

# Performance and Mechanism of the in-situ Restoration Effect on VHCs in the Polluted River Water Based on the Orthogonal Experiment: Photosynthetic Fluorescence Characteristics and Microbial Community Analysis.

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## Research Article

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1 Performance and mechanism of the in-situ restoration effect on VHCs in the polluted river  
2 water based on the orthogonal experiment: photosynthetic fluorescence characteristics and  
3 microbial community analysis.

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10

11     **Abstract:**

12     Volatile halogenated hydrocarbons (VHCs) attracted many attentions due to its toxic and  
13     persistent in the environment. In this research, a novel in situ ecological restoration reactor  
14     was applied to the degradation of VHCs in polluted river water. The optimized working  
15     condition adaptation of the in-situ restoration technique was evaluated through orthogonal  
16     tests. The experiments showed that when the water depth was 0.4 m, the HRT was 5d and the  
17     current velocity was 1 m/s, VHCs removal efficiencies could achieve favorable value. The  
18     CHCl<sub>3</sub> CCl<sub>4</sub>, C<sub>2</sub>HCl<sub>3</sub> and C<sub>2</sub>Cl<sub>4</sub> removal efficiency could reach 70.27%, 70.59%67.74% and  
19     81.82%, respectively. F test results showed that both HRT and water depth were significantly  
20     related to the removal efficiency of reactor. Besides, using underwater modulated chlorophyll  
21     fluorometer analyzed the rapid light curves (RLC) of plants in the experiment, which showed  
22     that the VHCs of damaged river was harmful to the physiological state of the plants.  
23     Moreover, the microbial community structures of fillers in the reactor were tested by high-  
24     throughput sequencing, the findings supported that the microbial community made a great  
25     response to adapt to the changes of environment of the reactor. The relative abundance of  
26     *Rhodocyclaceae* increased slightly, which hinted that it had good adaptability to VHCs in  
27     polluted river water. The research results confirmed that in situ ecological restoration reactor  
28     was an economical technology for removal VHCs in polluted river water.

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34     Key words: Volatile halogenated hydrocarbons; Orthogonal tests; in-situ ecological  
35     restoration; Microbial community

36      **1.Introduction**

37      Urbanization in China has accelerated the transformation of geomorphological features  
38      and ecological environment in urban river, which contained the decrease on the hydrological  
39      connectivity and the pollutants accumulated at the bottom of the river channels (Wang et al.,  
40      2017). With the process of urbanization continues to move forward, the destruction intensified  
41      of urban water environment, especially in urban rivers and lakes environment (Lu et al., 2018).  
42      Human activity changes the original shape of the river, injects additional pollutants to affect  
43      the water quality frequently (Hirsch et al., 2010). Volatile halogenated hydrocarbons (VHCs)  
44      were constituted with a series of diversified organic compounds, most of which with the  
45      characteristics of environmental persistence, bioaccumulation and biology toxicity (Zou et al.,  
46      2006; Song et al., 2017). Meanwhile, some VHCs instant vaporized into air environment as a  
47      consequence of low boiling points, adding air pollutant and the local people risk of illness in  
48      urban area (Mun & Townley, 2021). VHCs has been widely accepted as a major source of  
49      photochemical smog since the beginning of the 21th century. Besides, variety of VHCs  
50      produced by plastics petroleum and landfill leachate influx into urban river alteration the  
51      characteristics of water environment and initiation potential adverse effects on human (Wang  
52      et al., 2020). VHCs would cause continuous hazard in environmental media, which was the  
53      particularly sensitive and prominent in river within the city limits.

54      River pollution caused by human activities is very common in world, many regions and  
55      countries begin to show concerns for the river quality and take measures to treat it. As we all  
56      know, many factors such as the water depth and current velocity, regulate actual situation of  
57      the river and affect pollutant distribution in the damaged river (Jiang et al., 2017; Rugner et al.,

58 2019). Due to the complexity characteristics of river pollution, the traditional water treatment  
59 process is difficult for practical application (Giripunje et al., 2015).

60 Actually, both phytoremediation and slope wetland system are known as green and  
61 efficient in situ water environment remediation treatments, which could alter water quality  
62 and flow regime to a certain extent (Ladislas et al., 2011). *Submerged macrophytes* as a  
63 representative eco-engineering plants, it also is an important component of river ecosystem,  
64 which have capable of influencing various biotic and abiotic environments, including the  
65 status of the rivers (Zhang et al., 2018; Özgencil et al., 2020). Slope wetland system has  
66 characterized by good geomorphology adaptability and flexible controllability, which was  
67 suitable for recovery and remediation of damaged river (Wang et al., 2021). Aimed at  
68 maintenance and restoration polluted river water, the two technologies need to be suitable  
69 combination with low cost and simple management mode (Nilsson et al., 2015). The  
70 combination of the two methods can not only expand the suitable construct methods to the  
71 riverway physiognomy, but also improve the removal efficiency on the organic matters in the  
72 polluted river water. Additionally, both *Submerged macrophytes* and slope wetland are  
73 biological systems involved in the degradation of organic matter. And the plants in  
74 conjunction with microflora in the system, which would play cleaning function on the  
75 coupling system by the bioremediation function (Wang et al., 2017). Hence, the regulation on  
76 plant physiological activities and microbial community status was a vital concern, which  
77 involve in optimized the pollutant removal and performance on coupling system.

78 *Water pollution prevention action plan* has been applied to numerous black-stinking  
79 water bodies in China, which met the requirements on elimination eutrophication and promote

80 water resources management (Song et al., 2017). Many rivers restoration projects have been  
81 applied to urban river remediation through the last few years, but the degradation VHCs in  
82 polluted river water have been rarely reported. In view of VHCs are toxic and persistent in the  
83 environment, determining its content of polluted river water and proposed appropriate  
84 measures to restore the river ecosystem is essential measures to local population health.

85 In view of the above aspects, in situ restoration reactor was used as the research carrier,  
86 and the purification effect on VHCs in the polluted river water was investigated. After due  
87 consideration, different experimental factors were set on orthogonal the outdoor simulation  
88 experiment to determine the optimal conditions on the reactor. The underwater modulation  
89 chlorophyll fluorescence was employed to analyze the physiological state of plants at the in-  
90 situ restoration reactor at the same. In addition, the headspace gas chromatography was used  
91 as detected the content of VHCs in damaged river and tested the VHCs variation on polluted  
92 river water under the system treatment. Besides, the change of substrate microorganism was  
93 measured before and after the experiment to evaluate the flora on the dichlorination reaction.

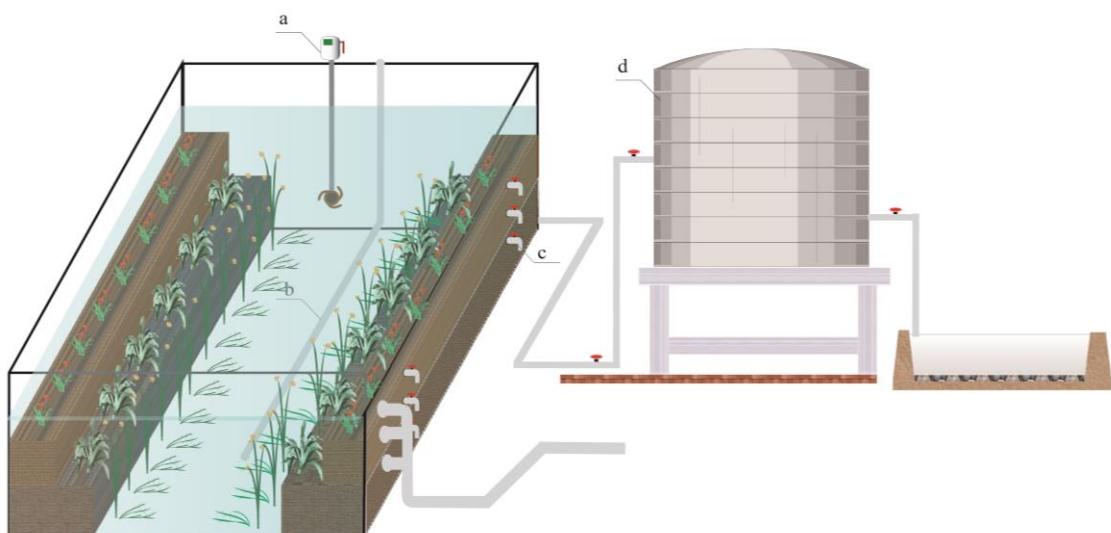
94 The aim of this research was to explore and optimize the actual operation requirement of the  
95 in-situ restoration reactor through microbial resource management and some relevant analysis  
96 instruments. And the impact on experimental factors were evaluated and pollutant  
97 decomposition mechanism in the reactor were comprehensively analyzed.

