

Isolation, characterization and application of theophylline-degrading *Aspergillus* fungi

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Abstract

Background: Caffeine, theobromine and theophylline are main purine alkaloid in tea. Theophylline is the downstream metabolite and stays a very low level in *Camellia sinensis*. In our previous study, *Aspergillus sydowii* could convert caffeine into theophylline in solid-state fermentation of tea through N-demethylation. In this study, tea-derived fungi caused theophylline degradation in the solid-state fermentation. The aim of this study was isolate theophylline-degrading fungi and to investigate their application in production of methylxanthines with theophylline as feedstock through microbial conversion.

Results: Of seven tea-derived fungi collected and identified by ITS, β -tubulin and calmodulin gene sequences, *Aspergillus ustus*, *Aspergillus tamarii*, *Aspergillus niger* and *Aspergillus sydowii* associated with solid-state fermentation of pu-erh tea have shown ability to degrade theophylline. *A. ustus* and *A. tamarii* could degrade theophylline highly significantly ($p < 0.01$) in liquid culture. 1,3-Dimethyluric acid, 3-methylxanthine, 3-methyluric acid, xanthine and uric acid were detected consecutively by HPLC in *A. ustus* and *A. tamarii* respectively. The data from absolute quantification analysis suggest that 3-methylxanthine and xanthine were main degraded metabolites in *A. ustus* and *A. tamarii* respectively. 129.48 ± 5.81 mg/L of 3-methylxanthine and 159.11 ± 10.8 mg/L of xanthine were produced by *A. ustus* and *A. tamarii* respectively in 300 mg/L of theophylline liquid medium.

Conclusions: For the first time, we confirmed that isolated *Aspergillus ustus*, *Aspergillus tamarii* degrade theophylline through N-demethylation and oxidation. We were able to biologically produce 3-methylxanthine and xanthine efficiently from theophylline through a new microbial synthesis platform with *A. ustus* and *A. tamarii* as appropriate starter strains.

Background

Caffeine (1, 3, 7-trimethylxanthine) is the most abundant methylxanthine alkaloids in tea, and also one of the major tea flavor constituents contributing to the bitter taste [1]. Caffeine was extensively established to be the final methylxanthine product biosynthesized through three steps of the methylation of xanthine in the root of tea tree [2]. Until now, the catabolism of caffeine has been relatively understood and established in plants, mainly in tea (*Camellia sinensis*) and coffee (*Coffea arabica*) [3]. The major catabolic pathway of caffeine is as follows: caffeine \rightarrow theobromine/theophylline \rightarrow 3-methylxanthine \rightarrow xanthine \rightarrow uric acid \rightarrow allantoin \rightarrow allantoic acid \rightarrow $\text{CO}_2 + \text{NH}_3$ [4, 5]. The other alternative catabolic pathways have been reported recently in *Camellia* plants [6, 7]. The first demethylation happened at the position N-7 of caffeine and is the speed-limiting step for caffeine catabolism [8]. Theophylline (1,3-dimethylxanthine) is a transient metabolite and stays a very low level due to the slow metabolism in tea leaves [9].

Although caffeine level remains stable in the processing of general teas (green tea, black tea, oolong tea and white tea) [10, 11], several microorganisms have been selected from the soil of tea and coffee plantations to degrade caffeine, which included *Pseudomonas putida* [12, 13], *Pseudomonas lutea* [14], *Serratia marcescens*, *Fusarium solani* [15], *Stemphyllium* sp., *Aspergillus tamarii*, *Penicillium commune*

[16]. Two possible mechanisms of caffeine catabolism in microorganisms are N-demethylation and oxidation [17]. Theophylline is the major metabolite formed in fungi through the N-demethylation of caffeine, which present marked differences from bacteria that theobromine (3,7-dimethylxanthine) or paraxanthine (1,7-dimethylxanthine) are major metabolites in caffeine catabolism [17, 18]. Moreover, *Pseudomonas putida* was established to not only use caffeine, theobromine, paraxanthine (1,7-dimethylxanthine), 7-methylxanthine as sole carbon and nitrogen sources, but also degrade theophylline and 3-methylxanthine [12, 19]. In addition, *Aspergillus niger*, *Talaromyces marneffeii*, and *Talaromyces verruculosus* isolated from cocoa pod husks were established to degrade theobromine and produce methylxanthine (Mensah et al., 2018) [20]. However, only the bacterial strain *Pseudomonas putida* CBB5 was confirmed to degrade theophylline via N-demethylation (Yu et al., 2009; Algharrawi et al., 2015) [19, 21]. Until now, the theophylline-degrading fungi and correlative metabolites were not completely definite.

Pu-erh tea is one of the most representative dark tea and natural microorganisms involved in solid-state fermentation (SSF) play an important role in tea processing [22, 23]. Microorganisms including bacteria and fungi have profound impact on substance metabolisms and correlation with quality formation of pu-erh tea [24, 25, 26]. *Aspergillus niger*, *Aspergillus tubingensis*, *Aspergillus fumigatus*, *Aspergillus acidus*, *Aspergillus awamori*, *Aspergillus tamarii*, *Blastobotrys adenivorans*, *Candida tropicalis*, *Fusarium graminearum*, *Pichia farinosa*, *Rasamsonia byssochlamydoidea*, *Rasamsonia emersonii*, *Rasamsonia cylindrospora*, *Rhizomucor pusillus*, *Rhizomucor tauricus* and *Thermomyces lanuginosus* have been detected in pu-erh tea [27, 28, 29, 30]. Theophylline has several applications in therapeutics, especially as anti-asthmatic, anticancer, anti-cellulite and combinatorial drug [31, 32, 33]. Caffeine content is changing in pu-erh tea, which was associated with the fungi appearing in solid-state fermentation [34, 35, 36, 37, 38]. We found that theophylline content was increased significantly ($p < 0.01$) in the solid-state fermentation and *Aspergillus sydowii* cause caffeine degradation [39]. After in-depth research, *Aspergillus sydowii* has a significant ($p < 0.05$) impact on caffeine metabolism and has potential value in theophylline production through aerobic fermentation [40, 41].

In present study, we found that theophylline content had a highly significant ($p < 0.01$) decrease during the later period of solid-state fermentation of pu-erh after a highly significant ($p < 0.01$) increase. Therefore, except for an isolated caffeine-degrading fungus identified as *Aspergillus sydowii* which caused the production of theophylline in the solid-state fermentation, theophylline-degrading fungi also could be found in the solid-state fermentation. In this paper, two theophylline-degrading fungi were isolated from the solid-state fermentation and identified as *A. ustus* and *A. tamarii* based on colonial morphology and ITS, β -tubulin and calmodulin gene sequences, respectively. Theophylline degradation metabolites and pathways were analyzed in fungi by high-performance liquid chromatography (HPLC). The application in the production of methylxanthines was investigated by using *A. ustus* and *A. tamarii*.

Methods

Materials and Reagents

Sun-dried green tea leaves (*C. sinensis* var. *assamica*) with moisture content 6.25% by weight were obtained from Yunnan province, China. Caffeine, theophylline, 3-methylxanthine, 1-methylxanthine, xanthine, 1,3-dimethyluric acid, 1-methyluric acid, 3-methyluric acid and uric acid were purchased from USA Sigma Company. SP fungal DNA kit was purchased from USA Omega Company. DNA marker, polymerase chain reaction (PCR) spread reagent, primers: ITS1 (5–*TCTAGGTGCTGCGG*–3) and ITS4 (5–*TCTCGCTATGATAGC*–3); Bt2a (5–*GGTCTATCGGTGCTGC*–3) and Bt2b (5–*ACCTCAGTGTAGTGACC*–3); and CF1L (5–*GCTGACTCGTACGAG*–3) and CF4 (5–*ATTTGCATCATGAGCTGC*–3) were purchased from Japan TaKaRa company. Other reagents were either analytical grade or chromatographic grade.

