Statistical analysis

Data were analyzed using SPSS 20.0 (Statistical Program for Social Sciences Version 20.0). Data acquisition was halted on 1st June 2017.

The chi-square test (2-tailed) was used when comparing the categorical variables; a value of $p \leq 0.05$ was considered statistically significant. Fisher's exact test and t-test were used under the right conditions. The Kaplan–Meier method was used to construct survival curves. Overall survival was calculated from the time of diagnosis to all-cause death. In univariate and multivariate analyses, log-rank tests were applied to analyze differences in OS between groups. Estimation was limited to the largest survival if it was censored.

Results

Patient characteristics. Table 1 summarizes all the characteristics at diagnosis of the 86 patients in the retrospective analysis in the study. Patients were divided into high level of CD33 expression (High-CD33) and low level of CD33 expression (Low-CD33) groups. We found significant differences in the WBC (white

blood cell) count, ratio of primitive cells and karyotype between the two groups. These results suggest that CD33 expression is higher in patients with high WBCs, a high proportion of primitive cells and a normal karyotype.

Gene analysis. The initial evaluation of AML patients is important and it has two prerequisite aims. On the one hand, to characterize the disease process and provide prognostic information, we can base the analysis on factors such as prior toxic exposure, antecedent myelodysplasia, and karyotypic or molecular abnormalities. These factors may impact responsiveness to chemotherapy and the risk of relapse. On the other hand, we assessed the patients' ability to tolerate chemotherapy by evaluating comorbidities. Both factors are taken into consideration when deciding treatment. Molecular abnormalities play an important role in prognostic stratification. We found molecular abnormalities, including mutant genes and fusion genes, by performing PCR (polymerase chain reaction) and NGS (next-generation sequencing) experiments. Samples for mutation detection were collected from de novo AML patients.

To determine the gene mutation or fusion gene in relation to the level of CD33 expression, we analyzed 86 patients in this retrospective study. In 62 patients (72.1%), molecular abnormalities were detected, which may greatly contribute to the development of AML. Some genes with a high frequency among these gene abnormalities, such as FLT3, NPM1, DNMT3A, IDH1, and CEBPA, are shown in Table 2. The results suggested that more patients with FLT3 mutations ($\chi^2 = 9.778$, $p = 0.002$) and NPM1 mutations ($\chi^2 = 11.305$, $p = 0.001$) presented high levels of CD33 expression. Of the 86 patients, 21 (24.4%) were positive for FLT3. Among these patients with FLT3 mutations, there were 19 patients with high CD33 expression and 2 patients with low CD33 expression. Similarly, there were 19 (22.1%) NPM1 mutation patients, of which 18 patients had high levels of CD33 expression.

Table 2
Some genes with a high frequency among gene abnormalities.

Gene	Low-CD33(n = 33)	High-CD33(n = 53)	P
FLT3	2	19	0.002
IDH2	2	5	0.880
RUNX1	4	2	0.297
ASXL1	6	3	0.111
KIT	3	4	1.000
DNMT3A	4	16	0.054
IDH1	3	6	1.000
TP53	2	1	0.673
AML1/ETO	3	4	1.000
U2AF1	1	2	1.000
NRAS	2	0	0.144
BCOR	1	2	1.000
CEBPA	3	11	0.154
TET2	2	7	0.490
WT1	3	3	0.863
HOX11	1	0	0.384
NPM1	1	18	0.001

Survival analysis. The median follow-up time for all patients in the present study, from the day of diagnosis to the last follow-up date, was 16.4 months (range, 0.3–68 months). The OS rates of patients in the low CD33 expression and high CD33 expression groups were 67.5% and 42.9%, respectively (log rank = 7.385, P = 0.007). The results are shown in the Figure.

Discussion

Acute myeloid leukemia (AML) is a heterogeneous hematological malignancy characterized by the proliferation of leukemia-initiating cells, and the majority of cases have a poor prognosis. It is the most common form of acute leukemia among adults, with the largest number of deaths due to leukemia each year. The median age at diagnosis is 67 years, and 54% of patients are diagnosed when they are 65 years or older (approximately one-third are diagnosed as ≥ 75 years)[13].