## 98 **2. Materials and methods**

### 99 **2.1 Experimental device**

100 To assure an effective and authenticity research, the experiment was carried out outdoors.  
101 One device was set up in the experiment, which dimension was 1m high, 2m wide and 7m

long. The experimental devices were consisted of water inlet pipe, water outlet, water tank and sampling ports. In the flank of all reactors, three wastewater sampling pipes are arranged in three rows with the height of 0.4m, 0.6m and 0.8m (to the bottom). Slope wetland contained three plants (*ScirpusvalidusVahl*, *Typha orientalis Presl* and *Lythrum salicaria L.*), which was planted from bottle to up, respectively. And planting density was set at 30clumps/m<sup>2</sup>. *Potamogeton wrightii Morong* and *Potamogeton pectinatus* was closely adjacent seeded around the *ScirpusvalidusVahl* in the sediment of the reactor, and the planting density was set at 30clumps/m<sup>2</sup>. The pushing flow device (QJB0.85) was purchased from Sapphire environmental protection (Nanjing, China), and impeller diameter was 220mm. In order to guarantee the replenishment and drain of the reactor steadily, the polluted river water employed in the reactor was supplied through the water tank. In addition, the polluted river water directly extracted from the damaged river by the water pump, which enhance the reliability of the simulation experiment and guarantee the authenticity of the results to a certain extent. The experimental reactor is shown in Fig.1.



**Fig.1** Experimental reactor (a. plug flow device b. water replenishing pipe c. sampling ports d. water tank)

119      **2.2 Experimental chemical and instrument**

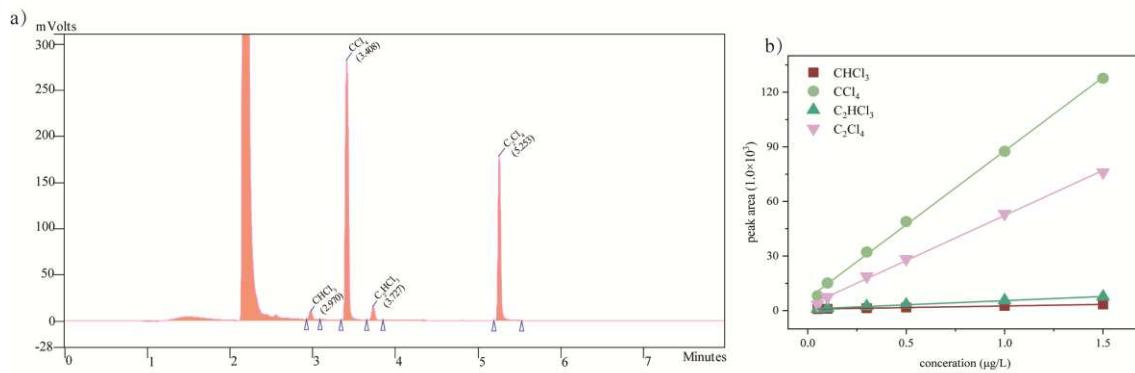
120      The chemicals obtained methanol and ethanol, which was used in the experiment were of  
121      guaranteed reagent (GR) and purchased from Beijing chemical plant (Beihua, China).  
122      Standard solutions (GSB07-1982-2005) of  $1,000 \mu\text{g}\cdot\text{mL}^{-1}$  VHCs in ethanol were purchased  
123      from standard sample Institute of Ministry of environmental protection, including  $\text{CHCl}_3$ ,  
124       $\text{CCl}_4$ ,  $\text{C}_2\text{HCl}_3$  and  $\text{C}_2\text{Cl}_4$ . Analytical grade of  $\text{NaCl}$  were purchased from Yongda Chemical  
125      Reagent Co., Ltd (Tianjin, China). Headspace bottle and micropipettor (1  $\mu\text{L}$ , 10  $\mu\text{L}$ , 50  $\mu\text{L}$ )  
126      were purchased from high pigeon instrument (Shanghai, China).  $1,000 \mu\text{g}\cdot\text{mL}^{-1}$  chloride  
127      standard solutions (GSB 07-1267-2000) was purchased from the resources platform of the  
128      national standard material. Main instruments of the experiment contained: headspace sampler  
129      (7697A, Aglient, America), headspace gas chromatograph (7890A, Aglient, America),  
130      capillary chromatographic column (HP-5MS, Aglient, America), ultra-pure water machine  
131      (CM-R0-C2, Anshi, China), underwater modulated chlorophyll fluorometer (DIVING-PAM-  
132      II, Zealqueit, China) and associated accessories.

133      **2.3 Gas chromatography operating conditions**

134      HS-GC analysis conducted with the environmental protection standards of HJ620-2011.  
135      The micropipette was used as transfer standard solutions of VHCs, which was diluted by  
136      methanol to obtain the  $10 \mu\text{g}\cdot\text{mL}^{-1}$  preparation solutions. Preparation solutions was allowed  
137      the headspace extraction and gas phase operations relatively handy. The  $\text{NaCl}$  was dried to a  
138      constant weight ( $350^\circ\text{C}$  for 3h) and then cooled at room temperature, which was added into  
139      headspace bottle followed immediately when it has cooled. At the same time, 5, 10, 20, 30,  
140      50 $\mu\text{L}$  preparation solutions was added up respectively into 10mL ultra-pure water in the

141 headspace bottle, and the mixture sufficient agitated respectively. After the above operations,  
142 the standard solution was obtained. HS-GC operations were composed of headspace injection,  
143 high temperature separation and analyte display. A robotic arm and a headspace generation  
144 unit was consisted the 111-space autosampler, which could heat samples and convey the  
145 bottles into the GC equipment. Chromatographic conditions: Initial temperature was 40°C for  
146 2 min and increased to 100 °C at 60 °C min<sup>-1</sup>, then 100 °C to 200 °C at 10 °C min<sup>-1</sup>. Detection  
147 temperature was set as 320 °C. Helium used as carrier gas and flow velocity was 1.0mL·min<sup>-1</sup>.  
148 The results were shown in the Fig.2a).

149 The signal was gathered from GC equipment, which reflected the characteristic on  
150 analytes by the position and size of peak area. The external standard method was used to  
151 establish standard line, the results were shown in the Fig.2b).

