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D-MT Prompts the Anti-Tumor Effect of Oxaliplatin by Inhibiting IDO Expression in
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ABSTRACT

Colon cancer is one of the most common malignant tumors in the digestive system. Although oxaliplatin, a chemotherapy drug, has been clinically used to treat colon cancer, its therapeutic effect is unsatisfactory. It has been proved that indoleamine dioxygenase 2,3 (IDO), an immune checkpoint, is a result of tolerance to chemotherapy. Herein, an IDO inhibitor, D-MT (indoximod, 1-Methyl-D-tryptophan), was combined with oxaliplatin to treat colon cancer in mice. T cell infiltration in tumor tissues, the ratios of immune cells in the spleens, and the tumor growth and survival of the mice were detected and recorded. The results showed that the combination of oxaliplatin and D-MT significantly inhibited tumor growth and prolonged the survival of tumor-bearing mice. More importantly, the combination treatment increased the ratios of CD4+ T, CD8+ T and NK cells from the spleen in tumor-bearing mice, and prompted T cell infiltration in tumor tissues. This study provided a new therapeutic strategy for colon cancer treatment in the clinic, especially for patients with oxaliplatin resistance.

Keywords:

colon cancer · IDO · oxaliplatin · anti-tumor immune response

Introduction

Colon cancer is one of the most common malignant tumors of the gastrointestinal tract, with its incidence rising. Worldwide, colon cancer ranks the third in the incidence of male malignancies and fourth in mortality, and ranks the second in the incidence of female malignant tumors and third in mortality^{1,2}. In 2018, about 1.8 million colon cancer cases were diagnosed, with 881,000 patients dying of the disease. In the last decade, the incidence and mortality rates of colon cancer have been increasing in certain countries, such as China¹. Colon cancer is a major burden for patients worldwide. At present, the main treatment for colon cancer is surgery, with chemotherapy often performed following surgery for patients with metastasis. The response rate of chemotherapy could reach up to 50%, but drug resistance has been reported in nearly all patients with colon cancer³. Therefore, it is important to find a more effective treatment.

Oxaliplatin is a new anticancer platinum-based drug commonly used in the treatment of metastatic colorectal and liver cancer. Due to the heterogeneity of a tumor, a single drug often does not achieve satisfactory results. As reported, oxaliplatin treatment would lead to a reduction in macrophages and raised the expression of HMGB1 related to immunosuppression in the colon⁴. Therefore, studies have been trying to combine oxaliplatin with other therapies to treat tumors, improving the therapeutic effect of oxaliplatin⁵⁻⁷. Studies have shown that Huaier can effectively improve the anti-tumor effect of oxaliplatin by downregulating the expression of YAP protein which located in cell nucleus and related with cell proliferation, apoptosis and migration⁸. With the development of immunology, tumor immunotherapy has received more and more attention⁹. It has been found that anti-tumor immunity in colon cancer patients is inhibited to varying degrees, and thus enhancing the anti-tumor immune response may become an effective means of treating colon cancer. Consequently, chemical and immune combination treatment has become a new approach to cancer treatment¹⁰.

Studies have confirmed that the tumor microenvironment plays an important role in the tumor development stage^{11,12}. Tumor cells have the ability to evade host immune surveillance, which is one of the most important pathological features of tumor

formation and metastasis. For cancer patients, it is critical to reactivate and enhance the anti-tumor capabilities of their immune cells. The occurrence and development of tumors are highly correlated with the host immune response. Clinical studies have found that, although tumor-bearing patients produce potent anti-tumor immune responses, malignant tumors can continue to proliferate and metastasize. Tumor formation and development are associated with multiple factors, such as tumor immune escape, activation of inhibitory tumor immune response and failure of effective tumor immunity. There is a large number of immunosuppressive factors in the tumor microenvironment. These immunosuppressive factors play an important role in inhibiting T cells, such as indoleamine dioxygenase 2,3 (IDO)¹³⁻¹⁵.

IDO is a rate-limiting enzyme of the immunosuppressive tryptophan (Trp)/kynurenine (Kyn) metabolic pathway, including IDO1 and IDO2, which are highly expressed by tumor tissues, thereby, inhibiting T cell anti-tumor immunity. IDO has been found to be highly expressed in several types of human cancer^{16,17}, such as melanoma¹⁸⁻²⁰, and colon^{21,22}, brain²³ and ovarian cancer²⁴. In tumors such as acute myeloid leukemia²⁵⁻²⁷, the high expression of IDO inhibits anti-tumor immunity; therefore, IDO can serve as a new target for the treatment of tumors^{28,29}. A variety of IDO inhibitors have been approved for clinical trials by the US FDA. D-MT has been shown to be effective in inhibiting the expression of IDO³⁰. A previous study found that the inhibition of the IDO expression in melanoma-bearing mice enhances the anti-tumor effect of pimozone³¹, but it remains unclear whether IDO inhibition can promote the anti-tumor effect of oxaliplatin by increasing the anti-tumor immune response in tumor-bearing mice.

The present study explored the therapeutic effect of oxaliplatin combined with D-MT, an IDO inhibitor, on colon cancer, as well as the underlying mechanism. This study will provide a new method and experimental and theoretical basis for the clinical treatment of colon tumor.

Results

Combination treatment with D-MT + oxaliplatin significantly inhibits IDO and p-Stat3 expression in CT-26 cells

To detect the effectiveness of D-MT + oxaliplatin on the expression levels of IDO and p-Stat3 in colon cancer cells, the dosage of D-MT or oxaliplatin was 2.5 $\mu\text{g/ml}$ (the data not shown). As shown in Fig. 1, both of the IDO and p-Stat3 expression levels were significantly inhibited in the CT26 cells after treatment with D-MT for 24 or 48h, while those of expression levels were increased by treatment with oxaliplatin only. Interestingly, with the combination treatment of D-MT and oxaliplatin, the IDO and p-Stat3 expression levels in the cells were comparable with those of the D-MT group.

Combination treatment with D-MT + oxaliplatin inhibits tumor growth and prolongs the survival of CT-26 cell-bearing mice

In order to determine the therapeutic effect of D-MT + oxaliplatin on tumor-bearing mice, mouse survival and tumor weight were recorded. As shown in Fig 2a,b and d, 7 days after the last treatment, tumor growth was significantly inhibited in the oxaliplatin group, or D-MT + oxaliplatin group. Of note, the average tumor weight of mice treated with D-MT + oxaliplatin was the lowest. In addition, treatment with D-MT + oxaliplatin significantly prolonged the survival time of colon tumor-bearing mice, as compared with that of mice in other groups (Fig. 2c).

Combination treatment with D-MT + oxaliplatin promotes cell apoptosis in tumor tissues

Next, in order to determine whether treatment with D-MT + oxaliplatin damaged tumor cells, cell apoptosis was examined in tumor tissues by TUNEL, and the expression of cleaved caspase-3 by western blotting. First, the TUNEL results showed that the number of cells with positive staining significantly increased in D-MT or oxaliplatin-treated tumor cells, indicating that increased cell apoptosis. Of note, the combination treatment induced the greatest number of apoptotic cells in the tissues (Fig. 3a and b). In addition, the western blotting results revealed a similar trend. As

compared with other groups, combination treatment significantly increased the expression of cleaved caspase-3 (Fig. 3 c and d).

Combination treatment with D-MT + oxaliplatin influences the expression of tumor-related proteins

The results showed that compared with the PBS group, single-agent therapy with D-MT or oxaliplatin significantly inhibited the expression of MMP2 and combination treatment showed the similar trends (Fig. 4). In addition, compared with the PBS group, treatment with D-MT inhibited the expression of p-Stat3, but treatment with oxaliplatin prompted the expression of p-Stat3 significant (Fig. 4). Interestingly, treatment with oxaliplatin significantly increased the expression of IDO. Treatment with D-MT or D-MT+oxaliplatin showed a reduction of IDO expression (Fig. 4), indicating that combination treatment has the synergic action.

Combination treatment with D-MT + oxaliplatin significantly increases T cell infiltration in tumor tissues

It has been shown that the activation of IDO in the tumor microenvironment can impair the survival and function of T cells³². it was therefore detected herein that the T cells infiltrated tumor tissues by IF. The data showed that single-agent treatment with oxaliplatin or D-MT increased CD4⁺ and CD8⁺ T cell infiltration in tumor tissues (Fig. 5a-d). Of note, as compared with other groups, combination treatment with D-MT + oxaliplatin could significantly increase CD4⁺ and CD8⁺ T cell infiltration, indicating that combination treatment could significantly prompts the survival of T cells in tumor-bearing mice.

Combination treatment with D-MT + oxaliplatin significantly increases the ratio of immune cells in spleens from tumor-bearing mice

In order to determine whether combination treatment improved the whole anti-tumor effect of tumor-bearing mice, the ratio of immune cells in the spleen, the largest peripheral immune organ in the body, was detected. The results of flow cytometry

showed that the ratio of CD8⁺ T and NK cells were increased in the spleens of mice treated with D-MT (Fig. 6b-f). Although treatment with oxaliplatin did not significantly raised the ratio of immune cells (Fig. 6 a-f), but the ratio of CD4⁺ T, CD8⁺ T and NK cells were significantly raised in mice treated with D-MT and oxaliplatin, as compared with the D-MT or oxaliplatin group (Fig. 6 a-f).

Combination treatment with D-MT + oxaliplatin significantly increases the concentration of TNF- α or IFN- γ in the sera of tumor-bearing mice

Finally, the concentration of TNF- α or IFN- γ was detected to play an anti-tumor role. The ELISA results showed that, as compared with the PBS group, the concentration of TNF- α or IFN- γ in the sera of tumor-bearing mice in the D-MT, oxaliplatin and combination groups was raised. Of note, the concentration of TNF- α or IFN- γ in the combination group was higher than that in other groups (Fig. 7).