Considerable progress has been made in anticancer treatments and breakthroughs in immunotherapies, but conventional AML chemotherapy, including induction and consolidation therapy, has not changed significantly. This highlights the need to develop new ways to improve prognosis, especially refractory and relapsed AML (R-R AML)[17–18].

Allogeneic hematopoietic stem cell transplantation (allo-HSCT) may remain the sole curative option. Therefore, immunotherapies, such as ADC, CAR-T and GO, bring promise and challenge the traditional regimens given to many R-R AML patients[9,11,19].

Although immunotherapy has been widely accepted as an effective strategy in the field of hematological malignancies, the lack of high specificity of target antigens and the heterogeneity of AML has led to slow progress in treating AML with similar strategies. Therefore, the challenge for us is to determine an antigen expressed specifically on the surface of leukemia cells in AML[4].

In this study, the results showed that the expression level of CD33, an independent risk factor affecting prognosis, was related to the OS of patients. At diagnosis, AML patients with a higher count of white blood cells, a higher percentage of primitive cells, and normal karyotypes had higher levels of CD33 expression. Patients with FLT3 and NPM1 mutations had higher expression of CD33. These positive results suggested that patients would most likely benefit from CD33 targeted therapy when de novo AML patients had the abovementioned clinical characteristics. In our opinion, taking into account all the available data can help improve the prognosis and survival of AML patients. Limitations of this study include the sample size and grouping the patients according to more detailed characteristics would be beneficial for analysis.

Of course, more research is required, preferably prospective clinical data, to optimize the regimen of targeting CD33 for AML. More clinical features or other factors need to be identified to help us choose CD33 as a targeted therapy for AML or in combination with chemotherapy.

Patients with FLT3 and NPM1 mutations had higher expression of CD33, which was consistent with the published literature[20]. The high expression of CD33 in NPM1-mutated patients may be related to the concomitant occurrence of NPM1 and FLT3 mutations. However, this conclusion needs to be supported by more clinical data. We can conclude that CD33 is a target for AML immunotherapy [21].

Targeting CD33 appears to be a suitable alternative in patients who lack hematologic stem cell donors. Ongoing efforts are needed to optimize the application to enhance therapeutic effects and decrease injury, including cytotoxicity and economic losses. To investigate the possibility of immune-based therapies beyond stem cell transplantation to treat hematologic malignancies and recommend targeting CD33 therapy for more extended list treatments, future research should focus on studies with higher quality parameters, such as larger sample sizes, randomized studies and prospective studies.

Responsive biomarkers will enable us to select patients who are more likely to benefit from immune checkpoints and monoclonal-based therapies. Exploiting the true potential of immune agents in AML

requires excellently designed clinical trials. Trials are ongoing and will guide further development of immune agents[17,21–24].

Declarations

Ethics approval and consent to participate

All procedures performed in studies involving human participants were in accordance with the ethical standards of the Research Ethics Committee of The First Affiliated Hospital of Henan University Science and Technology and with the 1964 Helsinki declaration and its later amendments. ALL written informed consent to participate in the study was obtained from AML patients for data to be analyse.

Consent for publication

All subjects have written informed consent.

Availability of data and materials

The datasets used or analysed during the current study are available from the corresponding author on reasonable request.

Statement of conflict of interest

The authors declare that they have no competing interests.

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Authors' contributions

The authors contributed equally to this work. Jingjing Liu analyzed and interpreted the patient data. Jiayin Tong and Haiping Yang collected patients' data. Jingjing Liu and Haiping Yang drafted and revised the manuscript. All authors have read and approved the final manuscript and confirmed the authenticity of all the raw data. The authors are accountable for all aspects of the work and ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

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References

1. Schneider, D., Xiong, Y., Hu, P., Wu, D., Chen, W., Ying, T., Zhu, Z., Dimitrov, D., S., Dropulic, B., Orentas, R. J. A Unique Human Immunoglobulin Heavy Chain Variable Domain-Only CD33 CAR for the Treatment of