Pu-erh tea solid-state fermentation, and determination of caffeine and theophylline

The solid-state fermentation of pu-erh tea was based on the natural microbiota existing on the leaves and fermentation environment. A 2 kg sample of sun-dried green tea leaves was mixed with 885 mL tap water to achieve given moisture content of 35% (w/w) [29]. During the fermentation, the leaves were mixed to ensure homogeneity and tap water was added to keep the appropriate moisture content at 25-35%. The whole fermentation continued for about 35 days and samples were collected every 5 days. The colony forming units (CFU) was calculated by per gram of the dry weight after 2 days of cultivation at 30 °C. Caffeine and theophylline contents were determined by high-performance liquid chromatography (HPLC) described by Zhou et al., [39, 40] using an Agilent 1200 series system and an Agilent C₁₈ Chromatogram column (250 mm×4.6 mm, 5 μm). The samples collected on day 20 and 25 were stored at 4 °C and selected for further isolation and identification of theophylline-degrading fungi.

Fungal identification of isolates

Fungal strains were isolated using potato dextrose agar (PDA) medium and they were counted by dilution-plating method (Wang, Peng and Gong, 2011). The colony forming units (CFU) was calculated by per gram of sun-dried tea leaves after 2 days of cultivation at 30 °C. Microscopic and morphological examinations of colonies were carried out according to mycological guide, and morphological features of their colonies are recorded in Table 1.

The fresh cells were obtained by centrifugation at 1700 g for 5 min after cultivation in 20 mL of Czapek Dox medium at 30 °C for 2 days on an incubator shaker (250 rpm) and freeze-dried at -80 °C [42]. DNA was extracted by using SP fungal DNA kit. The fungal primers ITS1 and ITS4, Bt2a and Bt2b, and CF1L and CF4 were used in the PCR to amplify the ITS, β-tubulin and Calmodulin regions, respectively [43]. The final volume of 50 μL, 1.0 μL of containing template DNA, 5 μL of 10 x buffer, 5 μL of dNTPs (2.5 mM), 0.5 μL of Taq polymerase, 1.0 μL (10 μM) of each primer, and 36.5 μL of sterile distilled water were used to

implement amplifications [23, 42]. The PCR reaction procedure was as follows. Pre-degeneration at 95 °C for 5 min, degeneration at 94 °C for 1 min, annealing at 54 °C for 1 min, extension at 72 °C for 1.5 min, with 35 cycles, extension at 72 °C for 10 min [44]. It was stored at 10 °C in the end of the reaction process.

Evaluation of growth of isolates in agar mediums

Qualitative screenings were carried out on Petri dishes containing a solid culture medium with dextrose (2 % w/v) (control culture) and a selection medium with theophylline instead of dextrose in three different concentrations: 600, 1200 and 1800 mg/L per plate, respectively [39]. Fungal mycelia were transferred to the surfaces of the agar plates with an inoculating loop (10 ul). Isolates were incubated at 30 °C for 5 days. Compared with the control culture; theophylline utilization capacity of the isolate was estimated by the size of the colony grown on the plates (Table 3).

Assessment of theophylline-degrading fungi in different theophylline liquid mediums

Theophylline liquid medium (TLM) composed of (g/L) NaNO_3 , 4.0; KH_2PO_4 , 1.3; $\text{Na}_2\text{HPO}_4 \cdot 7\text{H}_2\text{O}$, 0.19; MgSO_4 , 0.15; $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$, 0.26; MgSO_4 , 0.15; sucrose, 2.0 and theophylline, 0.3 in distilled water was prepared [45]. The isolate strains were transferred into PDA medium and incubated aerobically at 30 °C for 72 h on an incubator shaker (250 rpm), respectively. The spore suspension was adjusted to 1.0×10^7 CFU/mL for inoculation after eluting by using sterile saline solution with identical theophylline concentration. To investigate the influence of carbon and nitrogen source on theophylline degradation, the modifications used either 5 g/L sucrose or 10 g/L dextrose as carbon source in theophylline liquid medium with sucrose as carbon source (TLM-S) or theophylline liquid medium with dextrose as carbon source (TLM-D), and 1.01 g/L ammonium sulphate as nitrogen source in theophylline liquid medium with ammonium sulphate as nitrogen source (TLM-N), and 5 g/L sucrose and 1.01 g/L ammonium sulphate in theophylline liquid medium with sucrose and ammonium sulphate as carbon and nitrogen sources (TLM-SN), respectively. The spore suspension and liquid mediums were adjusted for pH 6.0 by phosphate buffer. For each isolate, control and experimental mediums (25 mL each) were inoculated with spore suspension with 4% inoculum size (v/v) that 1 mL spore suspension was inoculated into each medium, and biocidal treatment was defined as the control. Theophylline concentration was determined after cultivation at 30 °C for 5 days on an incubator shaker (130 rpm), respectively.

Analysis of theophylline degradation metabolites by selected isolates

Through comparison, sucrose as carbon source could promote theophylline degradation. Therefore, TLM-S was chosen as the optimal medium to analyze theophylline degradation by selected isolates. A series of TLM-S mediums with different initial theophylline concentrations (100, 200 and 300 mg/L, respectively) were set up each day of incubation and a 7-day period cultivation of each selected isolates were carried out on an incubator shaker (130 rpm, 30 °C). At intervals of up to 24 h for 7 days, an aliquot of each

culture was filtered through a 0.45 µm syringe filter, and theophylline concentration and related metabolites were determined by high-performance liquid chromatography (HPLC) according to the method from Mensah et al. [20].

Standard calibration curves were prepared from solutions of theophylline, 3-methylxanthine, 1-methylxanthine, xanthine, 1,3-dimethyluric acid, 1-methyluric acid, 3-methyluric acid and uric acid. 2 mL aliquots from each collected culture were filtered and analyzed by HPLC for theophylline, 3-methylxanthine, 1-methylxanthine, xanthine, 1,3-dimethyluric acid, 1-methyluric acid, 3-methyluric acid and uric acid. Internal standard method was used to aid in identification of metabolites. The concentrations of main degradation products (3-methylxanthine and xanthine) were analyzed in inoculated culture medium by selected isolates.

Influence of selected isolates on 3-methylxanthine and xanthine

Analysis of theophylline degradation products showed that N-demethylation was the main theophylline degradation pathway in fungi, and 3-methylxanthine and xanthine were main demethylated products. To explore the effects of selected isolates on 3-methylxanthine and xanthine, 3-methylxanthine and xanthine liquid mediums were prepared as above described with 5 g/L sucrose as carbon source and 0.3 g/L 3-methylxanthine or 0.3 g/L xanthine, respectively. Each isolate was inoculated with 4% inoculum size (v/v) and biocidal treatment was defined as the control. 3-Methylxanthine and xanthine concentrations were determined by HPLC after cultivation at 30 °C for 5 days on an incubator shaker (130 rpm), respectively.

Statistical analysis

Three biological replications were carried out to ensure validity and repeatability. All data are presented as mean value ± standard deviation (SD). The independent t-test using SPSS 20.0 for window was carried out to determine whether the significant difference at $p < 0.05$ level or the highly significant difference in $p < 0.01$ level exist by using one-way analysis of variance (one-way ANOVA) .

Results

Theophylline degradation exists in solid-state fermentation of pu-erh tea

Fungi count, caffeine and theophylline contents were determined in the natural solid-state fermentation of pu-erh tea, and results are presented in Figure 1. Fungi count (Fig. 1A) dramatically increased from day 0 to 10 and then increased slowly before day 20. After day 20, fungi count maintained a high level over 1.0×10^5 CFU/g. Because of the metabolic activity of fungi, caffeine content (Fig. 1B) was decreased highly significantly ($p < 0.01$) from 36.85 ± 1.02 mg/g to 25.46 ± 1.85 mg/g during fermentation. Theophylline content (Fig. 1C) was increased highly significantly ($p < 0.01$) before day 20, which confirmed that

caffeine-degrading fungi led to caffeine degradation and theophylline production. In our previous study [40], it was found that *Aspergillus sydowii* convert caffeine to theophylline. However, after day 20, theophylline content had a highly significant ($p < 0.01$) decrease from 11.18 ± 1.10 mg/g to 5.89 ± 0.65 mg/g, which shown that theophylline degradation appeared in solid-state fermentation except for caffeine degradation. Therefore, in consideration of fungal community, there existed theophylline-degrading fungi in the solid-state fermentation, which could be *Aspergillus sydowii* or other fungi.