Discussion

Colon cancer is a malignant tumor that seriously endangers human health in several countries. Despite the existence of certain drugs for the treatment of colon cancer, their therapeutic effect is generally low. Colon cancer treatment has therefore been attracting more and more attention. In the present study, it was determined that the D-MT, an IDO inhibitor, prompted an anti-tumor effect of oxaliplatin and strengthened the immune response against tumor in mice.

Oxaliplatin, a chemotherapy drug, is the standard first-line treatment for colon cancer in the clinic^{33,34}. Unfortunately, clinical treatment often fails due to oxaliplatin resistance, which is associated with complex mechanisms, such as DNA adduct repair, cell death mechanisms and autophagy³⁵. In addition, it was found shown that immunosuppressive mechanisms, such as the immune checkpoint or immunosuppressive factors, were also important reasons for drug resistance³⁶. It has also been shown that oxaliplatin could effectively inhibit the growth of colon cancer, and even increase cell apoptosis, which would release a number of antigens in tumor tissues. It has been proven that chemotherapy could induce immunogenic cell death³⁷⁻³⁹, thereby increasing cancer immunogenicity by promoting dendritic cell maturation and T cell infiltration in tumor tissues⁴⁰. Furthermore, oxaliplatin could also increase T cell infiltration⁴¹, which would favour a good prognosis for tumor patients⁴². The present results showed that oxaliplatin significantly increased T cell infiltration and the ratio of T cells in the spleen, indicating that oxaliplatin could play an anti-tumor effect against colon cancer.

However, despite the several factors that affect the occurrence of tumors, the therapeutic effect of oxaliplatin was limited and unsatisfactory⁴³. One of the reasons is that the tumors that develop to cause immune suppression in the body, such as overexpression of the immune checkpoint³⁶. IDO has been proven to catalyze the oxidative catabolism of Trp to Kyn, and regulate immune responses by impairing the survival and activity of T cells^{44,45}. IDO has therefore become a therapeutic target for cancer that could either be used alone or in combination with other treatments for tumors^{46,47}. However, our results showed that oxaliplatin could inhibit tumor growth

and increase the apoptosis of tumor cells, but could also increase the expression of IDO, which might interfere with the therapeutic effect of oxaliplatin. We therefore detected the therapeutic effect of combination treatment with D-MT and oxaliplatin. D-MT has been proven to effectively inhibit the IDO expression⁴⁸ and enhance the anti-tumor immune response. D-MT might play a role in disrupting tumor immune escape^{30,49,50}. In fact, the present study determined that, even though oxaliplatin increased the IDO expression, the protein level of IDO could also be reduced in mice treated with D-MT and oxaliplatin. The reason might be that D-MT prompted the anti-tumor effect of oxaliplatin. In addition, the IDO expression could be upregulated by the activation of Stat3⁵¹. Stat3 activation would promote the development of colon tumor and related with the poor prognosis⁵². It was therefore found that D-MT also inhibited the expression of p-Stat3 in tumor cells. In addition, study showed that prevention of Stat3 activation could inhibition the expression of MMP2⁵³ which could promote the metastasis in colon tumor cells⁵⁴. Our results also confirmed that D-MT reduced the expression of MMP2 in CT26 cells or tumor-tissues. It might be related with that D-MT significantly inhibited the migration of CT26 cells.

Furthermore, the present results showed that the combination treatment with D-MT + oxaliplatin not only increased T cell infiltration in tumor tissues, but also raised the ratio of T cells in the spleen. They also indicated that this strategy could strengthen the anti-tumor immune response in tumor-bearing mice. T cells were found to play an important role in tumor immunity⁵⁵ and inhibit the expression of IDO, which could reverse CD8⁺ T cell suppression in breast cancer cell-bearing mice⁵⁶. Interfering with the activity of IDO could lead to strengthening the anti-tumor immune response by increasing the amount of CD8⁺ T lymphocyte infiltration⁵⁷. In addition, the present data showed that combination treatment with oxaliplatin and D-MT increased the ratio of NK cells, which was also an important reason against tumor. This might be associated with the inhibition of IDO, whose activation could lead to the downregulation of the function of NK cells^{58,59}.

In the present study, it was confirmed that D-MT could strengthen the anti-tumor effect of oxaliplatin, which might be associated with the inhibition of IDO.

Combination treatment with oxaliplatin and D-MT significantly inhibited tumor growth and prolonged the survival of tumor-bearing mice. Of note, the combination strategy could prompt the anti-tumor immune response by increasing T cell infiltration in tumors. This study might provide a new therapeutic strategy for colon cancer treatment in the clinic, particularly in patients treated with oxaliplatin, in which treatment was ineffective.

Methods

All animal studies were carried out in compliance with the ARRIVE guidelines (<https://arriveguidelines.org/>) as detailed below.

Cell lines, mice and drugs

A mouse colon cancer CT-26 cell line was purchased from American Type Culture Collection (ATCC; Rockville, MD, USA) and kept in the laboratory. IDO inhibitor D-MT was purchased from Merck KGaA. Oxaliplatin was purchased from Hengrui Pharmaceutical Co., Ltd. The Balb/c mice (Female, 6-8 weeks old) were procured from Beijing Vital River Laboratory Animal Technology Co., Ltd. The mice were fed in pathogen-free conditions, housed under a 12 h light/dark cycle at a temperature of 25±2°C. The animal study was approved by the Ethics Committee of Xinxiang Medical University.

Animal experiments

The CT-26 cells were adjusted to 1×10^7 cells/ml, and a 0.1-ml cell suspension was injected subcutaneously into the upper part of the right leg of mice to establish the colon cancer-bearing mouse model. Next, 24 mice were randomly divided into the PBS, D-MT, oxaliplatin and D-MT + oxaliplatin groups. On the 7th day of the establishment of the model, PBS, D-MT (1 mg/mouse), oxaliplatin (50 µg/mouse), D-MT (1 mg/mouse) and oxaliplatin (50 µg/mouse) were injected once a day for 7 days. A further 7 days after the last treatment, the tumors were separated and weighted. Finally, each tumor was divided into two parts, one was used to extract the protein for western blotting, and another was fixed in 4% formalin for immunofluorescence (IF) detection. The spleens were also separated from the mice for flow cytometry. In addition, 7 days after establishing the mouse model of colon cancer, another 40 mice were randomly divided into the PBS, D-MT, oxaliplatin and D-MT + oxaliplatin groups and administered the appropriate treatment. Mouse survival was observed and recorded every day. All the mice were given euthanasia by cervical dislocation after sera collection. All animal studies were performed according to protocols approved by the Ethics Committee of Xinxiang Medical University.

Western blotting

The protein of cells or tumor tissues was extracted in lysis buffer (Beyotime Institute of Biotechnology), and then the concentrations were quantified according to the instructions of the Bicinchoninic Acid Protein Assay kit (Beyotime Institute of Biotechnology). Based on the concentration, 30 µg protein was isolated by 10% SDS-polyacrylamide separation gel before being transferred onto PVDF membranes. The membranes were then incubated with the primary antibodies [Tubulin, IDO, p-signal transducer and activator of transcription 3 (p-Stat3), signal transducer and activator of transcription 3 (Stat3), matrix metalloproteinase 2 (MMP2), cleaved caspase-3 or I κ B α ; Cell Signaling Technology, Inc.], diluted according to the manufacturer's instructions. After 2 h, the membranes were washed with 1xTBST and incubated with the secondary antibodies for 1 h. Finally, the specificity of antigen-antibody complexes was detected using enhanced chemiluminescence reagent (Cell Signaling Technology, Inc.) and the images were visualized by Fusion FX Spectra imaging system.

Immunofluorescence (IF) detection

Indirect IF was performed as previously described⁶⁰. Briefly, the tumor slides were incubated with the appropriate antibody (CD3, CD4 and CD8) at 4°C overnight in a wet box. The tumor slides were restored to 25°C at least 30 min after being incubated with the secondary antibody for 30 min. Finally, the slides were stained with DAPI and the images were captured using a fluorescence or a laser confocal microscope. The intensities of positive cells were analyzed using a scale of 0-3+: 0, no staining identified; 1+, <25% positive cells; 2+, 25-75% positive cells; 3+, >75% positive cells.

Terminal deoxynucleotidyl transferase dUTP nick end labelling (TUNEL)

The cell apoptosis in tumor tissues was detected by TUNEL assay kit (Beyotime Institute of Biotechnology), according to the manufacturer's instructions. Briefly, the TUNEL detection solution was dropwise added onto the surface of tumor section.

Following incubation in the dark for 60 min at 37°C, the sections were washed with PBS for 10 min, 3 times. Finally, the sections were dried and sealed with anti-fluorescence quenching solution, and the positive cells were observed using the fluorescence microscope. In addition, Hoechst 33342 solution (Beyotime Institute of Biotechnology) was used for staining living cells, according to the manufacturer's instructions.

Flow cytometry

The concentration of spleen cells was adjusted to 1×10^7 cells/ml, and each tube was added into 100 μ l cell suspension. The cells were incubated with antibodies for CD3, CD4, CD8 and CD49b (BioLegend, Inc.) at 4°C for 30 min. The ratio of cells was detected using flow cytometry (CytoFLEX; Beckman Coulter, Inc.).

Enzyme-linked immunosorbent assay (ELISA).

One week after the last treatment, the mice in each group were sacrificed and the sera collected. The concentration of tumor necrosis factor alpha (TNF- α) or interferon gamma (IFN- γ) was analyzed by ELISA kits (RayBiotech), according to the manufacturer's instructions.

Statistical analysis

Measurement data are expressed as the mean \pm SD of three independent experiments. Data were analyzed by SPSS 19.0 (IBM Corp.). One-way ANOVA was performed to test the difference among the different groups, and the Kaplan-Meier method with a log-rank test was used to analyze survival. $P < 0.05$ were considered to indicate a statistically significant difference.