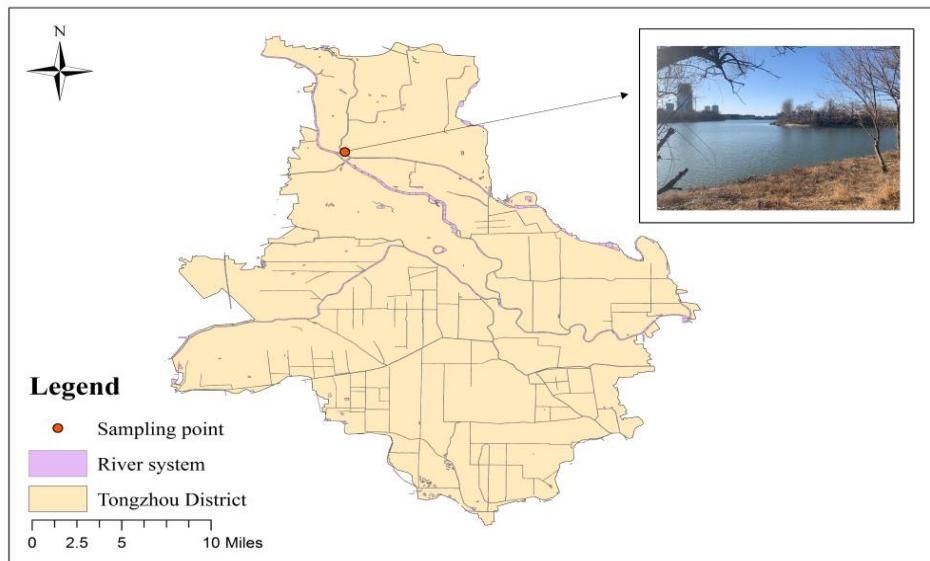


152 Fig.2 Gas chromatogram analysis of target compounds (a. Gas chromatogram of VHCs b.  
153 Standard line of VHCs)

## 155 2.4 Experimental site and influent quality

156 In this study, the Xiaozhong river in Tongzhou District of Beijing was selected as  
157 experimental site. The average annual atmospheric temperature of Tongzhou District is 13-  
158 24°C. The river runs through an urban area and it adopts the way of reclaimed water

159 replenishment, and the main stream is set with several sewage outlets, the self-purification  
160 capacity of water body is poor. The experimental site is located in Tongzhou Xiaozhong River  
161 experimental base (116°67'E, 39°93'N), and the sampling point is located at the confluence  
162 point of five rivers. Its variation can effectively reflect the interaction between upstream and  
163 downstream water quality. The experimental site is shown in Fig.3.



164

165 Fig.3 Experimental site

166 The Xiaozhong river receives multiplex sewage throughout the year, the pollutants  
167 mainly come from the surrounding domestic garbage, sewage treatment plants and rainfall,  
168 which aggravated river channel damaged situation (Zhang et al., 2020a). The integral state of  
169 river was sluggish, and some area existed stagnant zone. Using the 1000mL sample bottle,  
170 water samples were directly collected from the experimental site and were analyzed in the  
171 laboratory (sampling two times). The characteristics of polluted river water are shown in Tab.  
172 1

173 **Table 1** Characteristics of polluted river water

Parameter	$\text{CHCl}_3 (\mu\text{g} \cdot \text{L}^{-1})$	$\text{CCl}_4 (\mu\text{g} \cdot \text{L}^{-1})$	$\text{C}_2\text{HCl}_3 (\mu\text{g} \cdot \text{L}^{-1})$	$\text{C}_2\text{Cl}_4 (\mu\text{g} \cdot \text{L}^{-1})$	pH
Content	0.21	0.17	0.37	0.11	5.9~7.1

174 2.5 Experimental methods

In this study, in-situ restoration effect on polluted river water were evaluated by orthogonal tests under the water depth, HRT and current velocity as variables. In order to simulate the influence on river flow, considering the characteristics of wetlands at the same, the variation on HRT was applied to the reactor to meet the requirements. The HRT was calculated by the Eq.1. The three factors were selected as influencing factors, containing HRT(1, 3, and 5d), current velocity(1, 2, and 3m/s) and water depth(0.4, 0.6, and 0.8m) and three levels were taken for each factor. And the fictitious factors were added to the orthogonal matrix, for avoiding the influence of error caused by stochastic factors.

$$HRT = \frac{V}{O} \quad (1)$$

184       $V$ —The treatment volume in reactor ( $\text{m}^3$ )

185       $Q$ —The influent flow in reactor ( $\text{m}^3/\text{d}$ )

Meanwhile, the methods of outdoor simulation experiment were adopted to the experiment to assure the factuality of results. DNA was extracted from the soil and filler samples with a FastDNA skin for soil Isolation kit (116560200, MP, America) on the basis of product description. The extracted DNA was stored at -20°C and follow operation contain amplification, enrich and construct the template of miseq library was finished by Beijing Allwogene Tech and Monitoring Technology Co., Ltd.. The microbial community analysis of the experiment was completed by illuminamiseq pe300 sequencing platform. Water samples were collected in glass sampling bottle (50mL) and stored at 4°C until detect. The samples

194 before and after treatment conveyed by using headspace sampler (HS) and the content of  
195 VHCs in water samples was measured by headspace gas chromatograph (GC) with the help of  
196 capillary chromatographic column. Plants physiological status were analyzed by underwater  
197 modulated chlorophyll fluorometer.

198 **3. Results and discussion**

199 **3.1. The degradation on VHCs under orthogonal experiment**

200 **3.1.1. Construction of orthogonal experiment**

201 Transport of pollutants in the damaged river are closely associated with hydrodynamics  
202 situation on the polluted river water, which included current velocity, river flow, water depth  
203 and the like (Anwar Sadat et al., 2020). L<sub>9</sub>(3<sup>4</sup>) orthogonal experiment was applied in this study.  
204 Details of the factors and levels in orthogonal experiment is shown in Tab. 2.

205 **Table 2** Factors and levels in the orthogonal experiment

Factors	Level		
	1	2	3
Water depth (m)	0.4	0.6	0.8
HRT (d)	1	3	5
Current velocity (m/s)	1	2	3
Fictitious factors	1	2	3

206 **3.1.2 Orthogonal experiment results**

207 Aiming to experimental materials decrement, orthogonal experiments were constructed  
208 on representative points with mutual independence, which could validate the experimental  
209 result under various factors and obtained the optimization designation (Wang et al., 2019). The  
210 removal efficiency under different experimental variables were analyzed.

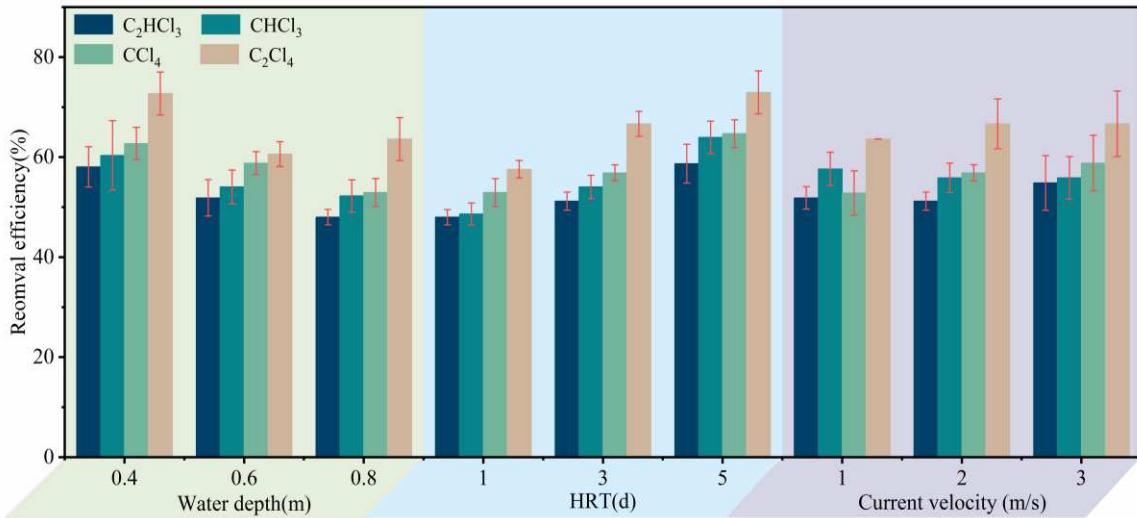


Fig.4 The VHCs removal efficiency under different influencing factors

As is shown in the Fig.4, the removal efficiency of four VHCs existed differences, which was associated with the change of influencing factors. The average removal efficiency of CHCl<sub>3</sub>, CCl<sub>4</sub>, C<sub>2</sub>HCl<sub>3</sub> and C<sub>2</sub>Cl<sub>4</sub> in the in-situ restoration reactor was 45.95%–70.27%, 47.06%–70.59%, 45.16%–67.74%, and 54.55%–81.82%, respectively. For the sake of determine the effect on experimental factors, significance tests of orthogonal experiment results were analyzed. The results were shown in the Table.3.