- Acute Myeloid Leukemia. *Front Oncol* 2018;8: 539.
2. Veronica A. Guerra, Courtney DiNardo, Marina Konopleva. Venetoclax-based therapies for acute myeloid leukemia. *Best Pract Res Clin Haematol* 2019;32(2):145–153.
 3. Ma, H., Padmanabhan, I. S., Parmar, S., Gong, Y. Targeting CLL-1 for acute myeloid leukemia therapy. *J Hematol Oncol* 2019;12(1): 41.
 4. Im, A. and S. Z. Pavletic. Immunotherapy in hematologic malignancies: past, present, and future. *J Hematol Oncol* 2017;10(1): 94.
 5. John, S., Chen, H., Deng, M., Gui, X., Wu, G., Chen, W., Li, Z., Zhang, N., An, Z., Zhang, C. C.. A Novel Anti-LILRB4 CAR-T Cell for the Treatment of Monocytic AML. *Mol Ther* 2018;26(10): 2487–2495.
 6. Yoshida, G. J., H. Saya. Therapeutic strategies targeting cancer stem cells. *Cancer Sci* 2016; 107(1): 5–11.
 7. Siveen, K. S., Uddin, S., Mohammad, R. M. Targeting acute myeloid leukemia stem cell signaling by natural products. *Mol Cancer* 2017;16(1): 13.
 8. Leipold, D. D., Figueroa, I., Masih, S., Latifi, B., Yip, V., Shen, B. Q., Dere, R. C., Carrasco-Triguero, M., Lee, M. V., Saad, O. M., Liu, L., He, J., Su, D., Xu, K., Vuilleminot, B. R., Laing, S. T., Schutten, M., Kozak, K. R., Zheng, B., Polson, A. G. Kamath, A. V. Preclinical pharmacokinetics and pharmacodynamics of DCLL9718A: An antibody-drug conjugate for the treatment of acute myeloid leukemia. *MAbs* 2018;10(8): 1312–1321.
 9. Mussai, F., Wheat, R., Sarrou, E., Booth, S., Stavrou, V., Fultang, L., Perry, T., Kearns, P., Cheng, P., Keeshan, K., Craddock, C., De Santo, C. Targeting the arginine metabolic brake enhances immunotherapy for leukaemia. *Int J Cancer* 2019;145(8): 2201–2208.
 10. Schurch, C. M. Therapeutic Antibodies for Myeloid Neoplasms-Current Developments and Future Directions. *Front Oncol* 2018; 8: 152.
 11. Norsworthy, K. J., Ko, C. W., Lee, J. E., Liu, J., John, C. S., Przepiorka, D., Farrell, A. T., Pazdur, R. FDA Approval Summary: Mylotarg for Treatment of Patients with Relapsed or Refractory CD33-Positive Acute Myeloid Leukemia. *Oncologist* 2018;23(9): 1103–1108.
 12. Kovtun, Y., Noordhuis, P., Whiteman, K. R., Watkins, K., Jones, G. E., Harvey, L. Lai, K. C., Portwood, S., Adams, S., Sloss, C. M., Schuurhuis, G. J., Ossenkoppele, G. Wang, E. S., Pinkas, J. IMG779. a Novel CD33-Targeting Antibody-Drug Conjugate with DNA-Alkylating Activity, Exhibits Potent Antitumor Activity in Models of AML. *Mol Cancer Ther* 2018;17(6): 1271–1279.
 13. NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). Acute Myeloid Leukemia .Version 3.2018-November 30, 2018.
 14. Braciak, T. A., Roskopf, C. C., Wildenhain, S., Fenn, N. C., Schiller, C. B., Schubert, I. A., Jacob, U., Honegger, A., Krupka, C., Subklewe, M., Spiekermann, K., Hopfner, K. P., Fey, G. H., Aigner, M., Krause, S., Mackensen, A. Dual-targeting triplebody 33-16-123 (SPM-2) mediates effective redirected lysis of primary blasts from patients with a broad range of AML subtypes in combination with natural killer cells. *Oncoimmunology* 2018;7(9): e1472195.