References

- 1 Bray, F. *et al.* Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA: a cancer journal for clinicians* **68**, 394-424, doi:10.3322/caac.21492 (2018).
- 2 Kumar, R. & Lewis, C. R. in *StatPearls* (2021).
- 3 Hu, T., Li, Z., Gao, C. Y. & Cho, C. H. Mechanisms of drug resistance in colon cancer and its therapeutic strategies. *World journal of gastroenterology* **22**, 6876-6889, doi:10.3748/wjg.v22.i30.6876 (2016).
- 4 Stojanovska, V. *et al.* Oxaliplatin-induced changes in microbiota, TLR4+ cells and enhanced HMGB1 expression in the murine colon. *PLoS One* **13**, e0198359, doi:10.1371/journal.pone.0198359 (2018).
- 5 Yin, X. *et al.* ID1 promotes hepatocellular carcinoma proliferation and confers chemoresistance to oxaliplatin by activating pentose phosphate pathway. *Journal of experimental & clinical cancer research : CR* **36**, 166, doi:10.1186/s13046-017-0637-7 (2017).
- 6 Wang, X. *et al.* Secretory Clusterin Mediates Oxaliplatin Resistance via the Gadd45a/PI3K/Akt Signaling Pathway in Hepatocellular Carcinoma. *Journal of Cancer* **9**, 1403-1413, doi:10.7150/jca.23849 (2018).
- 7 Ren, W. W. *et al.* MicroRNA-125b reverses oxaliplatin resistance in hepatocellular carcinoma by negatively regulating EVA1A mediated autophagy. *Cell death & disease* **9**, 547, doi:10.1038/s41419-018-0592-z (2018).
- 8 Tao, Y. *et al.* Huaier Augmented the Chemotherapeutic Sensitivity of Oxaliplatin via Downregulation of YAP in Hepatocellular Carcinoma. *Journal of Cancer* **9**, 3962-3970, doi:10.7150/jca.25909 (2018).
- 9 Tsai, H. F. & Hsu, P. N. Cancer immunotherapy by targeting immune checkpoints: mechanism of T cell dysfunction in cancer immunity and new therapeutic targets. *Journal of biomedical science* **24**, 35, doi:10.1186/s12929-017-0341-0 (2017).
- 10 Mahoney, K. M., Rennert, P. D. & Freeman, G. J. Combination cancer immunotherapy and new immunomodulatory targets. *Nature reviews. Drug discovery* **14**, 561-584, doi:10.1038/nrd4591 (2015).

- 11 Witz, I. P. The tumor microenvironment: the making of a paradigm. *Cancer microenvironment : official journal of the International Cancer Microenvironment Society* **2 Suppl 1**, 9-17, doi:10.1007/s12307-009-0025-8 (2009).
- 12 Arneth, B. Tumor Microenvironment. *Medicina (Kaunas)* **56**, doi:10.3390/medicina56010015 (2019).
- 13 Swanson, K. A., Zheng, Y., Heidler, K. M., Mizobuchi, T. & Wilkes, D. S. CD11c+ cells modulate pulmonary immune responses by production of indoleamine 2,3-dioxygenase. *American journal of respiratory cell and molecular biology* **30**, 311-318, doi:10.1165/rcmb.2003-0268OC (2004).
- 14 Hornyak, L. *et al.* The Role of Indoleamine-2,3-Dioxygenase in Cancer Development, Diagnostics, and Therapy. *Frontiers in immunology* **9**, 151, doi:10.3389/fimmu.2018.00151 (2018).
- 15 Ye, Q. *et al.* Expression of programmed cell death protein 1 (PD-1) and indoleamine 2,3-dioxygenase (IDO) in the tumor microenvironment and in tumor-draining lymph nodes of breast cancer. *Human pathology* **75**, 81-90, doi:10.1016/j.humpath.2018.02.004 (2018).
- 16 Mitsuka, K. *et al.* Expression of indoleamine 2,3-dioxygenase and correlation with pathological malignancy in gliomas. *Neurosurgery* **72**, 1031-1038; discussion 1038-1039, doi:10.1227/NEU.0b013e31828cf945 (2013).
- 17 Ye, J. *et al.* Tumoral indoleamine 2,3-dioxygenase expression predicts poor outcome in laryngeal squamous cell carcinoma. *Virchows Archiv : an international journal of pathology* **462**, 73-81, doi:10.1007/s00428-012-1340-x (2013).
- 18 Munn, D. H. *et al.* Expression of indoleamine 2,3-dioxygenase by plasmacytoid dendritic cells in tumor-draining lymph nodes. *The Journal of clinical investigation* **114**, 280-290, doi:10.1172/JCI21583 (2004).
- 19 Gerlini, G. *et al.* Indoleamine 2,3-dioxygenase+ cells correspond to the BDCA2+ plasmacytoid dendritic cells in human melanoma sentinel nodes. *The Journal of investigative dermatology* **130**, 898-901, doi:10.1038/jid.2009.307 (2010).

- 20 Speeckaert, R. *et al.* Indoleamine 2,3-dioxygenase, a new prognostic marker in sentinel lymph nodes of melanoma patients. *European journal of cancer* **48**, 2004-2011, doi:10.1016/j.ejca.2011.09.007 (2012).
- 21 Ferdinande, L. *et al.* Clinicopathological significance of indoleamine 2,3-dioxygenase 1 expression in colorectal cancer. *British journal of cancer* **106**, 141-147, doi:10.1038/bjc.2011.513 (2012).
- 22 Brandacher, G. *et al.* Prognostic value of indoleamine 2,3-dioxygenase expression in colorectal cancer: effect on tumor-infiltrating T cells. *Clinical cancer research : an official journal of the American Association for Cancer Research* **12**, 1144-1151, doi:10.1158/1078-0432.CCR-05-1966 (2006).
- 23 Zhai, L. *et al.* The role of IDO in brain tumor immunotherapy. *Journal of neuro-oncology* **123**, 395-403, doi:10.1007/s11060-014-1687-8 (2015).
- 24 Okamoto, A. *et al.* Indoleamine 2,3-dioxygenase serves as a marker of poor prognosis in gene expression profiles of serous ovarian cancer cells. *Clinical cancer research : an official journal of the American Association for Cancer Research* **11**, 6030-6039, doi:10.1158/1078-0432.CCR-04-2671 (2005).
- 25 Chamuleau, M. E. *et al.* High INDO (indoleamine 2,3-dioxygenase) mRNA level in blasts of acute myeloid leukemic patients predicts poor clinical outcome. *Haematologica* **93**, 1894-1898, doi:10.3324/haematol.13113 (2008).
- 26 Folgiero, V. *et al.* Indoleamine 2,3-dioxygenase 1 (IDO1) activity in leukemia blasts correlates with poor outcome in childhood acute myeloid leukemia. *Oncotarget* **5**, 2052-2064, doi:10.18632/oncotarget.1504 (2014).
- 27 Sobash, P. T. *et al.* Role of indoleamine 2,3-dioxygenase in acute myeloid leukemia. *Future Oncol* **16**, 3085-3094, doi:10.2217/fon-2019-0642 (2020).
- 28 Prendergast, G. C., Malachowski, W. J., Mondal, A., Scherle, P. & Muller, A. J. Indoleamine 2,3-Dioxygenase and Its Therapeutic Inhibition in Cancer. *International review of cell and molecular biology* **336**, 175-203, doi:10.1016/bs.ircmb.2017.07.004 (2018).
- 29 Monjazebe, A. M. *et al.* Blocking Indoleamine-2,3-Dioxygenase Rebound Immune Suppression Boosts Antitumor Effects of Radio-Immunotherapy in Murine

- Models and Spontaneous Canine Malignancies. *Clinical cancer research : an official journal of the American Association for Cancer Research* **22**, 4328-4340, doi:10.1158/1078-0432.CCR-15-3026 (2016).
- 30 Schmidt, S. K. *et al.* Influence of tryptophan contained in 1-Methyl-Tryptophan on antimicrobial and immunoregulatory functions of indoleamine 2,3-dioxygenase. *PloS one* **7**, e44797, doi:10.1371/journal.pone.0044797 (2012).
- 31 Jia, H. *et al.* The enhanced antitumour response of pimozide combined with the IDO inhibitor LMT in melanoma. *International journal of oncology* **53**, 949-960, doi:10.3892/ijo.2018.4473 (2018).
- 32 Liu, D. *et al.* Redox-Activated Porphyrin-Based Liposome Remote-Loaded with Indoleamine 2,3-Dioxygenase (IDO) Inhibitor for Synergistic Photoimmunotherapy through Induction of Immunogenic Cell Death and Blockage of IDO Pathway. *Nano letters* **19**, 6964-6976, doi:10.1021/acs.nanolett.9b02306 (2019).
- 33 Schmoll, H. J. *et al.* Effect of adjuvant capecitabine or fluorouracil, with or without oxaliplatin, on survival outcomes in stage III colon cancer and the effect of oxaliplatin on post-relapse survival: a pooled analysis of individual patient data from four randomised controlled trials. *The Lancet. Oncology* **15**, 1481-1492, doi:10.1016/S1470-2045(14)70486-3 (2014).
- 34 Shah, M. A. *et al.* Impact of Patient Factors on Recurrence Risk and Time Dependency of Oxaliplatin Benefit in Patients With Colon Cancer: Analysis From Modern-Era Adjuvant Studies in the Adjuvant Colon Cancer End Points (ACCENT) Database. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* **34**, 843-853, doi:10.1200/JCO.2015.63.0558 (2016).
- 35 Martinez-Balibrea, E. *et al.* Tumor-Related Molecular Mechanisms of Oxaliplatin Resistance. *Molecular cancer therapeutics* **14**, 1767-1776, doi:10.1158/1535-7163.MCT-14-0636 (2015).
- 36 Peng, J. *et al.* Chemotherapy Induces Programmed Cell Death-Ligand 1 Overexpression via the Nuclear Factor-kappaB to Foster an Immunosuppressive