**Table 3** Orthogonal matrix and significance test of results

Number of groups	Water depth (m)	HRT (d)	Current velocity (m/s)	Fictitious factors
1	0.4	1	1	1
2	0.4	3	2	2
3	0.4	5	3	3
4	0.6	1	2	3
5	0.6	3	3	1
6	0.6	5	1	2
7	0.8	1	3	2
8	0.8	3	1	3
9	0.8	5	2	1
Name	CHCl <sub>3</sub>	CCl <sub>4</sub>	C <sub>2</sub> HCl <sub>3</sub>	C <sub>2</sub> Cl <sub>4</sub>
$K_1$	181.08	188.23	174.19	218.19
$K_2$	162.16	176.47	155.59	181.83
Water depth(m)	$K_3$	156.76	158.82	143.99
	$\bar{K}_1$	60.36	62.74	58.06
	$\bar{K}_2$	54.05	58.82	51.86
	$\bar{K}_3$	52.25	52.94	48
	R	24.32	29.41	30.2
				63.64
				36.36

	$K_1$	145.95	158.82	143.99	172.74
	$K_2$	162.16	170.58	153.67	200.01
	$K_3$	191.89	194.12	176.11	218.9
HRT (d)	$\bar{K}_1$	48.65	52.94	48	57.58
	$\bar{K}_2$	54.05	56.86	51.22	66.67
	$\bar{K}_3$	63.96	64.71	58.7	72.97
	$R$	45.94	35.3	32.12	46.16
	$K_1$	172.97	176.47	155.59	190.92
	$K_2$	167.57	170.58	153.67	200.01
Current velocity (m/s)	$\bar{K}_3$	167.57	176.47	164.51	200.01
	$\bar{K}_1$	57.66	52.82	51.86	63.64
	$\bar{K}_2$	55.86	56.86	51.22	66.67
	$\bar{K}_3$	55.86	58.82	54.84	66.67
	$R$	5.4	5.89	10.84	9.09
	$K_1$	162.16	176.46	154.83	200.01
	$K_2$	167.57	170.59	156.76	190.92
Fictitious factors	$\bar{K}_3$	170.27	176.47	162.18	200.01
	$\bar{K}_1$	54.05	58.82	51.61	66.67
	$\bar{K}_2$	55.86	56.86	52.25	63.64
	$\bar{K}_3$	56.76	58.82	54.06	66.67
	$R$	8.11	5.88	7.35	9.09

220      Sum of square deviance are important index for description the degree of dispersion,  
221      which was calculated by Eq.(2) and Eq.(3) in this experiment.

222

223      
$$SS_t = \frac{1}{r} \sum_{i=1}^k K_{ij}^2 - T \quad (2)$$

224

$$T = \frac{S^2}{n} \quad (3)$$

225       $SS_t$ —Sum of square deviance with each factors

226       $r, k$ —The repeat times of each level, the number of levels with each factor

227       $K_{ij}$ —Total indicators of each factor level

228       $T$ —Sum of all index

229       $n$ —Number of experiments ( $n=9$ )

230       $N = df_a = df_b = df_c = df_d = m - 1 \quad (4)$

231      
$$MS_t = \frac{SS_t}{df_t} \quad (5)$$

232       $N$ —Degree of freedom

233  $MS_t$ —Variance of each factor

234 According to Eq.(4) and Eq.(5) calculated the variance of each factor. Final F value was  
235 obtained through Eq. (6) and identify the significance on the factors.

236

$$F_t = \frac{MS_t}{MS_e} \quad (6)$$

237 Using the forgoing formulas carried out the experiment data analysis of significance test.

238 The critical value  $F_{0.05}(2,4)$  of 19.247 was selected. The results were list in Tab. 4.

239 **Table 4** ANOVA of the orthogonal experimental results

	Source of variation	SS	DO F	MS	F	$F_{c1}$	$F_{c2}$	Sig
$\text{CHCl}_3$	Water depth	108.732	2	457.45	80.466	19.247	9.243	**
	HRT	361.902	2	180.951	31.83	19.247	9.243	**
	Current velocity	914.899	2	54.336	9.558	19.247	9.243	*
	Fictitious factors	11.37	2	5.685				
$\text{CCl}_4$	Water depth	146.085	2	73.046	18.948	19.247	9.243	*
	HRT	215.391	2	107.696	28.082	19.247	9.243	**
	Current velocity	7.709	2	3.855	1	19.247	9.243	
	Fictitious factors	7.67	2	3.835				
$\text{C}_2\text{HCl}_3$	Water depth	154.729	2	77.365	15.85	19.247	9.243	*
	HRT	180.994	2	90.497	18.698	19.247	9.243	*
	Current velocity	22.306	2	11.153	2.304	19.247	9.243	
	Fictitious factors	9.68	2	4.84				
$\text{C}_2\text{Cl}_4$	Water depth	238.703	2	119.352	12.986	19.247	9.243	*
	HRT	452.319	2	226.16	24.607	19.247	9.243	**
	Current velocity	18.382	2	9.191	1	19.247	9.243	
	Fictitious factors	18.382	2	9.191				

240 SS, sum of squares; DOF, total degree of freedom; MS, mean square;  $F_{c1}$ , critical F value at  $\alpha$   
241 level of 0.05;  $F_{c2}$ , critical F value at  $\alpha$  level of 0.1; Sig, significance (\*\* Extremely significant  
242 influence  $p<0.05$ ; \* Significant influence  $p<0.1$ )

243 As seen in Table.4, when the water depth was 0.4 m, the HRT was 5d and the current  
244 velocity was 1 m/s, VHCs removal efficiencies could reach favorable value. The factor of

245 HRT was the most influential factor in the removal efficiency of  $\text{CHCl}_3$ ,  $\text{CCl}_4$  and  $\text{C}_2\text{Cl}_4$ , and

246 water depth showed an extremely notable role on the CHCl<sub>3</sub> removal in the experiment  
247 (p<0.05). The sufficient contact time between the reactor and wastewater was directly  
248 affected by HRT, which involved in alter the accumulation of aquatic ecosystems in reactor,  
249 and the value of it should be extended appropriately to promote the good performance (Su et  
250 al., 2009). Besides, the factor of water depth also showed extremely significance on CHCl<sub>3</sub>  
251 removal (p<0.05) and exhibited significant influence on CCl<sub>4</sub>, C<sub>2</sub>HCl<sub>3</sub>, and C<sub>2</sub>Cl<sub>4</sub> (p<0.1).  
252 Water depth was used as a measure of treatment volume, which related to not only the total  
253 water volume, but also influence the active quantity of the in-situ restoration system. Mathon  
254 et al (2019) reported that the degradation of micropollutants were significant decrease  
255 corresponding to the increase of water level, which was in line with our results. And the effect  
256 of reactor existed a possible link with the light intensity, which was affected by the water  
257 depth. The appropriate light intensity was conducive for promoted potential function on the  
258 plants, such as biological function and metabolism status. The plants grow particularly well  
259 with the relatively low water depth, the removal of pollutants could be accelerated when the  
260 optimal condition was obtained.