Isolation and identification of theophylline-degrading fungi

Based on colony morphology, eleven filamentous fungi were initially selected and isolated from pu-erh tea. Among them, seven fungi were superior in number and coded orderly with numbers PT-1 to PT-7. Distinctive morphological features of the seven isolates were observed after cultivation at 30 °C for 5 days and documented in Table 1.

Table 1 Colony characteristics of theophylline-degrading fungi

Isolate	Shape	Surface	Colour	Exudate	Reference
PT-1	Circular	Rough	Black	None	[39]
PT-2	Circular	Rough	Olive green	Red-coloured	[39]
PT-3	Circular	Rough	Dark yellow colonies with white edges	Yellow sclerotium	[40]
PT-4	Irregular	Rough	Light yellow	Yellow sclerotium	[40]
PT-5	Circular	Rough	Greyish-green centre with yellow patches	Red pigment	[40]
PT-6	Circular	Rough	Iron gray bulge with milk white edges	None	Figure S1
PT-7	Irregular	Rough	Hazel green with gray back	None	Figure S2

The amplified sequences of ITS region were produced with sizes ranging between 502 and 546 base pairs. The amplified sequences of β -Tubulin region were produced with sizes ranging between 420 and 694 base pairs. The amplified sequences of Calmodulin region were produced with sizes ranging between 715 and 765 base pairs. Based on the DNA sequences (Table 2), seven dominating isolates were belonged to 6 *Aspergillus* spp. and 1 *Penicillium* sp., respectively. Through Neighbor-Joining analysis for both new fungi (Additional file: Figure S5), in the phylogram for *Aspergillus* species (Figure S5a and S5b), strain PE-6 was clustered with *Aspergillus ustus* and showed a 100% of identity to the tested *Aspergillus ustus* NRRL275; additionally, PE-7 strain was closely related to *Aspergillus tamarisii* NRRL20818 with 99.9% of identity.

Table 2 Identification of theophylline-degrading fungi by sequence determination

Isolate	Primers	Fragments (bp)	Species	Strain number	identity	Reference
PT-1	ITS1/ITS4	546	<i>Aspergillus niger</i>	NCBT110A	99.8%	[39]
PT-2	ITS1/ITS4	516	<i>Aspergillus sydowii</i>	NRRL250	99.8%	[39]
PT-3	TS1/ITS4	541	<i>Aspergillus pallidofulvus</i>	NRRL4789	99.9%	[40]
	Bt2a/Bt2b	516				
	CF1L/CF4	765				
PT-4	TS1/ITS4	532	<i>Aspergillus sesamicola</i>	CBS137324	99.8%	[40]
	Bt2a/Bt2b	515				
	CF1L/CF4	757				
PT-5	TS1/ITS4	525	<i>Penicillium manginii</i>	CBS253.31	99.6%	[40]
	Bt2a/Bt2b	420				
PT-6	TS1/ITS4	502	<i>Aspergillus ustus</i>	NRRL275	100%	Figure S3
	Bt2a/Bt2b	694				
PT-7	TS1/ITS4	532	<i>Aspergillus tamarisii</i>	NRRL20818	99.9%	Figure S4
	Bt2a/Bt2b	476				
	CF1L/CF4	715				

Evaluation results of theophylline-degrading fungi in solid mediums

The screening was carried out in agar solid mediums for the evaluation of the biocatalytic potential for the degradation of theophylline. All isolate tea-derived strains were inoculated into an agar solid medium with the presence of dextrose and they were also inoculated into a set of agar solid mediums with increasing theophylline concentrations. The colony diameters of potential theophylline-degrading fungi were measured and showed in Table 3.

Table 3 Growth of tea-derived fungi in agar solid medium (2 % w/v) with dextrose (2 % w/v) (control culture) or presence of theophylline instead of dextrose (30 °C, 5 d, pH 7.0).

Isolate fungi	Colony diameter (cm)			
	Control culture	600mg/L theophylline	1200 mg/L theophylline	1800 mg/L theophylline
<i>A. niger</i>	3.5 ± 0.5	0.5 ± 0.2	No growth	1.0 ± 0.5
<i>A. sydowii</i>	2.5 ± 1.0	0.5 ± 0.1	1.0 ± 0.3	0.5 ± 0.3
<i>A. pallidofulvus</i>	3.0 ± 0.5	No growth	No growth	0.5 ± 0.1
<i>A. sesamicola</i>	3.0 ± 0.5	No growth	No growth	0.5 ± 0.1
<i>P. mangini</i>	3.0 ± 1.0	No growth	No growth	No growth
<i>A. ustus</i>	2.5 ± 0.5	1.0 ± 0.3	1.5 ± 0.4	1.5 ± 0.4
<i>A. tamaraii</i>	3.0 ± 0.5	2.0 ± 0.5	2.5 ± 0.5	3.5 ± 1.0

Six tea-derived strains could survive in the agar solid mediums (2% w/v) with theophylline alone. *Aspergillus* spp. showed a better growth in higher evaluated concentrations. Particularly, *A. niger*, *A. sydowii*, *A. ustus* and *A. tamaraii* had growth in low theophylline concentration, which showed that these strains had a high utilization ratio of theophylline. A higher growth in lower concentration indicated a higher utilization of fungi direct or indirect [46]. Therefore, *A. niger*, *A. sydowii*, *A. ustus* and *A. tamaraii* were considered as the potential theophylline-degrading fungi.

Selection of theophylline-degrading fungi and optimal medium in liquid culture

For theophylline biodegradation in liquid culture, with biocidal treatment as the control, seven isolates were inoculated into the basic mediums with the presence of theophylline and sucrose or dextrose as carbon source, or ammonium sulphate as nitrogen source, respectively. Theophylline concentration was determined after cultivation at 30 °C for 5 days by HPLC and results are showed in Fig. 2. Through comparisons of each isolate, *A. ustus* and *A. tamaraii* could decrease theophylline concentration high significantly ($p < 0.01$) by about 65.15% and 95.98% in TLM-S medium, respectively. In addition, under the effect of *A. niger* and *A. sydowii*, theophylline concentrations were decreased slightly by about 1.03% and 5.19% in TLM-S medium, respectively, which showed that the theophylline utilizations of *A. niger* and *A. sydowii* were restricted. *A. pallidofulvus*, *A. sesamicola* and *P. mangini* had no significant ($p > 0.05$) impact on theophylline degradation. Hence, *A. ustus*, *A. tamaraii*, *A. niger* and *A. sydowii* were selected as the potential theophylline-degrading fungi for theophylline degradation in liquid culture.

Theophylline could be used as the sole carbon and nitrogen source. The presence of other carbon or nitrogen sources had a significant impact on theophylline degradation and pathway. The optimum liquid

medium was chosen by comparing theophylline degradation capacity in different mediums. In contrast with other mediums (TLM-D, TLM-N and TLM-SN), theophylline degradation level had a highly significant ($p < 0.01$) improvement in TLM-S medium, which showed that carbon source particularly sucrose promoted theophylline degradation and nitrogen source restrained theophylline degradation to a certain extent. Therefore, TLM-S medium was chosen to analyze the characterization of theophylline degradation in liquid culture.

Characterization of theophylline degradation by theophylline-degrading fungi

A. ustus, *A. tamarii*, *A. niger* and *A. sydowii* were inoculated into TLM-S mediums with increasing theophylline concentrations (100, 200 and 300 mg/L, respectively), and the Tissue-culture bottles were incubated in an orbital shaker (130 rpm, 30 °C). The inoculation bottles were taken every 24 h for the determination of theophylline and related metabolites by HPLC, and results are presented in Fig. 3. Under the effect of *A. ustus* and *A. tamarii*, theophylline content was decreased highly significantly ($p < 0.01$) in all substrate concentrations. However, the concentration of theophylline inoculated by *A. niger* and *A. sydowii* was slightly decreased ($p > 0.05$) in all substrate concentrations. Therefore, *A. ustus* and *A. tamarii* had an advantage in theophylline degradation than *A. niger* and *A. sydowii*. Both *A. ustus* and *A. tamarii* could degrade theophylline completely in low concentration (100 mg/L theophylline). However, *A. ustus* only degraded 79.00% theophylline in high concentration (300 mg/L theophylline), while *A. tamarii* could degrade theophylline almost completely in all concentrations, which showed that *A. tamarii* had a higher theophylline degradation capacity.