- Tumor Microenvironment in Ovarian Cancer. *Cancer research* **75**, 5034-5045, doi:10.1158/0008-5472.CAN-14-3098 (2015).
- 37 Casares, N. *et al.* Caspase-dependent immunogenicity of doxorubicin-induced tumor cell death. *The Journal of experimental medicine* **202**, 1691-1701, doi:10.1084/jem.20050915 (2005).
- 38 Tesniere, A. *et al.* Immunogenic death of colon cancer cells treated with oxaliplatin. *Oncogene* **29**, 482-491, doi:10.1038/onc.2009.356 (2010).
- 39 Sun, F. *et al.* Oxaliplatin induces immunogenic cells death and enhances therapeutic efficacy of checkpoint inhibitor in a model of murine lung carcinoma. *J Recept Signal Transduct Res* **39**, 208-214, doi:10.1080/10799893.2019.1655050 (2019).
- 40 Kroemer, G., Galluzzi, L., Kepp, O. & Zitvogel, L. Immunogenic cell death in cancer therapy. *Annual review of immunology* **31**, 51-72, doi:10.1146/annurev-immunol-032712-100008 (2013).
- 41 Galaine, J. *et al.* CD4 T cells target colorectal cancer antigens upregulated by oxaliplatin. *International journal of cancer* **145**, 3112-3125, doi:10.1002/ijc.32620 (2019).
- 42 Fridman, W. H., Pages, F., Sautes-Fridman, C. & Galon, J. The immune contexture in human tumours: impact on clinical outcome. *Nature reviews. Cancer* **12**, 298-306, doi:10.1038/nrc3245 (2012).
- 43 Healey, E. *et al.* Comparative effectiveness of 5-fluorouracil with and without oxaliplatin in the treatment of colorectal cancer in clinical practice. *Anticancer research* **33**, 1053-1060 (2013).
- 44 He, C. *et al.* Core-shell nanoscale coordination polymers combine chemotherapy and photodynamic therapy to potentiate checkpoint blockade cancer immunotherapy. *Nature communications* **7**, 12499, doi:10.1038/ncomms12499 (2016).
- 45 Zou, W. Immunosuppressive networks in the tumour environment and their therapeutic relevance. *Nature reviews. Cancer* **5**, 263-274, doi:10.1038/nrc1586 (2005).

- 46 Zhang, Y. *et al.* Tumor-Targeted Gene Silencing IDO Synergizes PTT-Induced Apoptosis and Enhances Anti-tumor Immunity. *Front Immunol* **11**, 968, doi:10.3389/fimmu.2020.00968 (2020).
- 47 Selvan, S. R., Dowling, J. P., Kelly, W. K. & Lin, J. Indoleamine 2,3-dioxygenase (IDO): Biology and Target in Cancer Immunotherapies. *Current cancer drug targets* **16**, 755-764 (2016).
- 48 Cepcova, D. *et al.* The protective effect of 1-methyltryptophan isomers in renal ischemia-reperfusion injury is not exclusively dependent on indolamine 2,3-dioxygenase inhibition. *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie* **135**, 111180, doi:10.1016/j.biopha.2020.111180 (2021).
- 49 Moreno, A. C. *et al.* The expanding roles of 1-methyl-tryptophan (1-MT): in addition to inhibiting kynurenine production, 1-MT activates the synthesis of melatonin in skin cells. *The FEBS journal* **280**, 4782-4792, doi:10.1111/febs.12444 (2013).
- 50 Gutzmer, R. *et al.* Atezolizumab, vemurafenib, and cobimetinib as first-line treatment for unresectable advanced BRAF(V600) mutation-positive melanoma (IMspire150): primary analysis of the randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet* **395**, 1835-1844, doi:10.1016/S0140-6736(20)30934-X (2020).
- 51 Zhang, Y. *et al.* Prolonged skin grafts survival time by IFN-gamma in allogeneic skin transplantation model during acute rejection through IFN-gamma/STAT3/IDO pathway in epidermal layer. *Biochemical and biophysical research communications* **496**, 436-442, doi:10.1016/j.bbrc.2017.12.152 (2018).
- 52 Heichler, C. *et al.* STAT3 activation through IL-6/IL-11 in cancer-associated fibroblasts promotes colorectal tumour development and correlates with poor prognosis. *Gut* **69**, 1269-1282, doi:10.1136/gutjnl-2019-319200 (2020).
- 53 Liang, Y. *et al.* Elevated IL-33 promotes expression of MMP2 and MMP9 via activating STAT3 in alveolar macrophages during LPS-induced acute lung injury. *Cell Mol Biol Lett* **23**, 52, doi:10.1186/s11658-018-0117-x (2018).

- 54 Li, H. *et al.* AKT2 phosphorylation of hexokinase 2 at T473 promotes tumorigenesis and metastasis in colon cancer cells via NF-kappaB, HIF1 alpha, MMP2, and MMP9 upregulation. *Cell Signal* **58**, 99-110, doi:10.1016/j.cellsig.2019.03.011 (2019).
- 55 Iwahori, K. Cytotoxic CD8(+) Lymphocytes in the Tumor Microenvironment. *Advances in experimental medicine and biology* **1224**, 53-62, doi:10.1007/978-3-030-35723-8_4 (2020).
- 56 Gao, J., Deng, F. & Jia, W. Inhibition of Indoleamine 2,3-Dioxygenase Enhances the Therapeutic Efficacy of Immunogenic Chemotherapeutics in Breast Cancer. *Journal of breast cancer* **22**, 196-209, doi:10.4048/jbc.2019.22.e23 (2019).
- 57 Huang, Z. *et al.* Enhanced cancer therapy through synergetic photodynamic/immune checkpoint blockade mediated by a liposomal conjugate comprised of porphyrin and IDO inhibitor. *Theranostics* **9**, 5542-5557, doi:10.7150/thno.35343 (2019).
- 58 De Martino, M. *et al.* Blockade of Stat3 oncogene addiction induces cellular senescence and reveals a cell-nonautonomous activity suitable for cancer immunotherapy. *Oncoimmunology* **9**, 1715767, doi:10.1080/2162402X.2020.1715767 (2020).
- 59 Ban, Y. *et al.* Effect of Indoleamine 2,3-Dioxygenase Expressed in HTR-8/SVneo Cells on Decidual NK Cell Cytotoxicity. *American journal of reproductive immunology* **75**, 519-528, doi:10.1111/aji.12481 (2016).
- 60 Ji, X. Y. *et al.* Detection of RAGE expression and its application to diabetic wound age estimation. *International journal of legal medicine* **131**, 691-698, doi:10.1007/s00414-016-1529-7 (2017).

Acknowledgements

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Conflict of interest

The authors declare that they have no conflict of interest.

Authors' contributions

All authors have read and approved the manuscript. TSZ and CXD designed the experiments. YXZ and YL participated in designing the experiment, analysing the data and revising the manuscript. YXZ, YL, ZAL, YJ, JG, WWR, RPL, GZZ, JL, MJL, XYL and SGW carried out the experiments. YXZ wrote the manuscript.

Figure legends

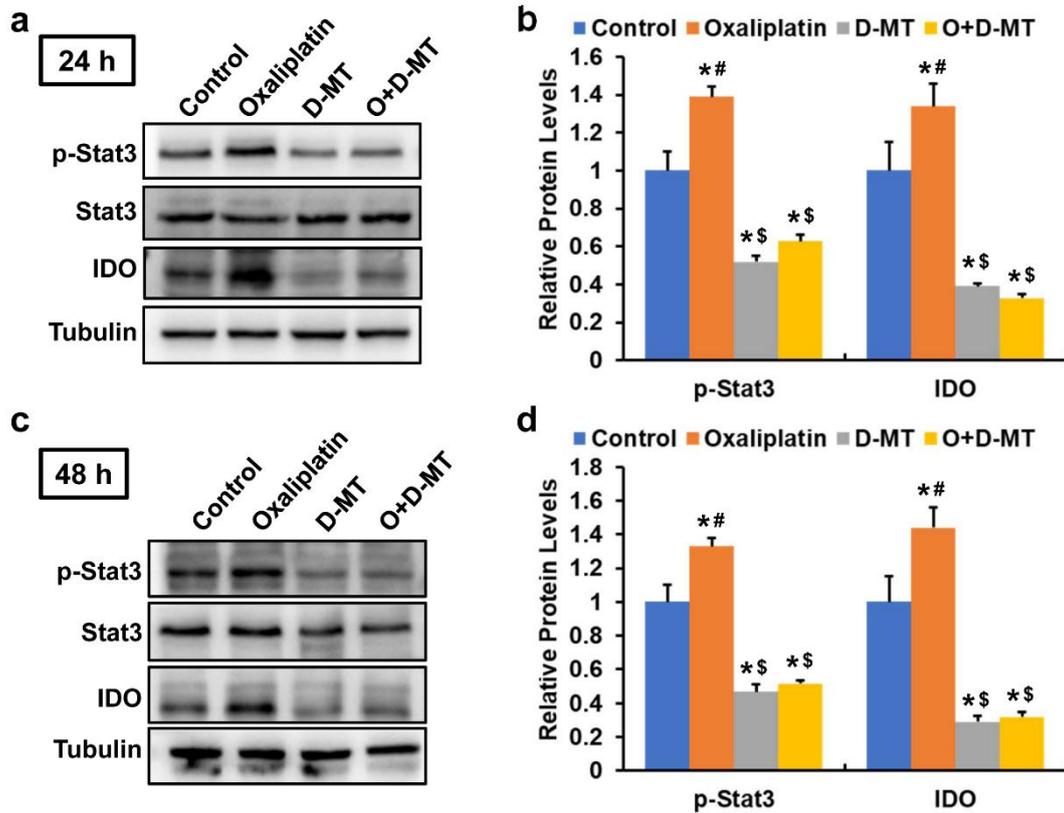


Figure 1. Effect of D-MT + oxaliplatin on the relative protein expression *in vitro*. CT-26 cells were seeded onto 6-well plates and treated with D-MT and oxaliplatin. The protein was extracted in lysis buffer 24 or 48 h after drug treatment, and the relative protein expression was detected by western blotting. **a** The protein expression of p-Stat3 and IDO was detected by western blotting 24 h after drug treatment. **b** Quantification of Fig. 1a. **c** The protein expression of p-Stat3 and IDO was detected by western blotting 48 h after drug treatment. **d** Quantification of the Fig. 1c. Data are presented as the mean \pm SD (n=3). * P <0.05 vs. the Control group; # P <0.05 vs. the D-MT group; $^{\$}$ P <0.05 vs. the Oxaliplatin group. IDO, indoleamine dioxygenase 2,3; Stat3, signal transducer and activator of transcription 3.

