

# Complete genome sequence of *Bacillus velezensis* YYC, a bacterium isolated from the tomato rhizosphere

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## Research Article

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# Abstract

The *Bacillus velezensis* YYC strain was isolated from the tomato rhizosphere. In a previous experiment, it increased tomato growth and induced systemic resistance against *Ralstonia solanacearum*. However, information on its genomic content is lacking. The complete genome sequence of the bacterium was described in this study. The genome size was 3,973,236 bp and consisted of 4,034 genes in total, with a mean G + C content of 46.52%. 86 tRNAs and 27 ribosome RNAs were identified. 14 clusters of secondary metabolites were identified. The KEGG database analysis showed that 69 genes were related to quorum sensing, which were important for cross-kingdom communication. In addition, genes involved in promoting plant growth and triggering plant immunity were identified from the genome. Based on digital DNA–DNA hybridizations (dDDH), *B. velezensis* YYC was the most closely related with *B. velezensis* FZB42. However, compared with *B. velezensis* FZB42, the lanipeptide biosynthesis gene cluster was special and only existed in the genome of *B. velezensis* YYC. The complete genome data of *B. velezensis* YYC will provide a basis for explanation of its growth-promoting mechanism and biocontrol mechanism.

## Introduction

*Bacillus velezensis* is an important member of plant growth-promoting rhizobacteria. This species was found to have multiple growth-promoting effects and to produce a variety of secondary metabolites with antibacterial activity (Chowdhury et al. 2015). *B. velezensis* can be widely isolated from diversified environments, such as plant rhizospheres, soil, rivers, human food, animal guts and seawater, and can easily be isolated and cultured (Ye et al. 2018). In our work, the *B. velezensis* YYC strain was isolated from the tomato rhizosphere in Heilongjiang Province, China. *B. velezensis* YYC increased tomato growth and induced systemic resistance against *Ralstonia solanacearum* (unpublished data). Strain YYC is a non-pathogenic bacterium. Genome sequencing of *B. velezensis* YYC will provide basic insight into the growth-promoting and biocontrol mechanism.

## Data description

### Genome sequencing, assembly and annotation

*B. velezensis* YYC strain was propagated in Luria-Bertani broth with shaking at 180 r/min overnight at 30°C. By alignments of the 16S ribosome RNA and housekeeping genes, it was identified as *Bacillus velezensis*. Bacterial genomic DNA extraction kit (Majorbio Bio-pharm Technology Co., Ltd., Shanghai, China) was used to extract genomic DNA. A TBS-380 fluorometer (Turner Bio Systems Inc., Sunnyvale, CA) was used to quantify the purified genomic DNA. PacBio RS II Single Molecule Real Time (SMRT) and Illumina sequencing platforms were used to sequence the genomic DNA. The sequencing yielded 170,436 reads, including 1,341,760,841 bp, with 337.7× sequence depth. A statistic of quality information was applied for quality trimming, by which the low-quality data could be removed to result in clean data. Using Unicycler (Version 0.4.7) (Wick et al. 2017), the reads were assembled into contigs. A complete genome

was generated by inspecting and completing the last circular step. Finally, using the Illumina reads, error correction of the PacBio assembly results was performed.

The number of protein coding sequences (CDSs) in the *B. velezensis* YYC genome was predicted by Glimmer (version 3.02) (<http://ccb.jhu.edu/software/glimmer/index.shtml>) (Delcher et al. 2007) and GeneMarkS software (version 4.3) (Besemer et al. 2005). The transfer RNA (tRNA) gene was analyzed by tRNAscan-SE v2.0 software (Version 2.0) (<http://trna.ucsc.edu/software>) (Chan et al. 2019). Barrnap software (Version 0.8) (<https://github.com/tseemann/barrnap>) was utilized to predict ribosome RNA genes. By aligning reads with the Nonredundant (NR), Swiss-Prot, Kyoto Encyclopedia of Genes and Genomes (KEGG) (Kanehisa et al. 2016), Gene Ontology (GO) (Ashburner et al. 2000), Cluster of Orthologous Groups of proteins (COG) (Galperin et al. 2015) and protein families (Pfam) (Finn et al. 2014) databases, all genes were annotated. The bioactive secondary metabolites were predicted by antiSMASH software (Version 4.0.2) (Weber et al. 2015).