A series of experiments was conducted to find out the theophylline degradation pathway through the identification of catabolic intermediates by HPLC using an internal standard method (Table 4). Related degradation metabolites were not found in the liquid culture inoculated by *A. niger* and *A. sydowii*. 1,3-Dimethyluric acid, 3-methylxanthine, 3-methyluric acid, xanthine and uric acid were detected consecutively in the liquid culture. 3-Methylxanthine was common and main metabolite through N-demethylation at the position N-1 of theophylline in *A. ustus* and *A. tamarii*. Xanthine was the further demethylated metabolite in theophylline degradation found in *A. ustus* and *A. tamarii* through N-demethylation at the position N-3 of 3-methylxanthine. In contrast to *A. ustus* isolate that additional metabolites including 1,3-dimethyluric acid and 3-methyluric acid were identified in the culture through the oxidation of theophylline and 3-methylxanthine, respectively [19]. Only uric acid was identified in *A. tamarii* culture as the oxidation product of xanthine, which showed the differences in degradation metabolites and pathways between *A. ustus* and *A. tamarii*.

Table 4 Theophylline degradation metabolites detected in the liquid media inoculated by *Aspergillus* fungi

Metabolite	Fungal isolates			
	<i>A. ustus</i>	<i>A. tamarii</i>	<i>A. niger</i>	<i>A. sydowii</i>
1,3-dimethyluric acid	+	-	-	-
1-methylxanthine	-	-	-	-
3-methylxanthine	+	+	-	-
1-methyluric acid	-	-	-	-
3-methyluric acid	+	-	-	-
xanthine	+	+	-	-
uric acid	-	+	-	-

TLM-S media were analyzed by HPLC for 1,3-dimethyluric acid, 1-methylxanthine, 3-methylxanthine, 1-methyluric acid, 3-methyluric acid, xanthine and uric acid.

Production of 3-methylxanthine or xanthine through theophylline degradation

Several xanthine derivatives including 3-methylxanthine have been synthesized chemically for use in medical industry [47]. Except for engineering a microbial platform for *de novo* biosynthesis of diverse methylxanthins [48], bioconversion from cheaper feedstocks such as caffeine, theophylline and theobromine was an effective pathway to produce high value methylxanthines via metabolically engineered microorganisms [21]. In this study, *A. ustus* and *A. tamarii* could degrade theophylline, and 3-methylxanthine and xanthine were detected as degradation metabolites. The microbial utilization of 3-methylxanthine and xanthine were investigated as substrate through liquid culture of isolates. The concentrations of 3-methylxanthine and xanthine were determined by HPLC after cultivation for 5 days. As shown in Fig. 4, *A. ustus* and *A. tamarii* had a significant ($p < 0.05$) or a highly significant ($p < 0.01$) impact on 3-methylxanthine degradation with a removal ratio of about 27.05% and 84.29%, respectively. Associated with the metabolites in theophylline degradation, 3-methyluric acid and xanthine were 3-methylxanthine degradation metabolites through oxidation and N-demethylation, respectively.

Although *A. ustus* and *A. tamarii* could utilize 3-methylxanthine and xanthine significantly, 3-methylxanthine and xanthine concentrations were accumulated largely in theophylline degradation. To investigate the application in the production of 3-methylxanthine and xanthine using theophylline-degrading fungi with theophylline as feedstock, the quantitative determination of 3-methylxanthine and xanthine were carried out in all theophylline concentrations inoculated by *A. ustus* and *A. tamarii*, respectively. The concentrations of 3-methylxanthine in *A. ustus* and *A. tamarii* cultures are presented in Fig. 5. We monitored the accumulation of 3-methylxanthine over the course of inoculated culture by *A. ustus* and *A. tamarii*. 3-Methylxanthine was detected in the culture medium after 24 h for the first time, and increased significantly with cultivation. Over a 7-day period cultivation of *A. ustus* (Fig. 5a), 49.68 ± 2.97

mg/L, 83.82 ± 3.35 mg/L and 129.48 ± 5.81 mg/L of 3-methylxanthine were accumulated and increased significantly with increasing initial theophylline concentrations, respectively. Due to high degradation capacity of 3-methylxanthine in *A. tamarii* culture, 3-methylxanthine concentration (Fig. 5b) stayed at a low level that only 56.72 ± 5.81 mg/L of 3-methylxanthine was accumulated in 300 mg/L of theophylline after a 7-day period cultivation. Hence, strain *A. ustus* exhibited a continuing accumulation of 3-methylxanthine over the course of liquid culture, and increasing initial theophylline concentrations could improve the production of 3-methylxanthine.

It is clear from the data presented in Fig. 6 that a reaction containing theophylline in *A. tamarii* culture provided linear conversion of theophylline to xanthine. Over a 7-day period cultivation of *A. tamarii*, 35.88 ± 6.65 mg/L, 103.95 ± 4.82 mg/L and 159.11 ± 10.8 mg/L of xanthine were accumulated and increased significantly with increasing initial theophylline concentrations through N-demethylation at the position N-3 of 3-methylxanthine, respectively. Therefore, xanthine was main metabolite in theophylline degradation over the course of *A. tamarii* liquid culture, which showed that *A. tamarii* could be used for the production of xanthine with theophylline as feedstock through the microbial conversion.

Discussion

Although caffeine and related methylxanthines are toxic to most bacteria and invertebrates [49], some bacteria and fungi have evolved the ability to metabolize caffeine [12, 15]. The certain microorganisms cause the decrease in caffeine level occasionally [36, 37]. In our previous studies [39, 40], it is found that caffeine content was decreased significantly and *A. sydowii* led to caffeine degradation. As the main metabolite, theophylline content was increased significantly and became the most abundant methylxanthine alkaloids instead of caffeine in pu-erh tea [41]. In this study, we confirmed that *A. sydowii* convert most of degraded caffeine to theophylline; otherwise, a new phenomenon (Fig. 1) that theophylline degradation was found in solid-state fermentation of pu-erh tea. This suggested that potential theophylline-degrading fungi utilizing theophylline as carbon or nitrogen sources could be found in fungal community.

Molecular identification of fungi is mostly dependent on PCR amplified sequences of ITS, β -Tubulin and Calmodulin genes, particularly the genera *Aspergillus* and *Penicillium* [50, 51]. The amplified sequencing of ITS, β -Tubulin and Calmodulin regions, and colony morphology indicated that their seven candidate isolates were belonged to the *Aspergillus* and *Penicillium* genera, and identified specifically as *A. niger*, *A. sydowii*, *A. pallidofulvus*, *A. ustus*, *A. sesamicola*, *A. tamarii* and *P. mangini*. This was in line with the observation in literature that *Aspergillus* spp. and *Penicillium* spp. are amongst the filamentous fungal genera commonly associated with pu-erh tea [52, 53,54], particularly *A. niger*, *A. sydowii* and *A. tamarii* having been widely reported as the dominant fungi in the solid-state fermentation of pu-erh tea.

The assessment in agar mediums with theophylline as sole carbon and nitrogen sources showed that six *Aspergillus* isolates could survive in theophylline agar medium, which suggested that those 6 candidate isolates (Table 3) used theophylline as carbon or nitrogen sources directly or indirectly. The data from the liquid culture inoculated by seven isolates with biocidal treatment as the control indicated that *A. ustus*

and *A. tamarii* could degrade theophylline significantly ($p < 0.05$) or highly significantly ($p < 0.01$); moreover, the theophylline utilizations of *A. niger* and *A. sydowii* were restricted. Though the effect of C/N ratio on growth is strain-dependent [55], increasing C/N ratio would generally favour fungal growth, which influenced microbial metabolism to a certain degree. Through comparisons of theophylline degradation capacities in different theophylline liquid mediums containing sucrose or dextrose as carbon source, or ammonium sulphate as nitrogen source, sucrose enhanced theophylline degradation highly significantly ($p < 0.01$). Therefore, TLM-S medium was the optimization for theophylline degradation and chose to analyze theophylline degradation metabolites in liquid culture.