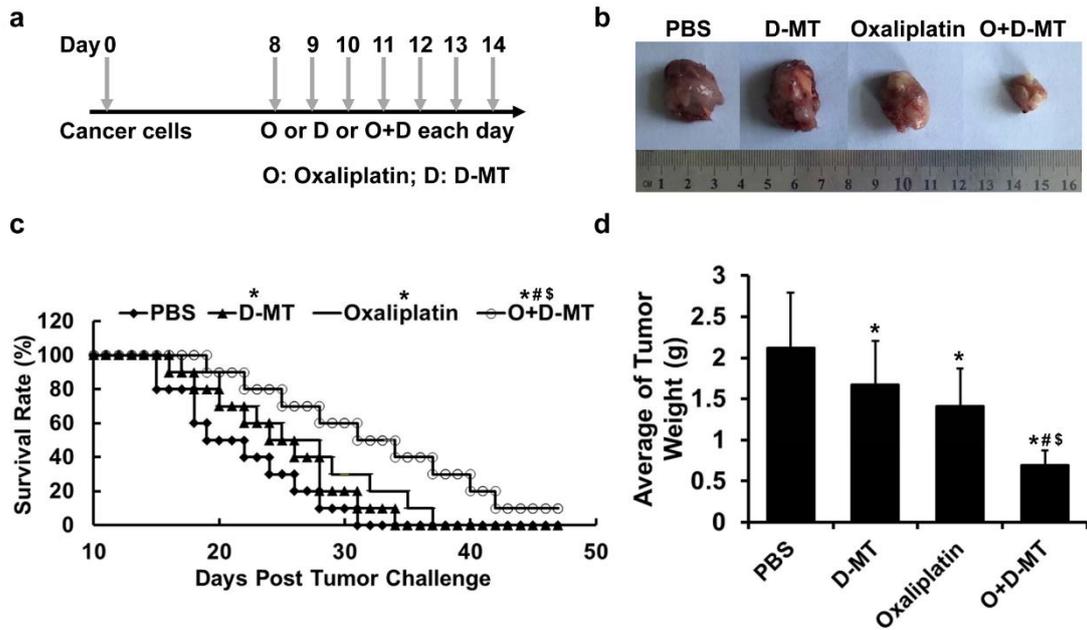


Figure 2. Effect of treatment with D-MT + oxaliplatin on tumor growth *in vivo*. The mice were s.c. inoculated with 1×10^6 CT-26 cells and treated with PBS, D-MT, oxaliplatin, D-MT and oxaliplatin. Survival and tumor weight were then recorded. **a** Schedule of combination treatment. **b** Images of the representative tumor in each group (n=10). **c** Survival rate of CT-26 cell-bearing mice. **d** Tumor weight analysis (n=6). * $P < 0.05$ vs. the Control group; # $P < 0.05$ vs. the D-MT group; \$ $P < 0.05$ vs. the Oxaliplatin group.

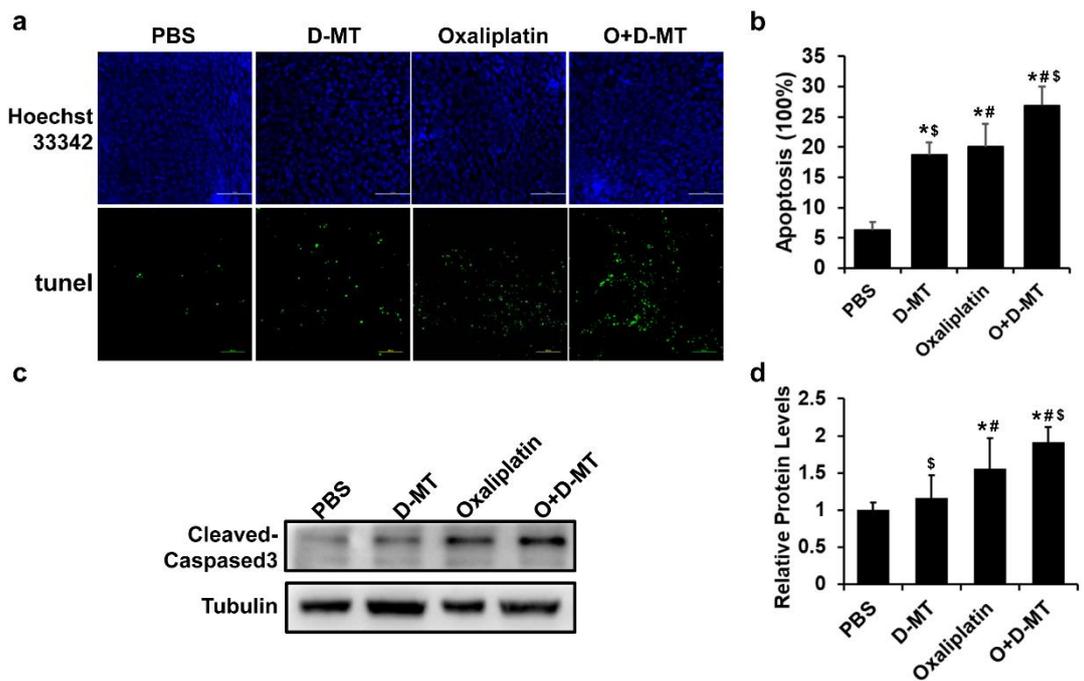


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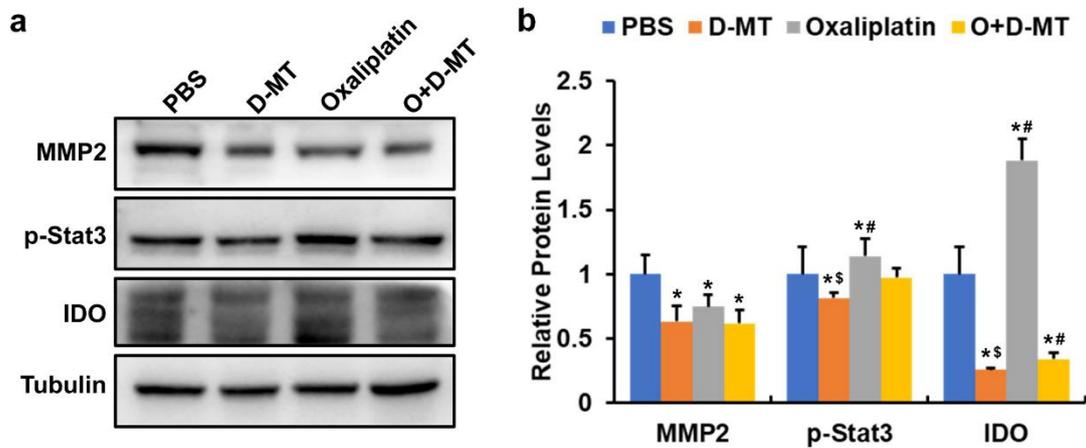