261 Current velocity had a significant influence on the CHCl<sub>3</sub> removal, but it was not  
262 obvious on the others pollutants removal effects. Some previous studies showed that a low  
263 current velocity was promoted to flocculate and precipitate of pollutants in damaged river,  
264 which was conducive to the removal of pollutants (Navarro-Ortega et al., 2010; Pan et al., 2016).  
265 The variation on the current velocity affected the reactor by altering the degree of architecture  
266 of submerged macrophytes, influencing the association on leaf-water interface and adjusting  
267 the attachment of bacteria, comprehensively. The significant influence of current velocity was

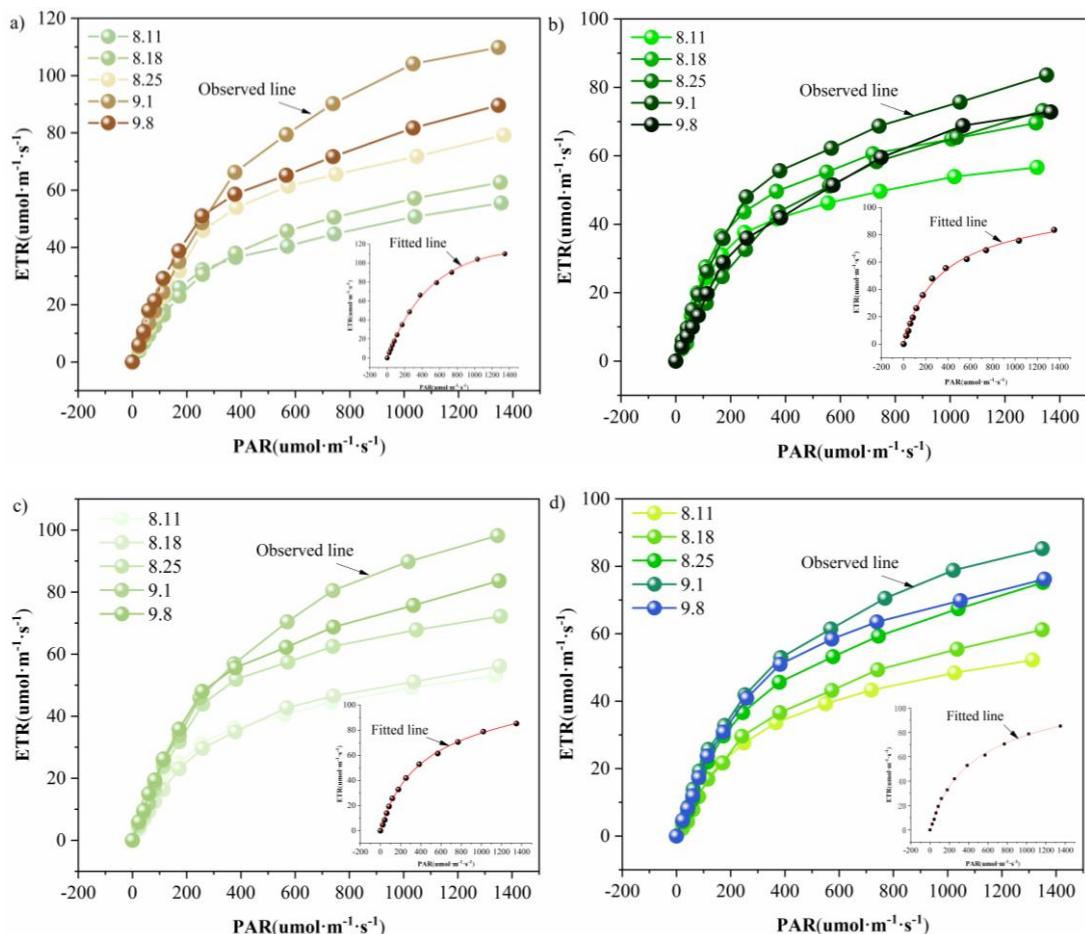
268 relatively low might be the distribution of VHCs was relatively stable under low current  
269 velocity. In addition, both sum of squares and R value of fictitious factors was relatively low  
270 indicated that no significant interaction effects existed among the variables (Xiong et al., 2020).

271 The situation mainly because extended HRT, decreased current velocity could strengthen  
272 interaction between the restoration area and polluted river water, which was conductive to the  
273 adsorption of VHCs through plants and optimizing mass transfer by epiphytic biofilms (Li et  
274 al., 2018). The decrease on current velocity would weaken the influence on flow shear,  
275 adjusting the attachment and distribution of biofilms. The increase of water depth increased  
276 the treatment volume with in situ restoration reactor, exacerbated the formation of anaerobic  
277 environment and the hydraulic dead zone (Liu et al., 2016), which would cause the removal  
278 efficiency improve more difficult.

279 In addition, the F value in the variance analysis showed that HRT and water depth had a  
280 significant impact on the VHCs removal, and the order of the factors could be summarized as  
281 HRT, water depth and current velocity for in situ restoration reactor. The evaluation index of  
282 orthogonal test was contact with its impact on the results, which exhibited positively  
283 correlation on the two indexes. Both HRT and water depth exhibited the highest significance  
284 on the VHCs removal, but current velocity had a bad correlation. However, the accuracy of  
285 orthogonal experiment seemed to be lower than single factor analyses, which was the  
286 unavoidable disadvantage of this scheme. The reason for the situation could be attributed to  
287 the restriction of the level setting (Guo et al., 2017). Considering the shortcomings of the  
288 orthogonal test, the further research should be added parallel experiment to improve the  
289 accuracy of orthogonal experiment.

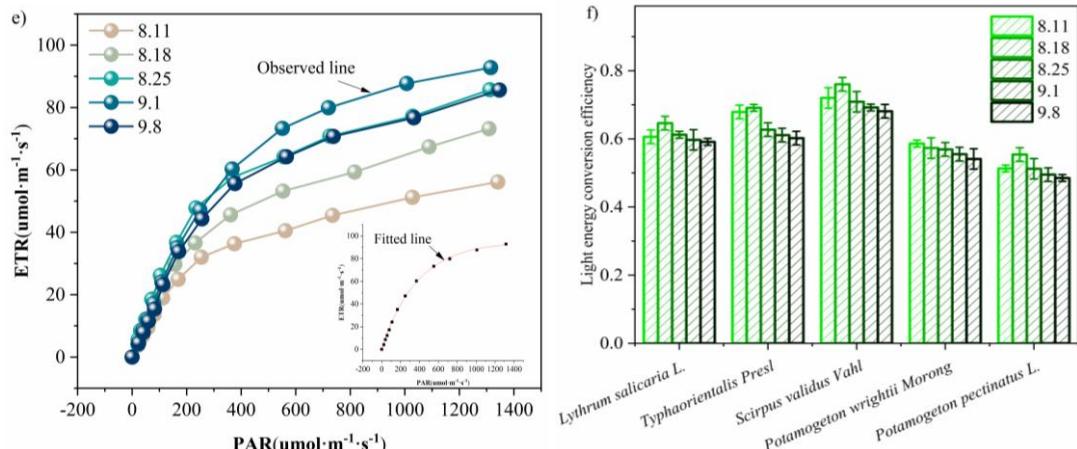
290     **3.2. Photosynthetic fluorescence characteristics upon plants**

291     The photosynthesis ability of plants was a drive factor that regulate the growth and  
 292     metabolism, implied the net assimilation rates of plants, which characterized the plant stress  
 293     responses at the same (Zhang et al., 2020b). Rapid light curves (RLC) are plots of mechanisms  
 294     of photosynthesis for 10s, it could reflect the influence of differ environmental factors on the  
 295     photosynthetic reactions (White & Critchley, 1999). The plots were measured with DIVING-  
 296     PAM-II, the results were shown in the Fig.5.