15. Olejniczak, S. H., Stewart, C. C., Donohue, K., Czuczman, M. S. A quantitative exploration of surface antigen expression in common B-cell malignancies using flow cytometry. *Immunol Invest* 2006;35(1): 93–114.
16. Xing, S., Wang, B., Gao, Y., Li, M., Wang, T., Sun, Y., Shen, Y., Chao, H. Cytogenetics and associated mutation profile in patients with acute monocytic leukemia. *Int J Lab Hematol* 2019;41(4): 485–492.
17. Saygin, C., H. E. Carraway. Emerging therapies for acute myeloid leukemia. *J Hematol Oncol* 2017;10(1): 93.
18. Walter, R. B., Appelbaum, F. R., Estey, E. H., Bernstein, I. D. Acute myeloid leukemia stem cells and CD33-targeted immunotherapy. *Blood* 2012;119(26): 6198–6208.
19. Zhang, C. C., Yan, Z., Pascual, B., Jackson-Fisher, A., Huang, D. S., Zong, Q., Elliott, M., Fan, C., Huser, N., Lee, J., Sung, M. Gemtuzumab Ozogamicin (GO) Inclusion to Induction Chemotherapy Eliminates Leukemic Initiating Cells and Significantly Improves Survival in Mouse Models of Acute Myeloid Leukemia. *Neoplasia* 2018;20(1): 1–11.
20. Liu, Yan-Rong, Zhu, Hong-Hu, Ruan, Guo-Rui, Qin, Ya-Zhen, Shi, Hong-Xia, Lai, Yue-Yun, Chang, Yan, Wang, Ya-Zhe, Lu, Dan, Hao, Le, Li, Jin-Lan, Li, Ling-Di, Jiang, Bin, Huang, Xiao-Jun. NPM1-mutated acute myeloid leukemia of monocytic or myeloid origin exhibit distinct immunophenotypes. *Leukemia Research* 2013;37(7): 737–741.
21. Assi, R., Kantarjian, H., Ravandi, F., Daver, N. Immune therapies in acute myeloid leukemia: a focus on monoclonal antibodies and immune checkpoint inhibitors. *Curr Opin Hematol* 2018;25(2): 136–145.
22. Tarlock, K., Alonzo, T. A., Gerbing, R. B., Raimondi, S. C., Hirsch, B. A., Sung, L. Pollard, J. A., Aplenc, R., Loken, M. R., Gamis, A. S., Meshinchi, S. Gemtuzumab Ozogamicin Reduces Relapse Risk in FLT3/ITD Acute Myeloid Leukemia: A Report from the Children's Oncology Group. *Clin Cancer Res* 2016;22(8): 1951–1957.
23. Richard F. Schlenk, Verena I. Gaidzik, Peter Paschka, Claudia Leis, Julia Krzykalla, Walter Fiedler, David Nachbaur, Daniela Weber, Thomas Kindler, Michael Lubbert, Mohammed Wattad, Katharina Gotze, Maisun Abu Samra, Silke Kapp-Schwoerer, Thomas Schroeder, Heinz A. Horst, Michael Girschikofsky, Elisabeth Koller, Axel Benner, Felicitas Thol, Michael Heuser, Arnold Ganser, Konstanze Dohner, Hartmut Dohner. Gemtuzumab Ozogamicin in NPM1-Mutated Acute Myeloid Leukemia: Early Results From the Prospective Randomized AMLSG 09–09 Phase III Study. *J Clin Oncol* 2020;38(6):623–632.
24. Niktoreh, N., Lerijs, B., Zimmermann, M., Gruhn, B., Escherich, G., Bourquin, J. P., Dworzak, M., Sramkova, L., Rossig, C., Creutzig, U., Reinhardt, D., Rasche, M. Gemtuzumab ozogamicin in children with relapsed or refractory acute myeloid leukemia: a report by Berlin-Frankfurt-Munster study group. *Haematologica* 2019;104(1): 120–127.