Except for caffeine, theophylline and theobromine are main purine alkaloid in tea, which both have close connection with caffeine metabolism that theophylline is catabolite of caffeine and theobromine is precursor of caffeine biosynthesis in *Camellia sinensis*. As mentioned above, the pathway of caffeine degradation metabolism is comparatively clear in microorganisms that both N-demethylation and oxidation were found in caffeine degradation, and N-demethylation was main pathway in fungi [17, 18, 56]. In addition, several bacteria and fungi have been shown to be able to utilize or degrade theobromine, which include *Pseudomonas putida*, *Aspergillus niger*, *Talaromyces marneffeii*, and *Talaromyces verruculosus* [12, 18, 20]. Mensah et al. confirmed the existence of subsequent demethylation and oxidation in theobromine degradation through the detection of correlative metabolites by HPLC under the effects of theobromine-degrading fungi [20]. However, theophylline degradation pathways and metabolites were not completely clear in fungi. In this study, it is confirmed that *A. ustus* and *A. tamarii* isolated from pu-erh tea could degrade theophylline in liquid culture. Based on the downstream metabolites detected in the liquid media, theophylline degradation pathways were established and shown in Fig. 7., which showed that both N-demethylation and oxidation were theophylline degradation pathways found in *A. ustus* and *A. tamarii*. Theophylline and related demethylated metabolites can be oxidized to 1,3-dimethyluric acid, 3-methyluric acid and uric acid, respectively. The differences in the degradation metabolites suggested that *A. ustus* and *A. tamarii* were different in degradation pathways, except the common pathway that theophylline \rightarrow 3-methylxanthine \rightarrow xanthine. Therefore, N-demethylation was main theophylline degradation pathway, which was similar to caffeine catabolism and theobromine catabolism in fungi.

3-Methylxanthine and xanthine were common downstream metabolites detected in *A. ustus* and *A. tamarii* cultures. 3-Methylxanthine and other xanthine derivatives have been shown various biomedical effects as adenosine antagonist and inhibitors of Primary Amine Oxidase [57, 58]. Besides chemical synthesis, researchers produced methylxanthines in *Escherichia coli* strains that converts fed theophylline to 3-methylxanthine, or using the microbial host *Saccharomyces cerevisiae* through *de novo* biosynthesis [21, 48]. Due to theophylline degradation characteristic, *A. ustus* and *A. tamarii* would be applied in the production of 3-methylxanthine and xanthine with theophylline as feedstock through microbial conversion. The differences in accumulated concentrations of 3-methylxanthine and xanthine suggested that *A. ustus* benefits the production of 3-methylxanthine, while *A. tamarii* benefits the production of xanthine. After a 7-day period cultivation in 300 mg/L of theophylline liquid medium, *A. ustus* could produce 129.48 ± 5.81 mg/L of 3-methylxanthine, and *A. tamarii* could produce 159.11 ± 10.8 mg/L of xanthine, respectively.

Conclusions

Our present work describes theophylline degradation pathways in tea-derived fungi, and explores the application in production of methylxanthines. *A. ustus* and *A. tamarii* isolated from solid-state fermentation of pu-erh tea and identified based on ITS, β -tubulin and calmodulin gene sequences, were confirmed to degrade theophylline significantly ($p < 0.01$) in liquid culture through the sequential selections. Extensive experiments were carried out to detect related degradation metabolites by using HPLC, finding N-demethylation and oxidation in theophylline catabolism. Through the absolute quantitative detection, it is showed that 3-methylxanthine and xanthine were main degraded metabolites in *A. ustus* and *A. tamarii* respectively, which suggests that *A. ustus* benefits the production of 3-methylxanthine, while *A. tamarii* benefits the production of xanthine with theophylline as feedstock. This reports theophylline degradation pathway in *Aspergillus* fungi and represents a new microbial synthesis platform for production of methylxanthines using theophylline through the inoculation of *A. ustus* and *A. tamarii* respectively.

Abbreviations

SSF, solid-state fermentation; CFU, colony forming units;

HPLC, high-performance liquid chromatography; PDA, potato dextrose agar;

PCR, polymerase chain reaction; ITS, internal transcribed spacer;

dNTPs: deoxy-ribonucleoside triphosphates; TLM, theophylline liquid medium;

TLM-S, theophylline liquid medium with sucrose as carbon source;

TLM-D, theophylline liquid medium with dextrose as carbon source;

TLM-N, theophylline liquid medium with ammonium sulphate as nitrogen source;

TLM-SN, theophylline liquid medium with sucrose and ammonium sulphate as carbon and nitrogen sources; SPSS: Statistical product and service solutions; SD, standard deviation; ANOVA: Analysis of variance.

Declarations

Ethics approval and consent to participate

Not applicable

Consent for publication

Not applicable

Availability of data and materials

The data that support the findings of this study are available from the corresponding author upon reasonable request.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

ZBX, MCQ and XT designed the study. ZBX and MCQ carried out the laboratory work for the study. LXH and ZCQ contributed to HPLC work. MCQ and WTT contributed to the fungal identification of the isolates. LXH contributed to the writing of the manuscript.

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Additional Files

Additional File 1: Figure S1. Colony characteristics (a, b) and conidial structure (c, d) of strain PE-6.

(a): Front on PDA medium at 25 °C for 7 days. (b): Back on PDA medium at 25 °C for 7 days.

(c) Mature conidia heads, conidia stems and antipodal cells (200×). (d): Hyohae, conidia stems and conidiums (400×).

Additional File 1: Figure S2. Colony characteristics (a, b) and conidial structure (c, d) of strain PE-7.

(a): Front on PDA medium at 25 °C for 7 days. (b): Back on PDA medium at 25 °C for 7 days.

(c) Conidia heads(200×). (d): Conidia stems, sterigmas and conidiums (400×).

Additional File 1: Figure S3. The received sequences of strain PT-6 (502bp ITS sequence and 694 bp Calmodulin sequence).

Additional File 1: Figure S4. The received sequences of strain PT-7 (532bp ITS sequence,476bp β -tubulin sequence and 715bp Calmodulin sequence).

Additional File 1: Figure S5. Neighbor-Joining consensus trees of **a** *Aspergillus ustus* PE-6 and **b** *Aspergillus tamarisii* PE-7.

The identification was based on the ITS and Calmodulin genes for *A. ustus* PE-6, and the ITS, β -tubulin and Calmodulin genes for *A. tamarii* PE-7. The numbers over branches represent bootstrap confidence values (%) based on 1000 replicates. The scale bar denotes the nucleotide substitution per sequence.