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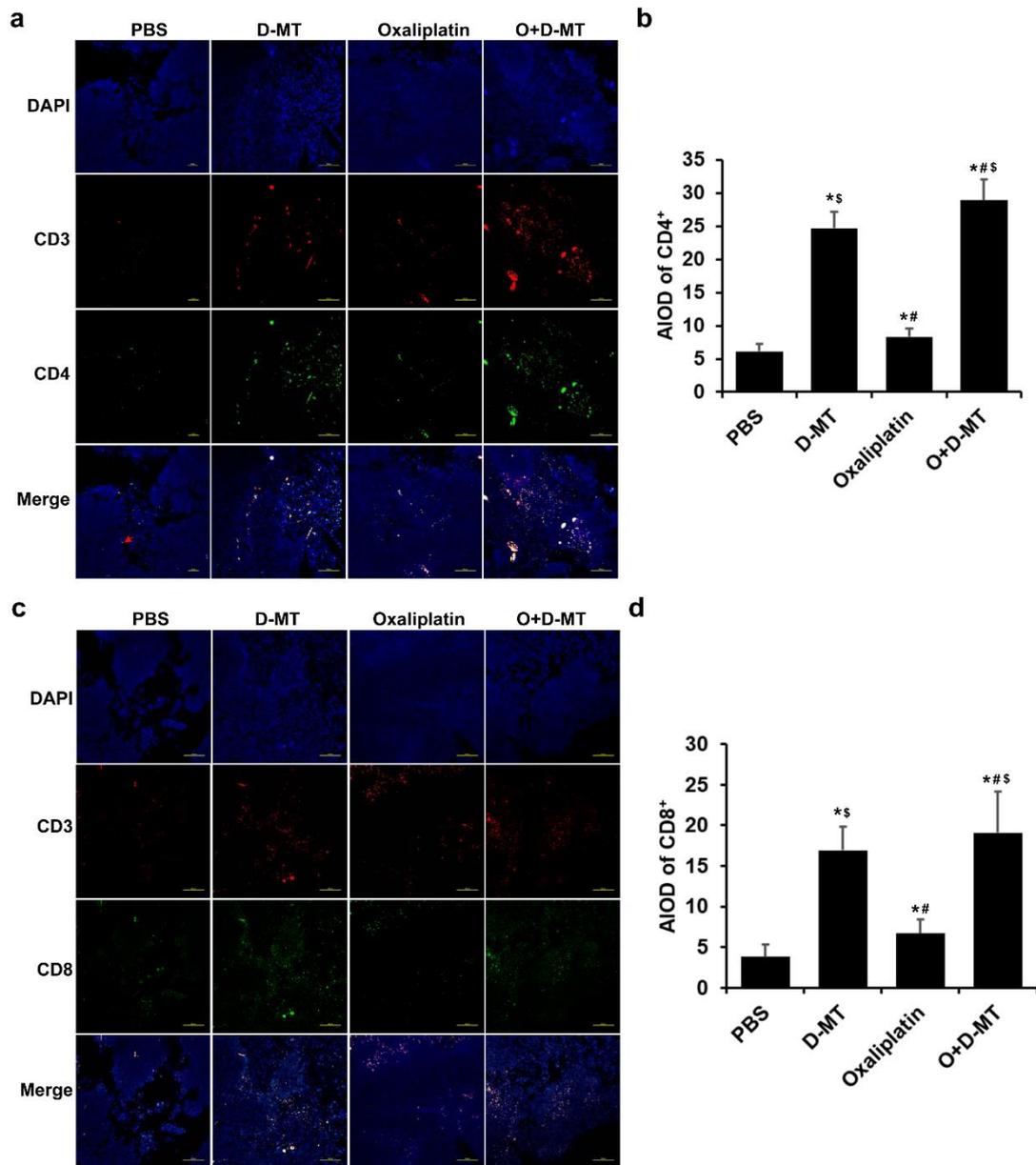


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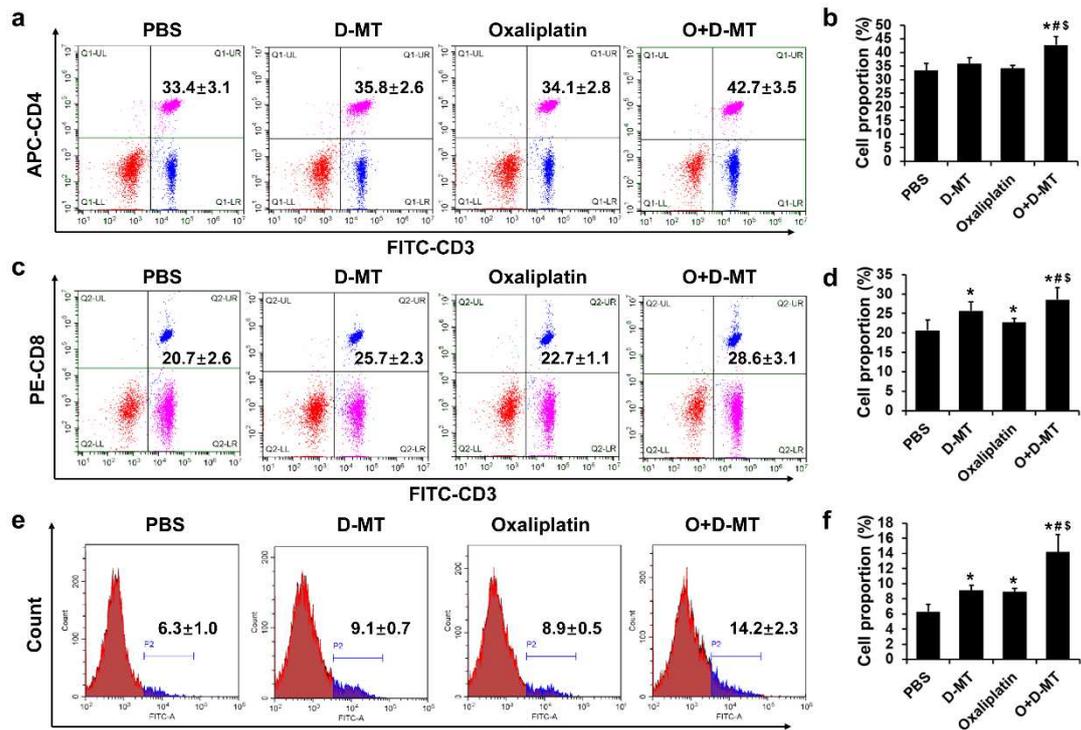


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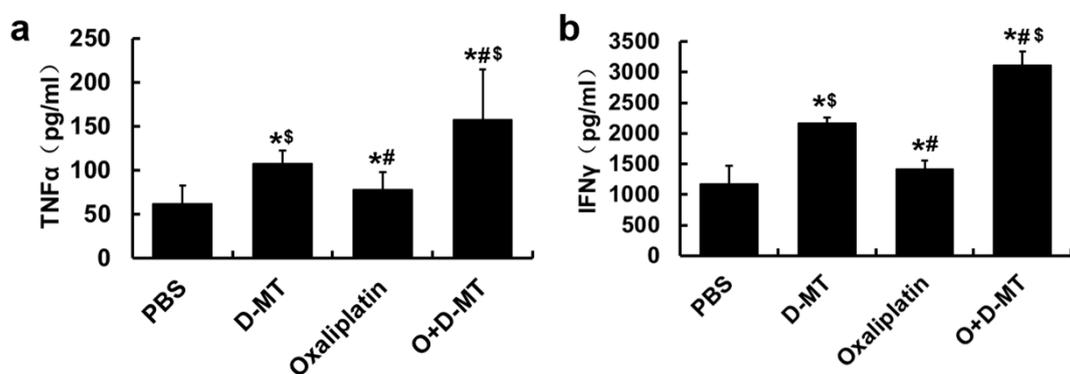


Figure 7. Effect of treatment with D-MT + oxaliplatin on the concentration of cytokines.