### **General genome features of *B. velezensis* YYC**

Whole-genome sequencing showed that the *B. velezensis* YYC strain contained a genome size of approximately 3,973,236 bp, with an average G + C content of 46.52%. The Glimmer program predicted that the number of protein coding sequences (CDSs) was 4,034, and the average gene length was 877.29 bp. Furthermore, a total of 86 tRNA and 27 ribosome RNA genes were identified and analyzed in the genome. By aligning the genome to sequences from diverse databases, including the NR, Swiss-Prot, Pfam, COG, GO and KEGG databases, the numbers of identified genes were 4,034, 3,533, 3,337, 3,013, 2,668, and 2,163, respectively.

The KEGG database analysis showed a great number of two-component systems (113 genes) and ABC transporters (117 genes). Meanwhile, 69 genes were related to quorum sensing, which were important for cross-kingdom communication (Schikora et al. 2016).

## **Secondary metabolites related to biocontrol**

AntiSMASH version 4.0.2 analysis identified 14 clusters of secondary metabolites (Fig. 1). Seven clusters of secondary metabolites were related to the synthesis of bacillaene, macrolactin, bacilysin, fengycin, difficidin, surfactin and bacillibactin. Some of these substances have antagonistic effects on bacteria, fungi, and viruses (Moldenhauer et al. 2007; Chowdhury et al.

2015; Wu et al. 2015; Zhang et al. 2018). Fengycin was found to induce resistance to plant diseases (Farzand et al. 2019).

### **Comparative genomics of diverse *Bacillus* strains**

In addition, based on digital DNA–DNA hybridizations (dDDH), *B. velezensis* YYC was the most closely related with *B. velezensis* FZB42 (Table 1). And it shared 96.80% identity with the strain that was used as biofertilizer and biocontrol agent (*B. velezensis* FZB42). Compared with FZB42, lantipeptide biosynthesis

gene cluster was special and only existed in the genome of *B. velezensis* YYC. It is engaged in the synthesis of locillomycin, which was related to hemolytic activity, swarming motility, biofilm formation, and colony morphology (Luo et al. 2019). In addition, four clusters encoding new metabolites with no reported description previously. Comparative analysis of secondary metabolite clusters of these strains were summarized for comparisons (Supplementary Table S1).

Table 1  
Pairwise comparisons of *B. velezensis* YYC with type strain genomes

Subject strain	dDDH (d0, in %)	C.I. (d0, in %)	dDDH (d4, in %)	C.I. (d4, in %)	dDDH (d6, in %)	C.I. (d6, in %)	G + C content difference (in %)
<i>Bacillus velezensis</i> FZB42	96.8	[95.1– 98.0]	90.8	[88.5– 92.6]	97.5	[96.3– 98.4]	0.05
<i>Bacillus velezensis</i> NRRL B-41580	91.8	[88.8– 94.0]	85.4	[82.7– 87.7]	93.3	[91.0– 95.0]	0.21
<i>Bacillus methylophilicus</i> KACC 13105	95	[92.7– 96.6]	84.6	[81.9– 87.0]	95.5	[93.7– 96.8]	0.09
<i>Bacillus siamensis</i> KCTC 13613	89.1	[85.8– 91.8]	56.9	[54.2– 59.7]	85.4	[82.3– 88.1]	0.19
<i>Bacillus vanillea</i> XY18	89	[85.6– 91.7]	56.9	[54.2– 59.7]	85.3	[82.2– 88.0]	0.2
<i>Bacillus amyloliquefaciens</i> DSM 7	82.7	[78.8– 86.0]	56	[53.2– 58.7]	79.8	[76.3– 82.8]	0.44
<i>Bacillus nakamurai</i> NRRL B-41091	73.4	[69.5– 77.1]	31	[28.6– 33.5]	61.2	[57.9– 64.4]	1.26
<i>Bacillus tequilensis</i> NCTC13306	30.9	[27.5– 34.5]	21.3	[19.0– 23.7]	27.5	[24.6– 30.6]	2.54
<i>Bacillus spizizenii</i> TU-B- 10	33.6	[30.2– 37.2]	21	[18.8– 23.4]	29.2	[26.3– 32.3]	2.7
<i>Bacillus subtilis</i> NCIB 3610	32.5	[29.1– 36.0]	20.9	[18.7– 23.3]	28.4	[25.5– 31.5]	3.13

The dDDH values were provided along with their confidence intervals (C.I.).