297

298



299

300 Fig.5 Photosynthetic fluorescence characteristics upon plants (a. *Lythrum salicaria L.* b.301 *Typhaorientalis Presl* c. *Scirpus validus Vahl* d. *Potamogeton wrightii Morong* e, *Potamogeton*302 *pectinatus L.* f. Light energy conversion efficiency)

303 As is shown in Fig.5, the RLCs of different plants were obtained under similar

304 temperature (20°C), which indicated the plants existed some difference at the intrinsic

305 characteristics of photosynthetic ability and electron transport, especially in stressful

306 environment. On the whole, the relative electron transfer efficiency (ETR) of plants increased

307 with the increase of photosynthetically active radiation and reaction time at the on the peak

308 value, and then decrease slightly. This phenomenon on plants caused by the physiological

309 mechanism for the adaption to environmental stress. The increase in ETR for the several

310 plants were considerably and the values of *Lythrum salicaria L.* was the highest, which

311 exhibited it had the excellent pollutant resistance. The variation on ETR was linked with the

312 adjustment on growth characteristics and photosynthesis pigments for plants (Yaghoubian et

313 al., 2016). The VHCs damaged the photosynthetic apparatus was a cumulative process in

314 plants, which were consistent with the change on RLCs under different experiment plants in

315 our experiment (Hou et al., 2018). In addition, five kinds of experimental plants existed

316 significance difference ( $p<0.01$ ), which supported that prominent diversity exists in the  
317 adaptation abilities for different types of plants.

318 As is shown in Fig.5-f), it was found that the  $F_v/F_m$  values of experiment plants were  
319 decline with the time extended, which showed the physiological state of the plant was  
320 inhibited to a certain extent. The variation on the  $F_v/F_m$  value was related to the conversion  
321 efficiency of photosystem II, which was linked with non-cyclic electron transport rates and  
322 implied the assimilation capability of plants with the help of light energy (Takahashi & Badger,  
323 2011). The decline of  $F_v/F_m$  value supported that there existed some damage in the  
324 photosynthetic machinery of experiment plants. In addition, Krall et al (2010) proposed the  
325  $F_v/F_m$  value near 0.8 was conducive for exploited full potential on the photosynthetic activity  
326 and productivity of plants. All the  $F_v/F_m$  value of experiment plants lower than 0.8, which was  
327 demonstrated the results that physiological state of the plants was endangered.

328 Using the presetting program of fluorescence induction curves and light curves, the  
329 parameters of the photosynthesis were obtained. According to the according to the model of  
330 Frankenbach and SeroDio (2017), the best RLCs of five kinds of experiment plants were fitted  
331 by the Eq. (7).

332

$$ETR = \frac{PAR}{a \cdot PAR^2 + b \cdot PAR + c} \quad (7)$$

333 And the detailed parameters of curves included the maximum photosynthetic electron  
334 transfer rate ( $ETR_{max}$ ), the initial slope ( $\alpha$ ) and half-saturation light intensity ( $I_k$ ) was  
335 calculated by the Eq. (8), Eq. (9) and Eq. (10), respectively.

336

$$\alpha = \frac{1}{c} \quad (8)$$

337

$$ETR_{max} = \frac{1}{b + 2 \cdot \sqrt{a \cdot c}} \quad (9)$$

338  $\alpha$ —Initial slope, electrons/photon

339  $ETR_{max}$ —Maximum electron transfer rate,  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$

340 The relevant parameters were carried out through above mathematical calculation, and

341 the results were shown in the Table.5.

342 **Table 5** Detailed parameters of rapid light curves equation upon experiment plants

Name	Fast light curve equation	$\alpha$	$ETR_{max}$
<i>Lythrum salicaria L.</i>	$E = \frac{P}{1.039E^{-6} \cdot P^2 + 0.005 \cdot P + 4.013}$	0.249	211.421
<i>Typhaorientalis Presl</i>	$E = \frac{P}{1.615E^{-6} \cdot P^2 + 0.006 \cdot P + 3.836}$	0.261	171.238
<i>Scirpus validus Vahl</i>	$E = \frac{P}{5.562E^{-8} \cdot P^2 + 0.007 \cdot P + 3.82}$	0.262	137.175
<i>Potamogeton wrightii Morong</i>	$E = \frac{P}{6.411E^{-7} \cdot P^2 + 0.01 \cdot P + 3.67}$	0.272	101.423
<i>Potamogeton pectinatus L.</i>	$E = \frac{P}{4.331E^{-7} \cdot P^2 + 0.01 \cdot P + 3.111}$	0.321	95.788

343 As shown in Table. 5, by comparison, the parameters of RLCs were obvious differences

344 between the different plants.  $\alpha$  was the initial slope of the curve, which mean the efficiency of

345 light energy utilization. According to the parameters in the Table.5, the value of *Potamogeton*

346 *pectinatus L.* was highest, and the value of *Potamogeton wrightii Morong* was the second largest,

347 which exhibited that they had the strongest tolerance to strong light. The phenomenon

348 supported that the two kinds of macrophyte with submerged structure could growth healthily in

349 the water.  $ETR_{max}$  is a measure of maximum electron transfer rate, which was related to the

350 calvin cycle metabolism and showed the vitality of plants (San Bautista et al., 2011). The

351  $ETR_{max}$  value of *Lythrum salicaria L.* was highest in our test, which accord with our

352 observation. It might be provided indirect information about the physiological condition of

353 plants, which hinted the plant had superior stress resistance (Brestic & Zivcak, 2013).

354 **3.3. The microbial community structures**

355 The removal of VHCs in in situ restoration reactor mainly relies on biological function,

356 which was similar with the pollutant removal role on constructed wetland (Tang et al., 2020).