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Figures

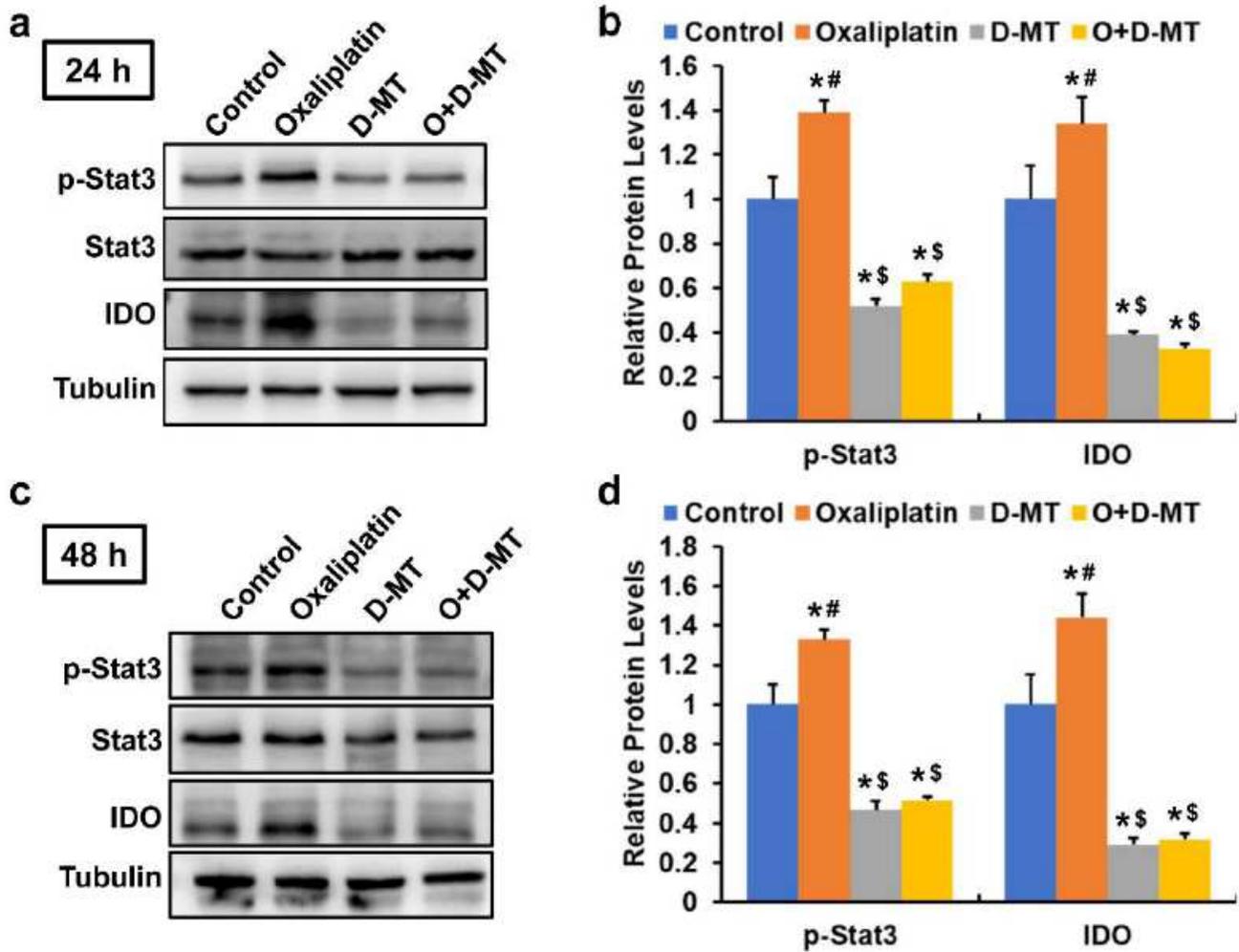


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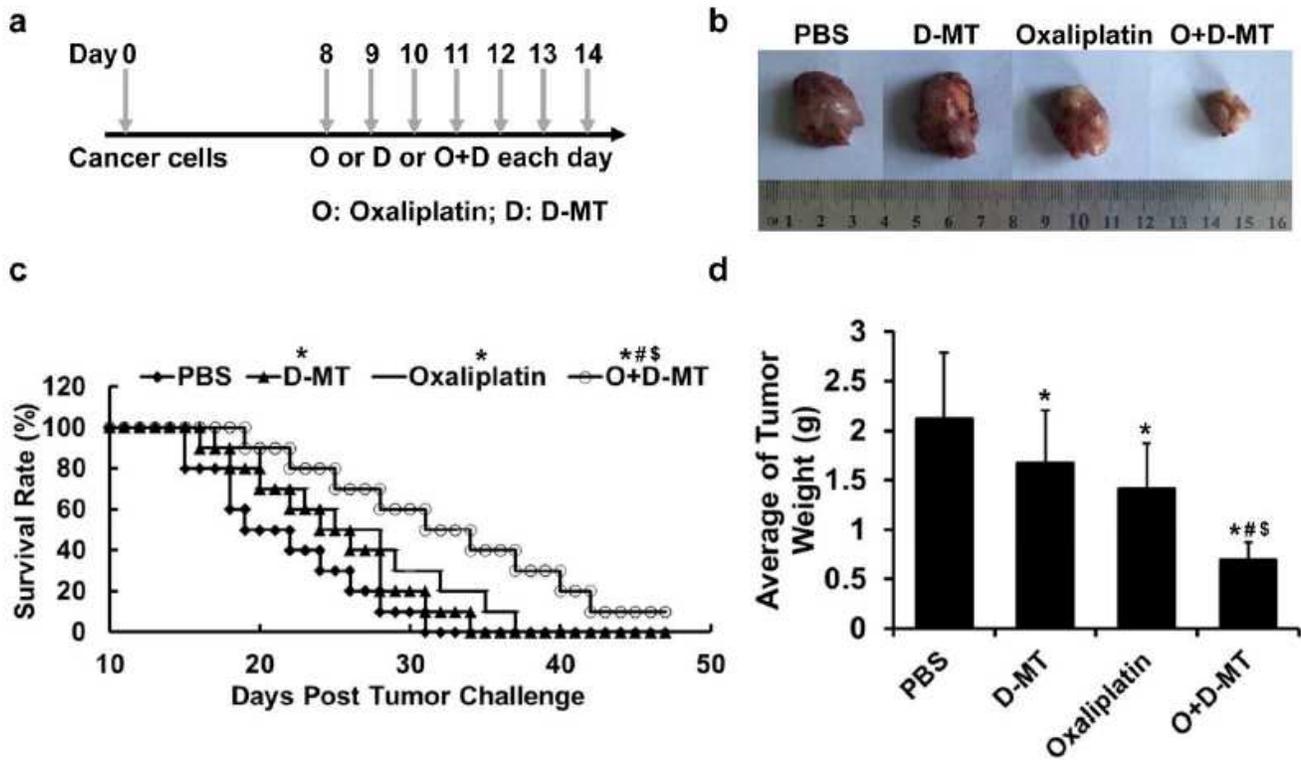


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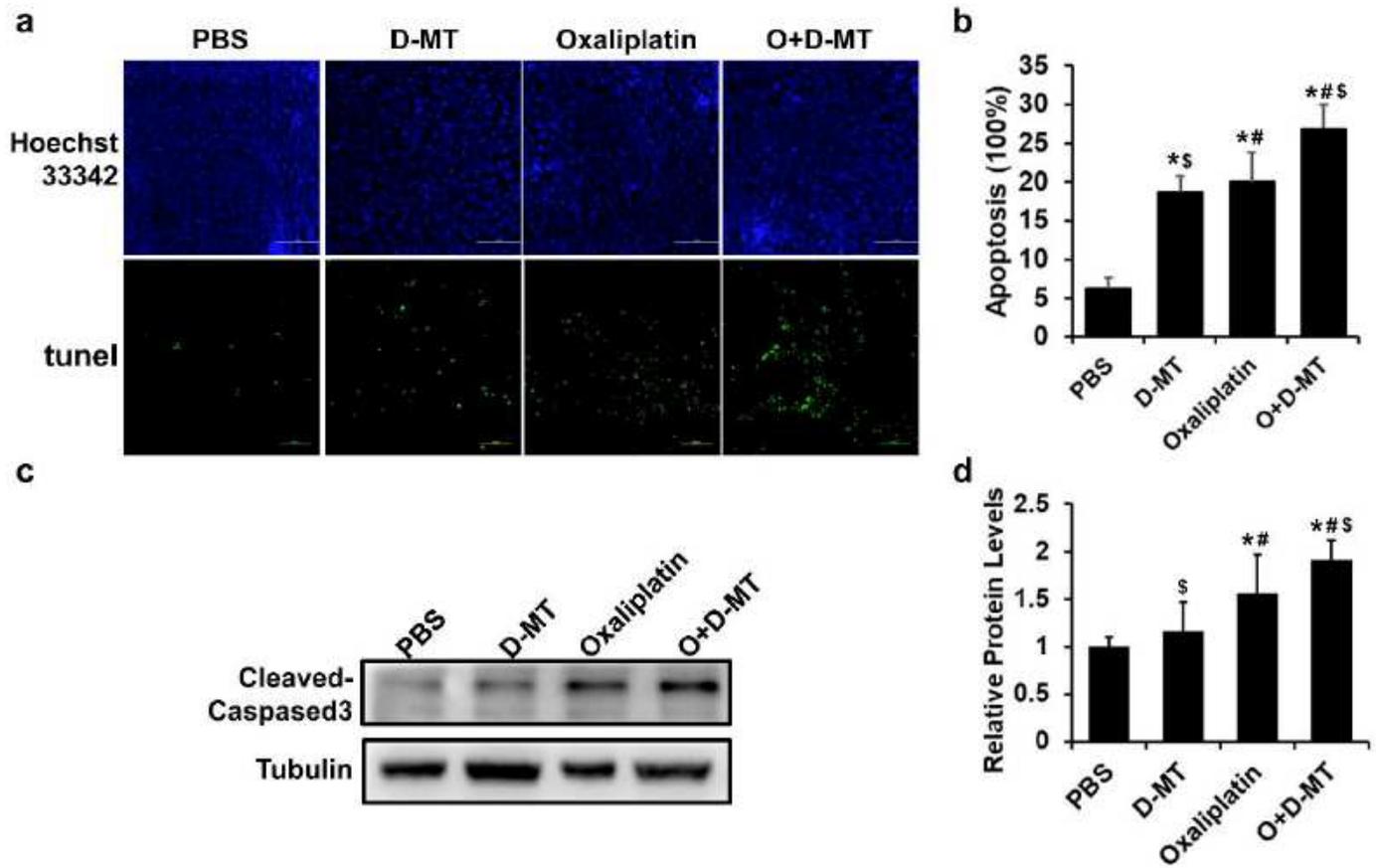


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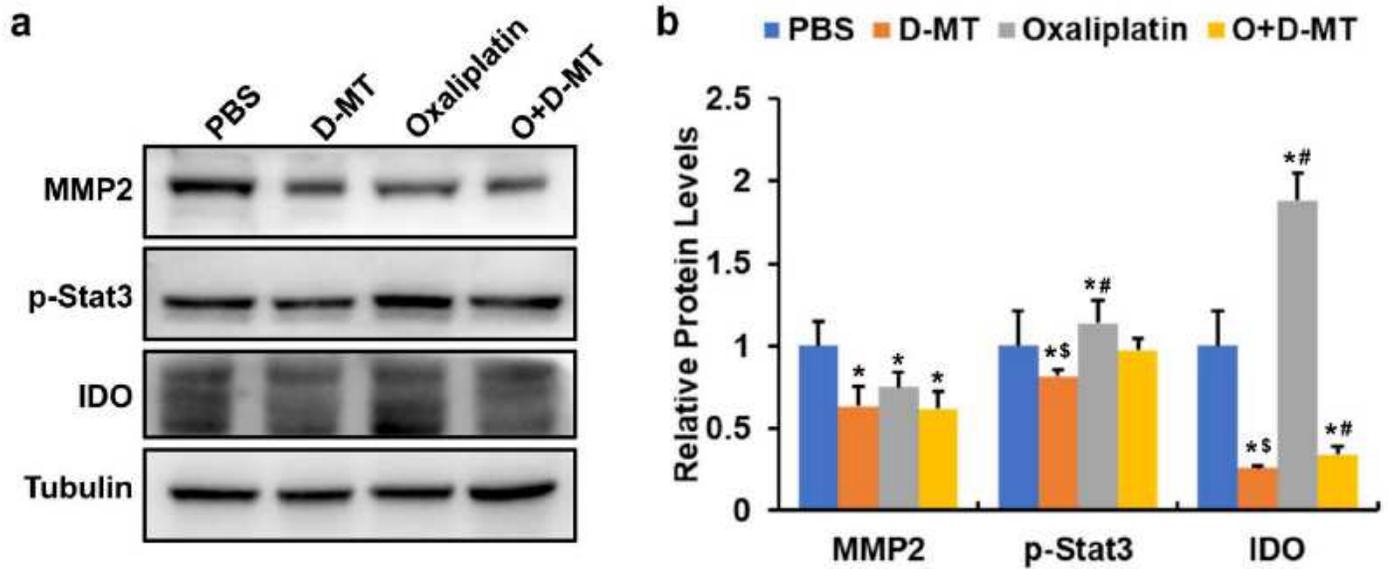