## Genes involved in promoting plant growth and triggering plant immunity

In addition to producing secondary metabolites with antifungal or antibacterial activity, *B. velezensis* YYC contains a various of genes implicated in biofilm formation and root colonization (Table 2). *B. velezensis* YYC contained the genes encoding acetolactate synthase (*ilvH*, *ilvB*), acetolactate decarboxylase (*alsD*),

and butanediol dehydrogenase (*butB*), which have plant growth–promoting effects including stimulating root formation and increasing systemic disease resistance (He et al. 2013; Jayakumar et al. 2020). *B. velezensis* YYC also has the genes required for synthesis of 2,3-butanediol (*alsD*), the compound reported to trigger systemic resistance (He et al., 2013).

Table 2  
 Representative genes of *B. velezensis* YYC probably involved in plant bacterium interactions

Gene	Position	Protein	Description
<i>spo0A</i>	2432908 – 2432108	Sporulation transcription factor Spo0A	Biofilm formation
<i>sinR</i>	2466273– 2466614	Transcriptional regulator	Biofilm formation
<i>abrB</i>	45929 – 45645	Transition state regulator Abh	Biofilm formation
<i>resE</i>	2260204– 2261985	Sensor histidine kinase	Biofilm formation
<i>lytS</i>	2757501– 2759282	Sensor histidine kinase	Biofilm formation
<i>ycbA</i>	248181– 249473	Sensor histidine kinase	Biofilm formation
<i>sacB</i>	3926734– 3928215	Levan sucrose	Root adhesion
<i>efp</i>	2452165– 2452722	Elongation factor P	Essential for swarming motility
<i>comP</i>	3033195– 3035495	Histidine kinase	Regulator of surfactin production
<i>fliD</i>	3414679– 3416199	Flagellar capping protein	Elicitation of plant basal defence
<i>flgK</i>	3419847– 3421364	Flagellar hook-associated protein FlgK	Elicitation of plant basal defence
<i>xynB</i>	1846654– 1848069	1,4-beta-D-xylan xylohydrolase	Carbohydrate metabolism
<i>lacR</i>	1216343– 1217104	Lactose phosphotransferase system repressor	Lactose metabolism
<i>lacG</i>	1214690– 1216090	6-phospho-beta-galactosidase	Hydrolyzation of phospholactose
<i>lacE</i>	1212603– 1214300	Phosphotransferase system	Cellobiose degradation
<i>lacF</i>	1214312– 1214626	Phosphotransferase system	Cellobiose degradation
<i>alsD</i>	3488161– 3488928	Acetolactate decarboxylase	Synthesis of 2,3-butanediol
<i>pta</i>	3639358– 3640329	Phosphotransacetylase	Strongly upregulated by root exudate

Gene	Position	Protein	Description
<i>ilvH</i>	2697631– 2698149	Acetolactate synthase	Promote plant growth
<i>ilvB</i>	2698146– 2699963	Acetolactate synthase	Promote plant growth
<i>alsD</i>	3488161– 3488928	Acetolactate decarboxylase	Promote plant growth
<i>butB</i>	629733– 630773	Butanediol dehydrogenase	Promote plant growth

## Nucleotide sequence accession number

The complete genome sequence of *B. velezensis* YYC was deposited in GenBank under accession number CP075055. (BioProject: PRJNA728388, BioSample: SAMN19079027).