Figures

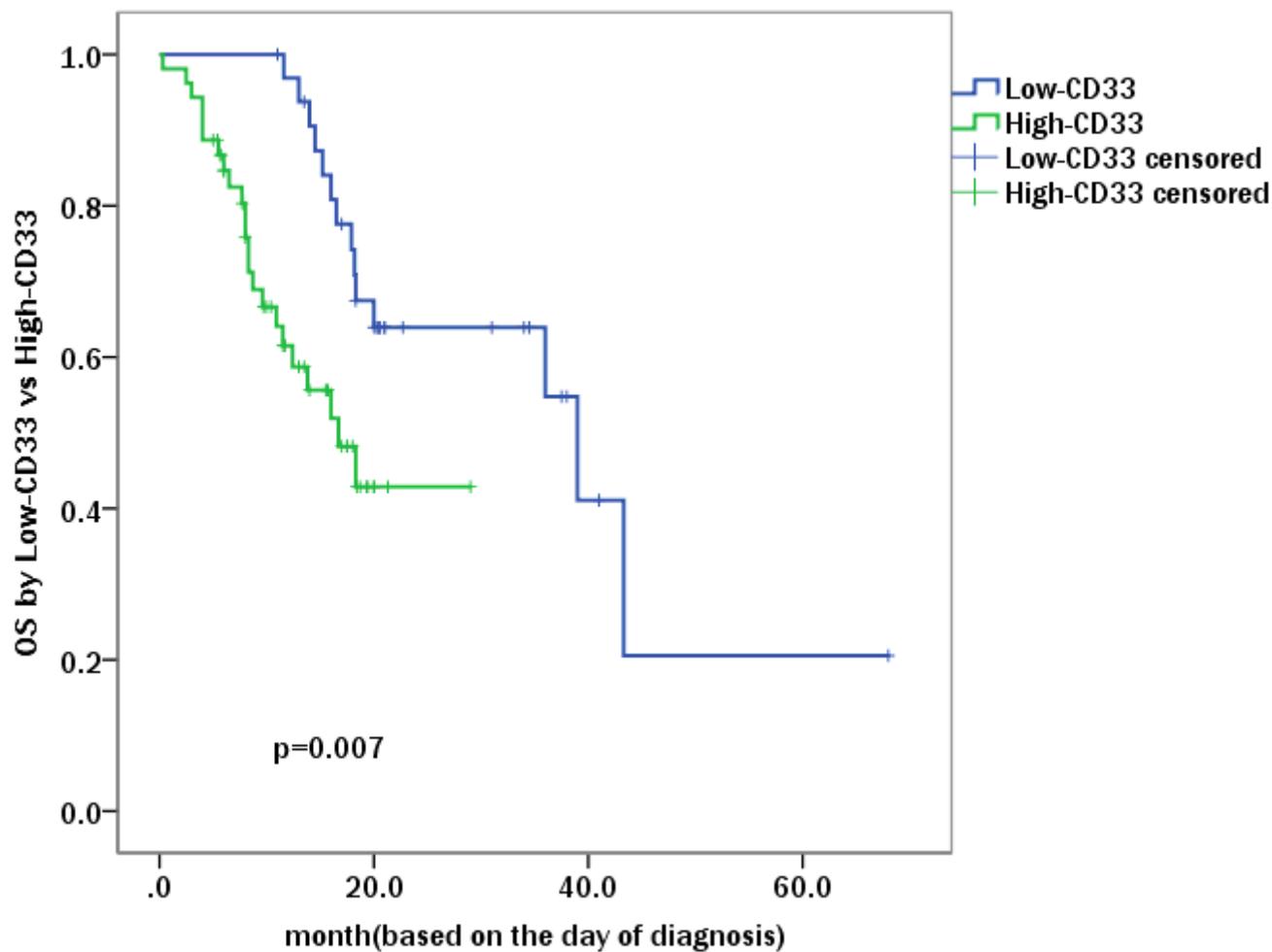


Figure 1

OS of 86 patients with AML based on the expression level of CD33 (low CD33 and high CD33 expression). $P < 0.01$. AML, Acute myeloid leukemia; OS, Overall survival.