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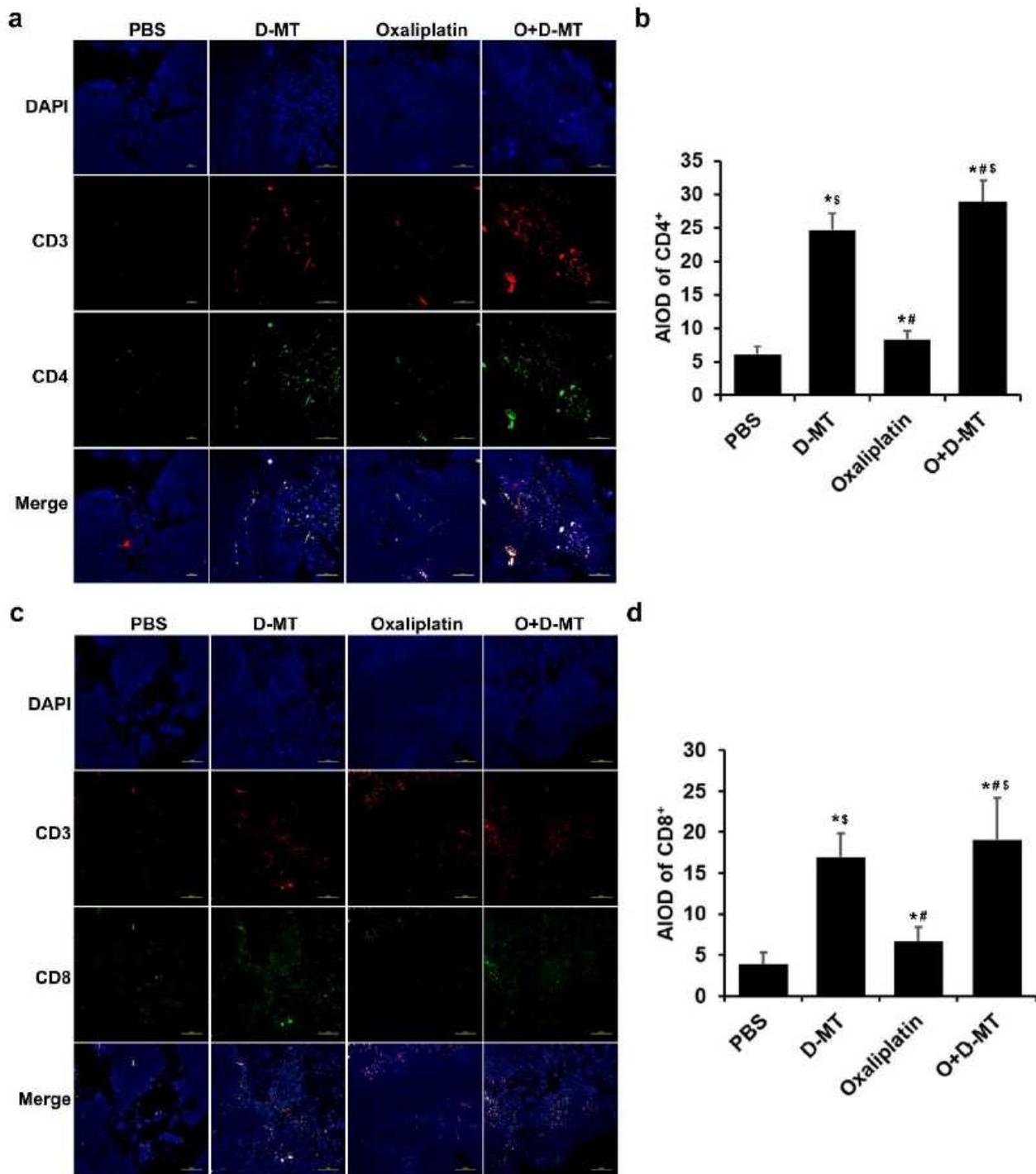


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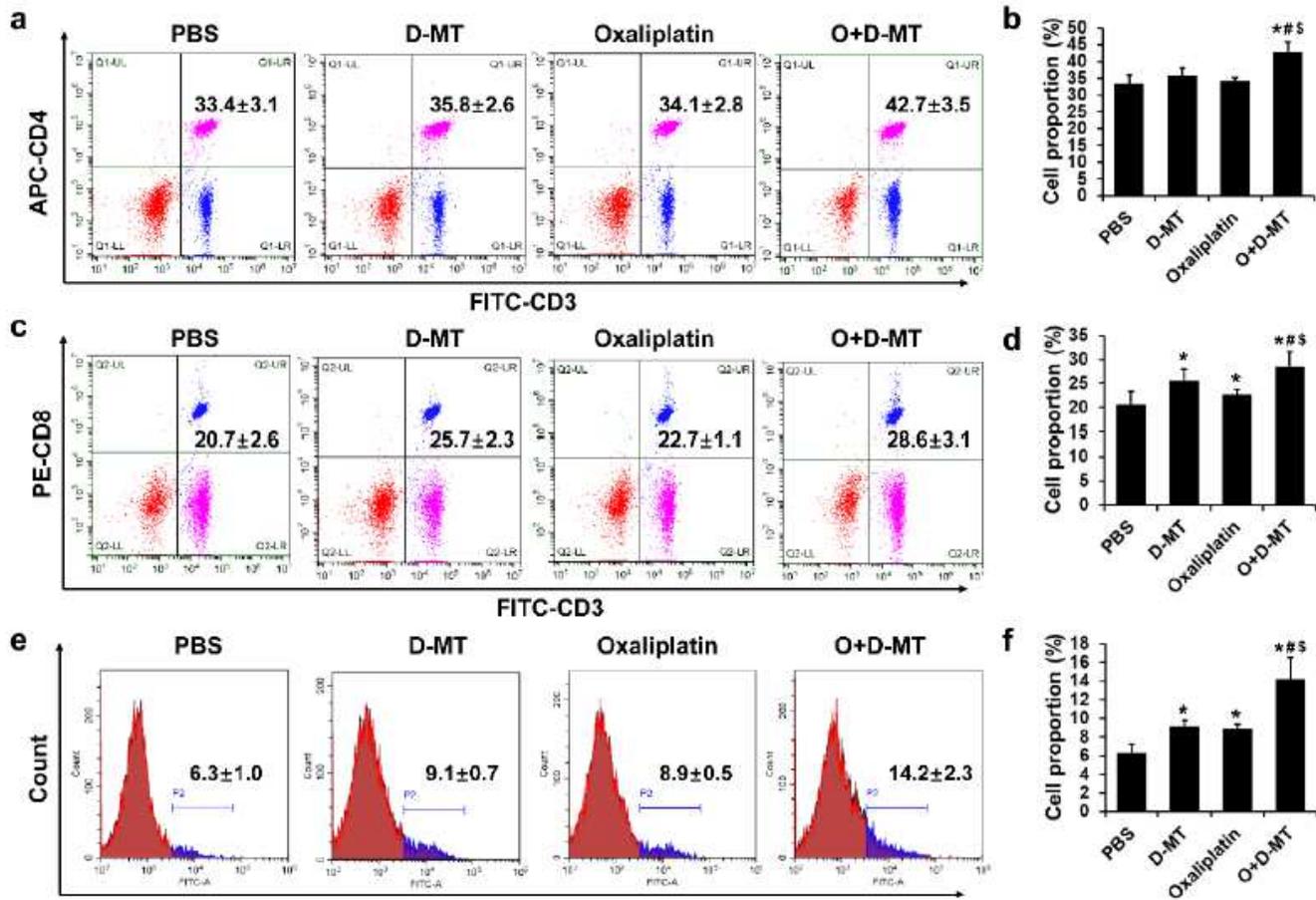


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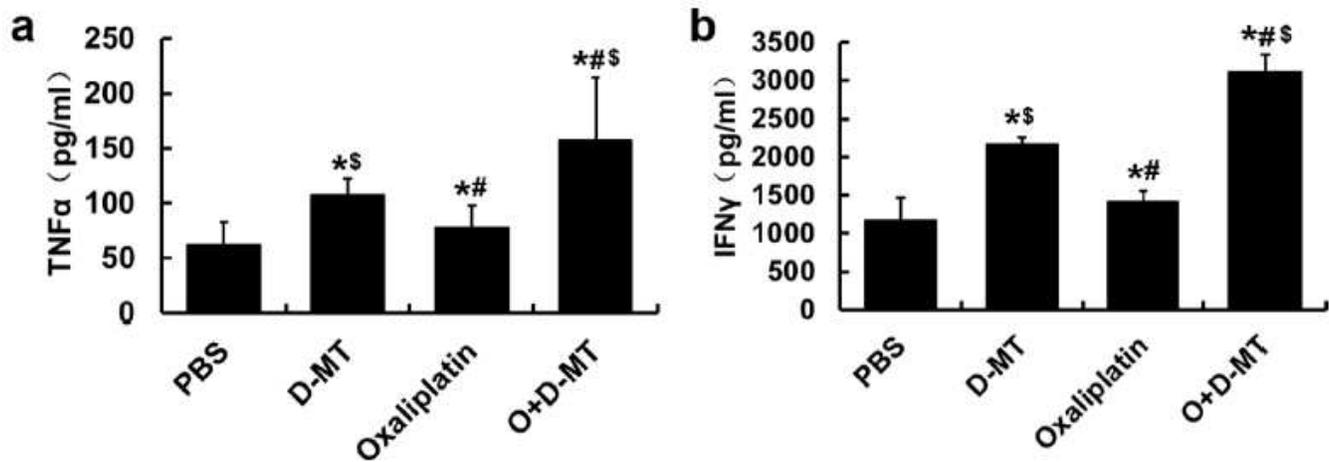


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