Figures

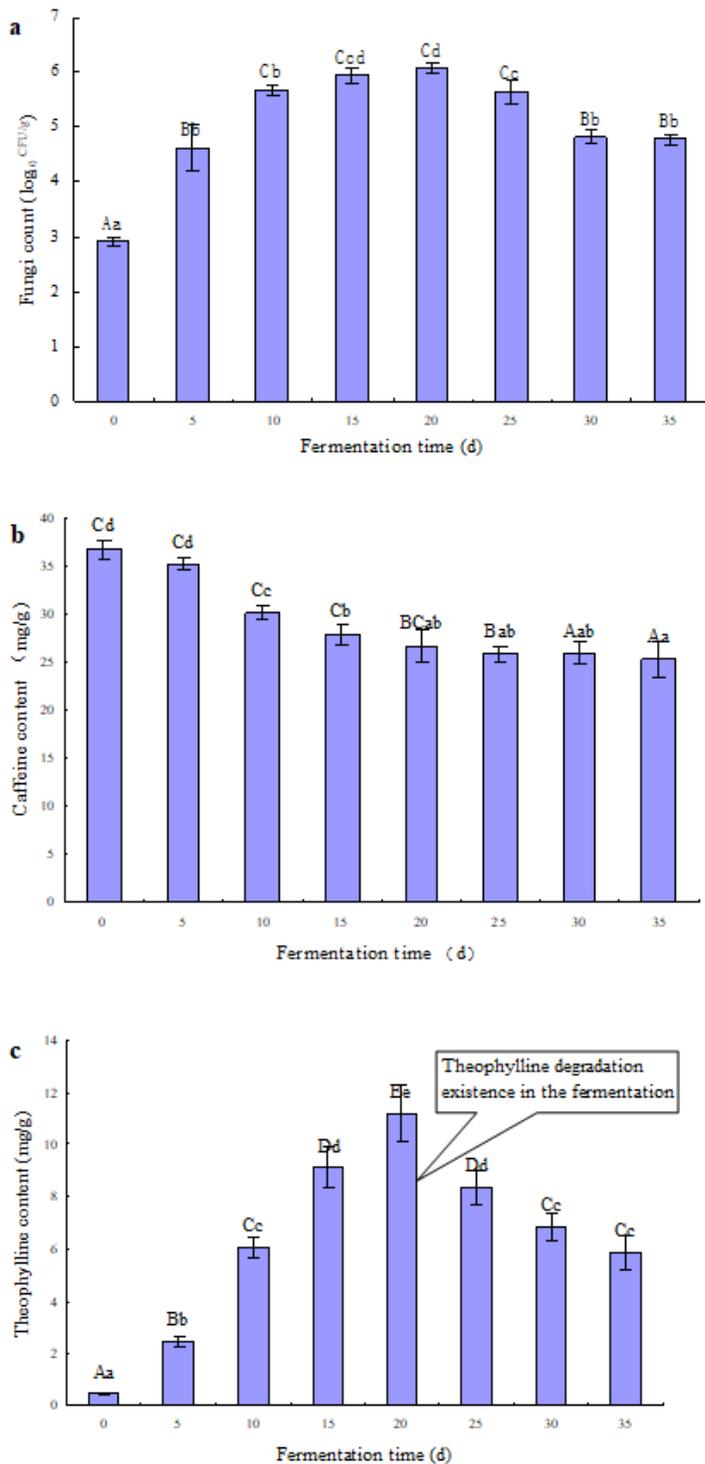


Figure 1

Changes of fungi count (a), caffeine content (b) and theophylline content (c) during the solid-state fermentation of pu-erh tea. All data were present by mean value \pm SD of three replications. The lowercase letters indicated a significant difference at $p < 0.05$ levels and the uppercase letters indicated a highly significant difference at $p < 0.01$ levels using the independent t-test of SPSS 20.0. The different letters show significant differences.

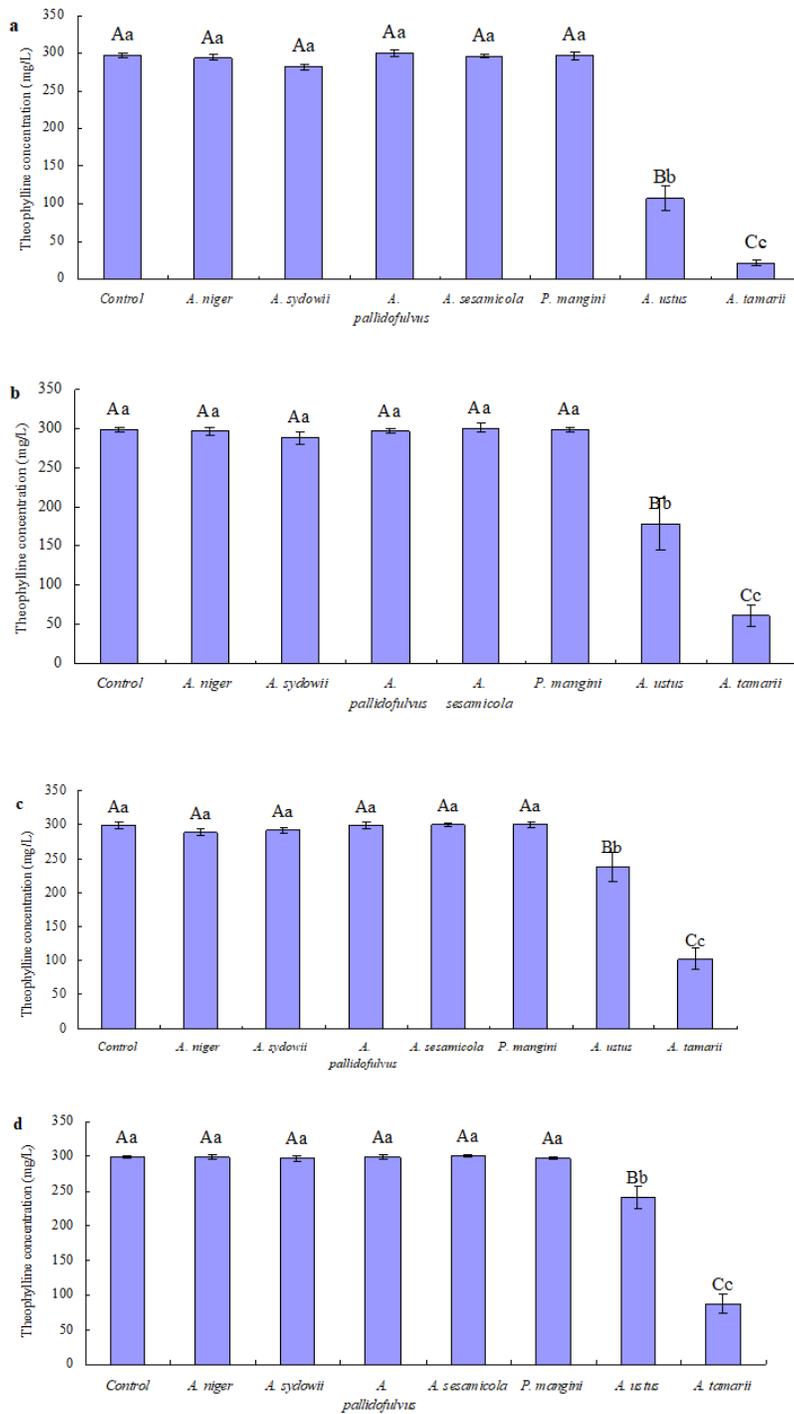


Figure 2

Theophylline degradation capacity of isolate fungi strains in different theophylline liquid mediums. (a) TLM-S = theophylline liquid medium with sucrose as carbon source; (b) TLM-D = theophylline liquid medium with dextrose with sucrose as carbon source; (c) TLM-N = theophylline liquid medium with ammonium sulphate as nitrogen source; (d) TLM-SN = theophylline liquid medium with sucrose and ammonium sulphate as carbon and nitrogen sources, respectively. Biocidal treatment was defined as the control. All data were present by mean value \pm SD of three replications. The lowercase letters indicated a significant difference at $p < 0.05$ levels and the uppercase letters indicated a highly significant difference at $p < 0.01$ levels using the independent t-test of SPSS 20.0. The different letters show significant differences.