## Declarations

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### Compliance with ethical standards

**Conflict of interest** The author(s) declare no conflict of interest.

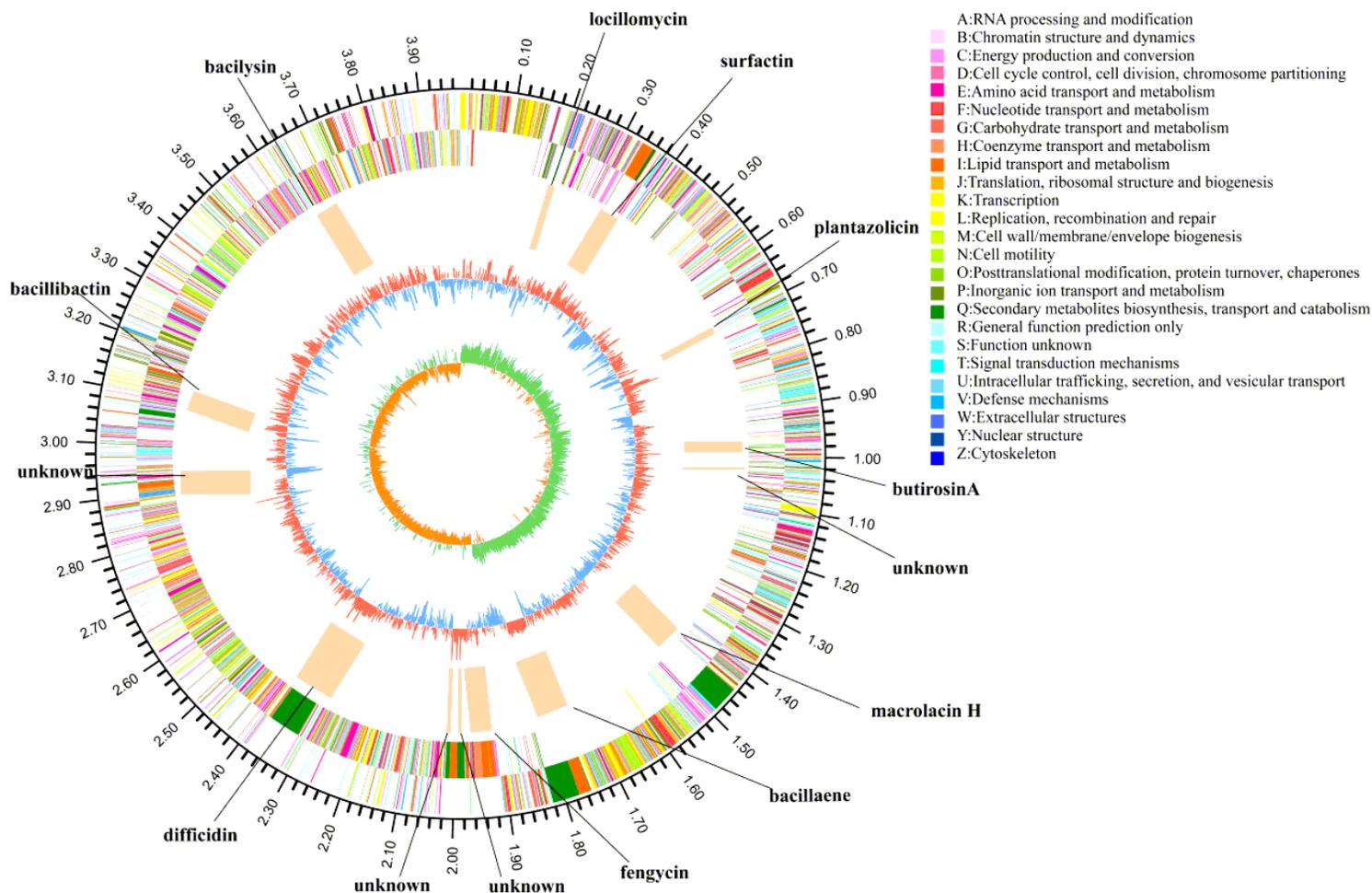
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## Figures



**Figure 1**

Circular representation of *Bacillus velezensis* YYC genome. First circle represents the genome size of *B. velezensis* YYC; Second and third circles are positive and negative chain genes, respectively, all genes are color coded according to their COG annotation functions; Fourth circle indicates nomenclature and locations of predictive secondary metabolite clusters excluding microcins; Fifth circle is GC content (red indicates greater than average value and blue indicates less than average).

## Supplementary Files

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