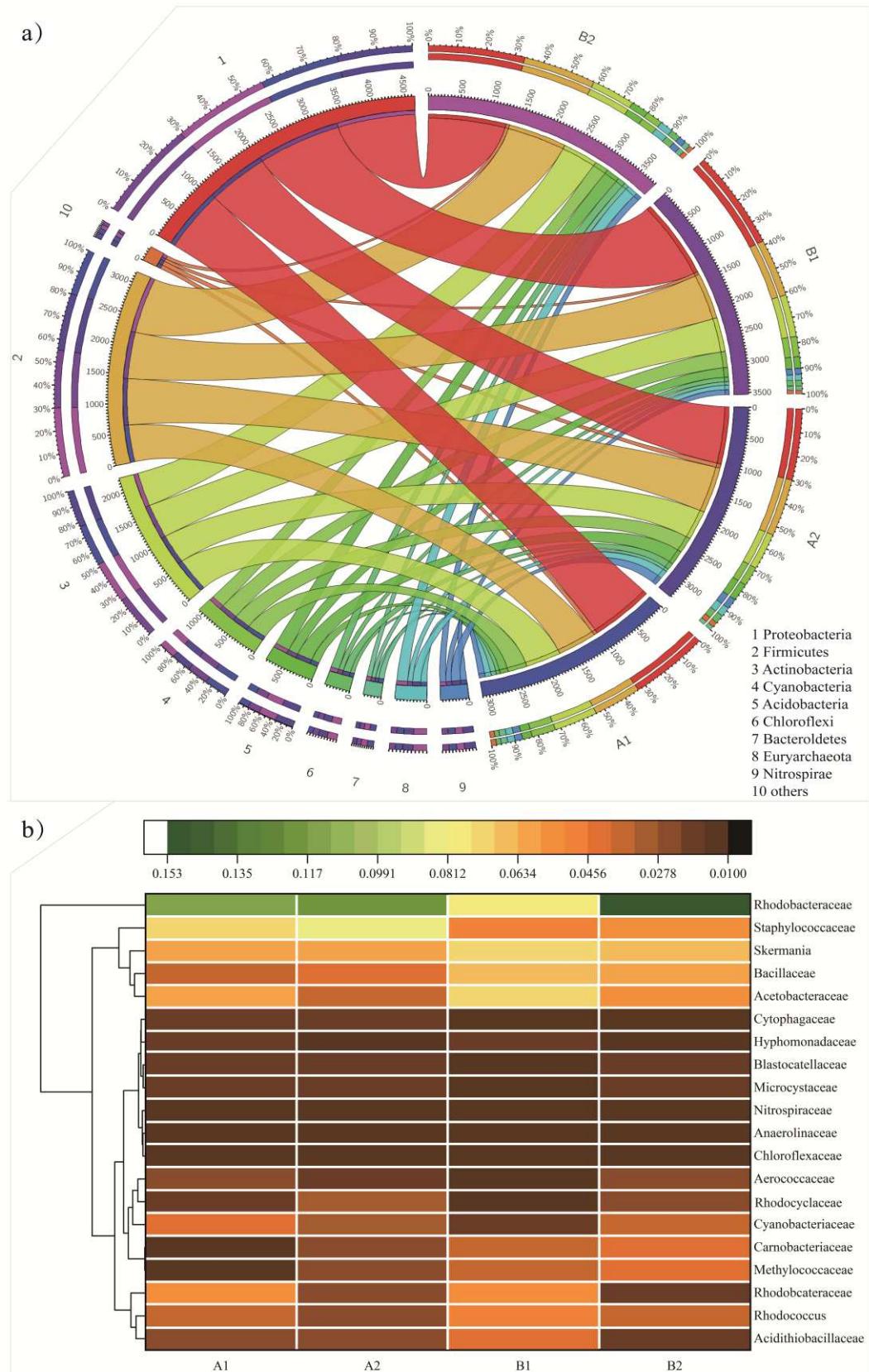
357 in situ restoration reactor of different plants and substrates can induce changes in microbial

358 community structure, which influenced on the energy and complex compounds

359 transformation efficiencies. Give full consideration to the characteristics of the fillers, the

360 samples of ceramsite and zeolite were collected during the experiment and analyzed in the

361 end. The results were shown in the Fig.6.



362

Fig.6. Relative abundance of the main microbial composition (a) phyla level (b) Family level

364 The distribution of microorganisms at phyla level in fillers are shown in the Fig.6. A1  
365 and A2 are ceramsites at the 9d and 27d, B1 and B2 are zeolites at the 9d and 27d, and a total  
366 of 3101, 3320, 3530, 3805 OTUs were identified from the four samples, respectively.  
367 Generally, 10 phyla of the bacterial communities were contained in the samples. The biomass  
368 samples taken from the in-situ restoration reactor were dominated by *Proteobacteria*,  
369 *Firmicutes*, *Cyanobacteria*, and *Actinobacteria* respectively at the phylum level, which was  
370 account for over half of the microbial community. These microbial communities were always  
371 play an important role in the degradation of organic matter, which had been found in the  
372 rhizosphere zone usually (Li et al., 2020a; Man et al., 2020). *Proteobacteria* and *Firmicutes*  
373 had been reported that included various bacteria with hydrolysis and could degrade complex  
374 compounds, which was common flora of macrophyte rhizospheres in the fillers of wetland  
375 (Huang et al., 2020; Yang et al., 2020). *Cyanobacteria* was a common component of soil-root  
376 interface in wetland substrate, in which some species involved in nutrients removal (Mandal et  
377 al., 1992). Besides, *Actinobacteria* were quotidian microorganisms in wetland ecosystem,  
378 whose existence was advantageous to the recycling of refractory biomaterials (Hassan et al.,  
379 2017).

380 As is shown in the Fig.6-a), after 18 days of operation of the in-situ restoration reactor,  
381 the distribution of biomass was more balanced both in zeolites and ceramsites. There are  
382 some previous researches supported that the microbial community structure would be evolved  
383 in order to constantly adapt to the variation of the environment, which might be related to the  
384 decrease of *Proteobacteria* (Kobayashi et al., 2020; Zheng et al., 2020). Some of *Acinobacteria*  
385 had specific degradation capacity, such as *Streptomyces*, *Rhodococcus*, which could

immensely helpful for the degradation of recalcitrant compound (Fatahi-Bafghi, 2019). The decrease of the relative abundance of *Actinobacteria* may be related to the acidogenic decomposition process on the VHCs during the experiment. *Firmicutes* contained many microbes, which was responsible for the structure on the bacterial colony. The increase of *Firmicutes* implied that the stability of bacterial biofilm was improved in the fillers (Chouari et al., 2005). Additionally, the content of *Chloroflexi* increased in the fillers, whose existence was advantageous to the dichlorination process in the reactor. In fact, *Firmicutes* and *Chloroflexi* contained a number of microorganisms had with the potential to biodegrade VHCs, such as *Cyanobacteria*, *Bacillus* and *Staphylococcus*, and the change of the two phylum was conducive to the improvement of in situ restoration reactor performance (Guo et al., 2020; Yuan et al., 2017). According to the changes of *Bacteroidetes*, it was founded that there existed different on the content of zeolites and ceramsites. Wolinska et al (2017) found *Bacteroidetes* could be used as a biological index of microbial community structure, and the amount of it would increase when the bacterial community encountered poor environment. And the amount of *Bacteroidetes* increased in the zeolites was conformed to the characteristics of materials, the fillers could adsorb considerable VHCs. And because the zeolites had better adsorption capacity, generous VHCs would accumulate on this filler, which exacerbated the difficult on the living of microorganisms. In addition, some researchers have been reported that *Acidobacteria*, *Cyanobacteria* and *Nitrospirae* were essential factors in wetland substrate, which could maintain the function of wetland (Li et al., 2020b). All these floras were observed as the dominant bacteria in experiment reactor, while the proportion of these phylum were declined in the later period of experiment. It was proved that there existed

408 certain adjustment on the microbial community to suit the change of the influent.

409 As is shown in the Fig.6-b), 20 species were determined as dominant bacteria, numbers  
410 from *Rhodobacteraceae*, *Staphylococcaceae*, *Skermania* were most frequently detected at the  
411 family level. Some previous research has been reported that some bacterial strains belong to  
412 *Rhodobacteraceae* showed excellent capacity to biodegrade a wide range of organic  
413 compound, which were used as bioremediation (Cappelletti et al., 2020). It was the most  
414 abundant family in zeolites and ceramsites, no matter in initial or last stage during the  
415 experiment. *Staphylococcaceae* was the second most abundant phylum in our results and the  
416 relative abundance of *Skermania* was followed closely. *Staphylococcaceae* and *Skermania*  
417 included a group of microorganisms had nutrients removal capacity, often as a basic  
418 component of activated sludge or wetland structure (Milobedzka & Muszynski, 2015; Wang et  
419 al., 2016). Furthermore, *Methylococcaceae* was predominance bacterial group, some  
420 researchers demonstrated that it contained some types methanotrophs that could utilize the  
421 methane to gain energy (Albers et al., 2015). The proportion of *Methylococcaceae* at the family  
422 level was promoted obviously, which was probably related to the affluent CH<sub>4</sub> originated from  
423 the reductive action on the VHCs decomposition.