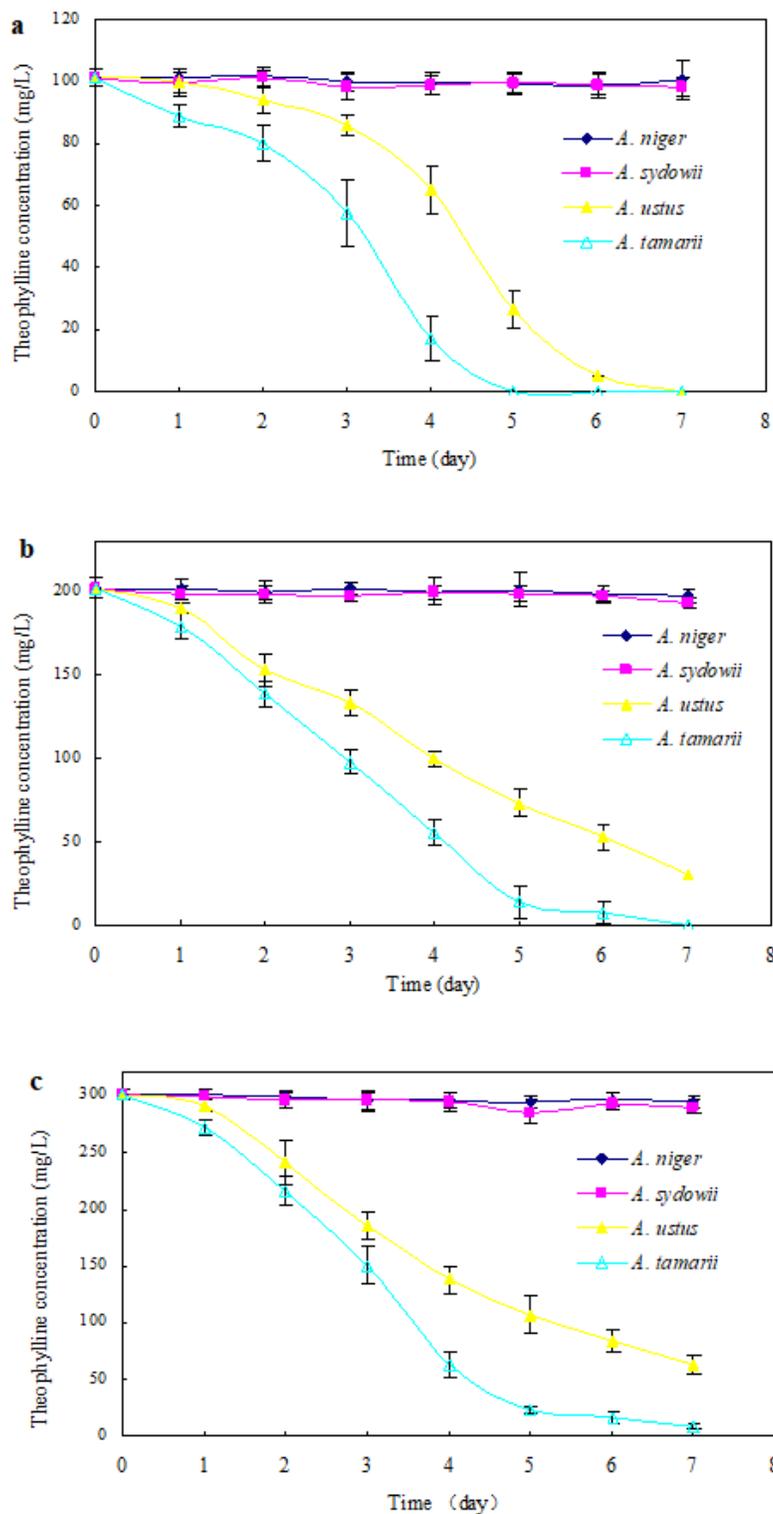


Figure 3

Effect of potential theophylline-degrading fungi on theophylline catabolism in different substrate concentrations. Theophylline concentrations were 100 mg/L (a), 200 mg/L (b), and 300 mg/L (c), respectively. All data were presented by mean value \pm SD of three replications.

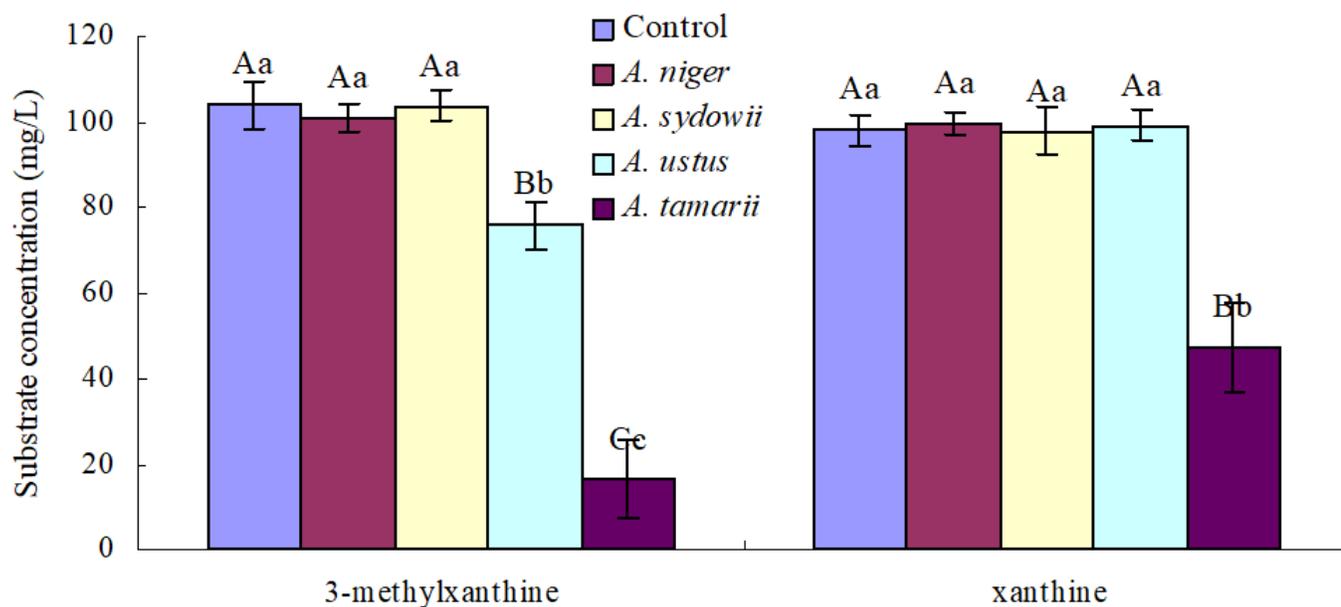


Figure 4

Effect of potential theophylline-degrading fungi on 3-methylxanthine and xanthine, respectively. Biocidal treatment was defined as the control. All data were present by mean value \pm SD of three replications. The lowercase letters indicated a significant difference at $p < 0.05$ levels and the uppercase letters indicated a highly significant difference at $p < 0.01$ levels using the independent t-test of SPSS 20.0. The different letters show significant differences.

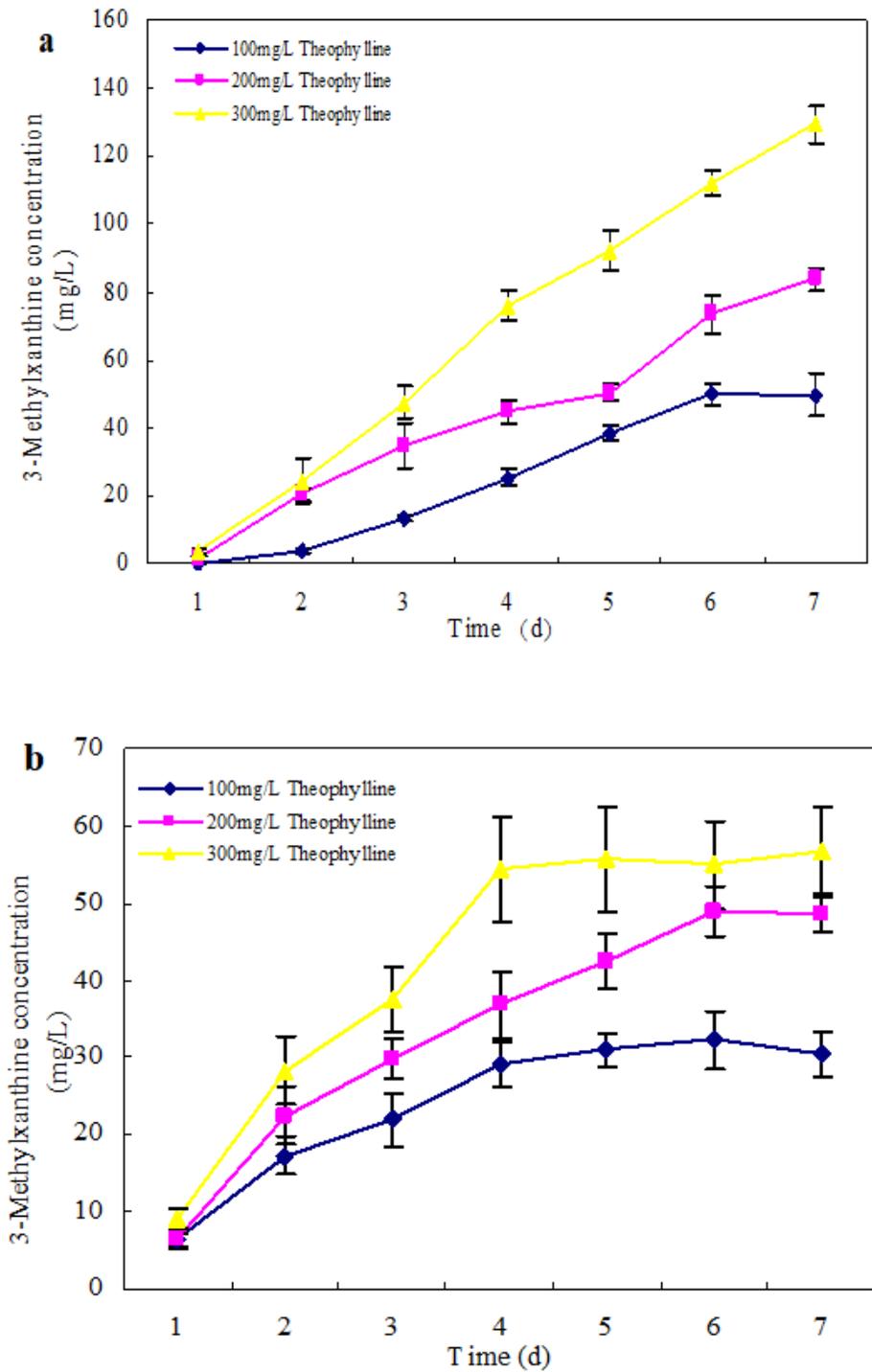


Figure 5

Effects of theophylline concentrations on 3-methylxanthine production by theophylline-degrading fungi. Liquid culture assays were performed using TLM-S mediums with different theophylline concentrations inoculated by *Aspergillus ustus* (a) and *Aspergillus tamarii* (b), respectively. Theophylline concentrations were 100 mg/L (filled rhombus), 200 mg/L (filled square), and 300 mg/L (filled triangle). Concentrations of 3-methylxanthine were present by mean value \pm standard deviations (SD) of three replications.

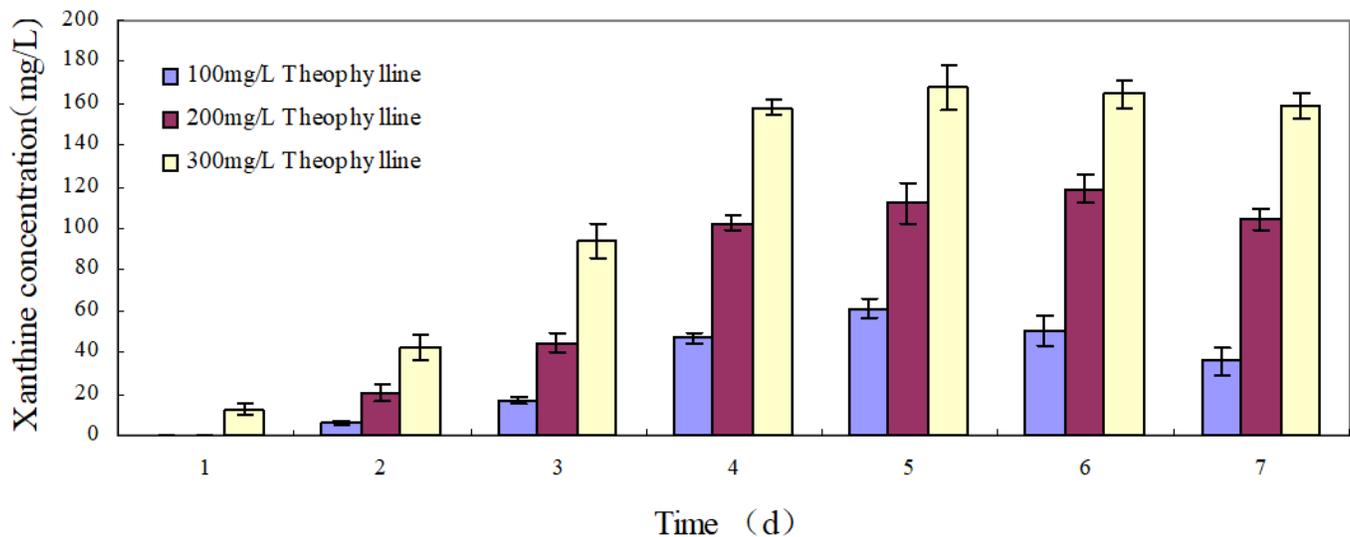


Figure 6

Effects of theophylline concentrations on xanthine production by *Aspergillus tamarii*. Liquid culture assays were performed using TLM-S mediums with different theophylline concentrations inoculated by *Aspergillus tamarii*, respectively. Concentrations of xanthine were present by mean value \pm standard deviations (SD) of three replications.

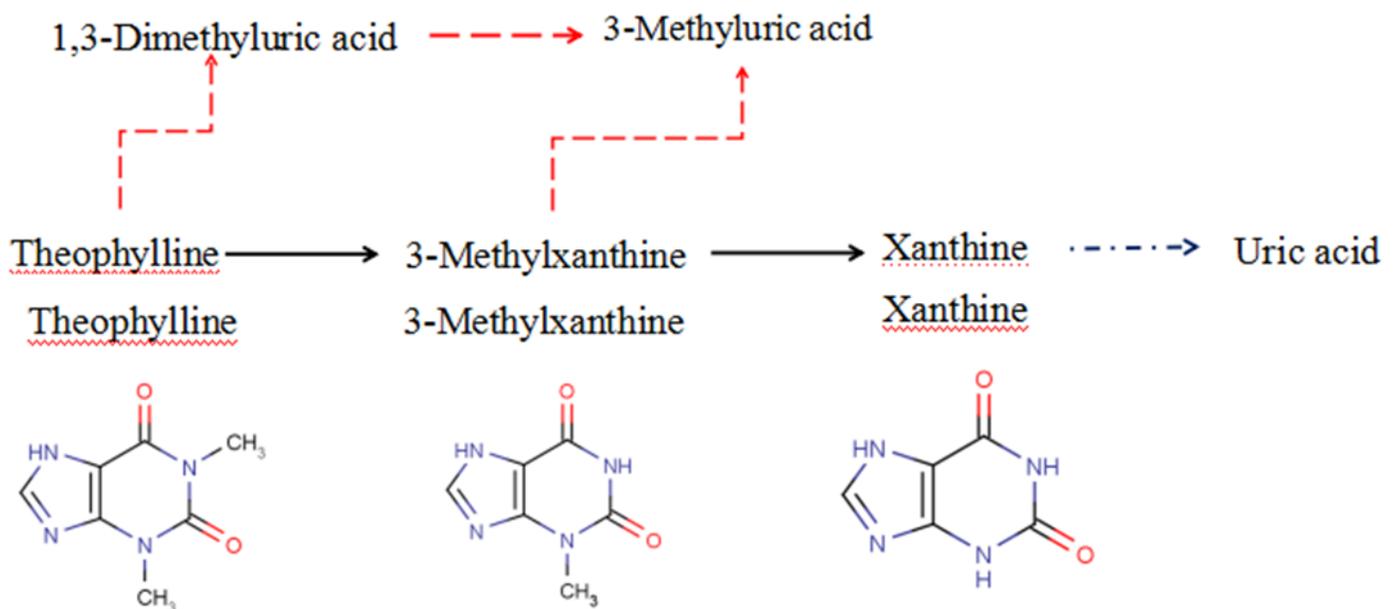


Figure 7

Theophylline degradation pathways employed by *A. ustus* and *A. tamarii*. The black arrows () indicate the common pathways to *A. ustus* and *A. tamarii*. The red arrows () indicate a pathway detected for *A. ustus* only. The blue arrows () indicate a pathway detected for *A. tamarii* only.