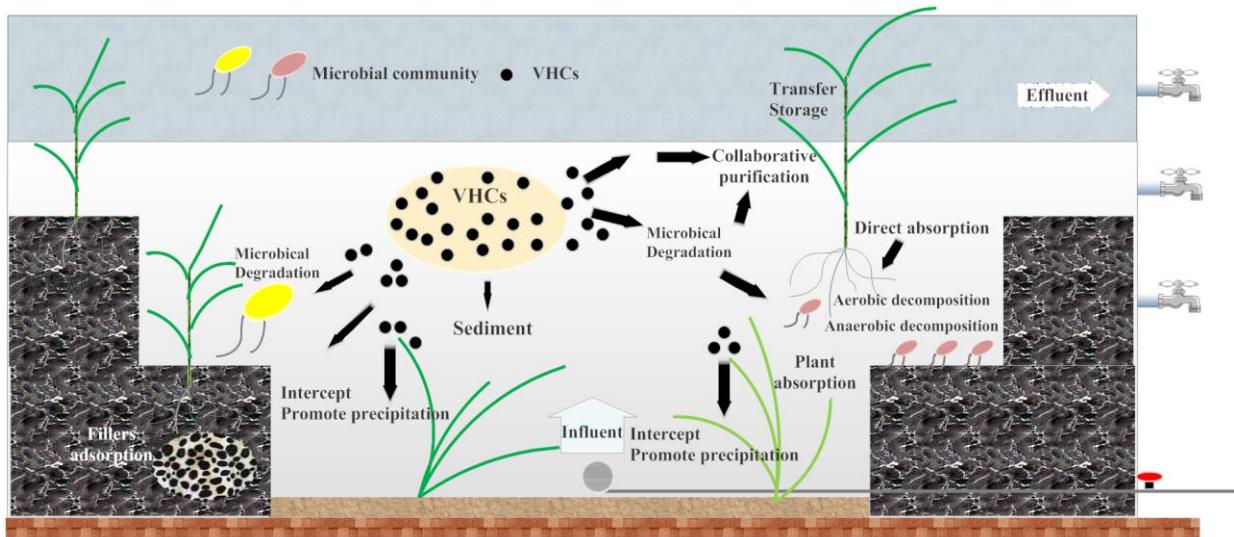
424 Additionally, both the number of *Bacillaceae* and *Cytophagaceae* exhibited that the  
425 opposite trends in the two fillers, which showed the enrichment in the ceramsites and decline  
426 in the zeolites. It appeared be link with the active bacteria were affected by the distribution on  
427 the pollutant concentration. The relative abundance of *Chloroflexaceae* was clearly more  
428 abundant in the later stage on the experiment. It has been widely reported that  
429 *Chloroflexaceae* was a kind of photosynthetic bacteria, which might be live close vicinity to

430 the primary producers and along with the abundance around the roots (Nubels et al., 2002).  
431 We believe the positive change on it connected with the diffusion on the plant roots. In  
432 addition, the number of *Rhodocyclaceae* in the fillers increased, which degraded  
433 trichloroethene in the in-situ restoration reactor and could assist dechlorinate carbon  
434 tetrachloride to form CH<sub>4</sub> and CO<sub>2</sub> (ZivEl et al., 2011). The abundance of family *Skermania*  
435 and *Rhodococcus* decreased slightly, both of the two kinds of microorganisms from the  
436 phylum *Acidobacteria*, which is consistent with some previous studies that *Acidobacteria* was  
437 difficult to use VHCs cometabolism (Maaike et al., 2015).

438 Overall, the results showed that the change of polluted river water led to the change of  
439 microbial community structure. In fact, the microbial communities would make a great  
440 response to adapt to the changes of environment, which was in line with the previous  
441 experimental results, and this situation could be described to two points. For one view, the  
442 microenvironment was regulated by rhizosphere of aquatic macrophytes. For another, the  
443 microorganisms were stimulated by the actual polluted river water, which would enhance  
444 community diversity, especially under a relatively high nutrient status (Navel et al., 2012;  
445 Zhang et al., 2010). But on the whole, the dominant microbial community stay relatively stable  
446 under the diverse hydraulic conditions, which help the degradation and conversion of the  
447 VHCs and was essential to the dechlorinating function of the in-situ reactor.

448 **3.4. Mechanism of VHCs degradation**

449 VHCs degradation needed multiple interactions, particularly in the in-situ restoration  
450 reactor. In-situ ecological restoration reactor was adopted to treat the VHCs in polluted river  
451 water, the processes were shown in the Fig.7.



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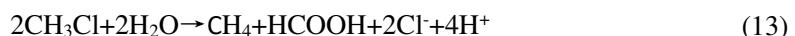
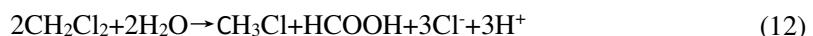
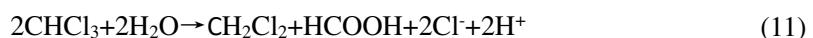
453

Fig.7 Degradation mechanism of VHCs in reactor

454 The material and energy cycle of in-situ remediation reactor ecosystem was the core of  
 455 its function exertion. The degradation reaction process of dehalogenation in plants were  
 456 complex. Under the action of peroxidase, the conversion of VHCs was usually completed by  
 457 substitution reaction. In this experiment, the purification effect of plants depended on the  
 458 process of gas deposition and particulate matter deposition. On the one hand, through the  
 459 exchange of gas deposition between the pores and the liquid surface, the migration trajectory  
 460 of pollutant particles was changed. On the other hand, the effect of hydraulic shock could be  
 461 reduced by macrophyte and the settlement of pollutant particles could be promoted by the  
 462 way of particle sedimentation.

463 The micropores structure of the filler was helpful to adsorb the small molecule pollutant  
 464 particles, which cause the separation of VHCs from volatile organic compounds (Li et al.,  
 465 2020c). In addition, adsorption is a self-energy reduction process. According to the second law of  
 466 thermodynamics, system has a tendency to reduce its energy, which accelerated the pollutant  
 467 molecular was absorbed in the rough surface of zeolite and ceramsites (Long et al., 2018).

468 Actually, the bacteria colonies are generally favoured the anaerobic environment to finish  
469 dichlorination process (Jagnow et al., 1977). In terms of microbial remediation, first of all, the  
470 substitution reaction was the main degradation avenue of VHCs by microorganisms under  
471 anaerobic conditions. The redox potential of VHCs containing halogen was relatively high  
472 (260-570mV), and it was more likely to become the electron acceptor of dehalogenated  
473 microorganisms in anaerobic environment. VHCs compounds got electrons and halogen  
474 atoms were replaced by hydrogen atoms to complete reduction and dehalogenation reaction.  
475 Or the dechlorination is an inducible process depended on the functional bacteria for broken  
476 the C-Cl bonds, formed the C-H bonds and finished biodegradation process (Dong et al.,  
477 2018). The basic processes might be shown in the Eq. (10), Eq. (11), Eq. (12), Eq. (13), Eq.  
478 (14), and Eq. (15):



479 **4. Conclusions**

480 Through outdoor simulation testing, in situ restoration method was applied to degrade  
481 VHCs of polluted river water, which showed that it was feasible. Favorable removal effect of  
482  $\text{CHCl}_3$ ,  $\text{CCl}_4$ ,  $\text{C}_2\text{HCl}_3$  and  $\text{C}_2\text{Cl}_4$  could be obtained through in situ ecological restoration. In  
483 the experiment, orthogonal tests were applied to evaluated different influencing factors. The  
484 findings in this research suggested that when the water depth was 0.4 m, the HRT was 5d and

485 the current velocity was 1 m/s, VHCs removal efficiencies could achieve favorable value, the  
486 removal efficiency of CHCl<sub>3</sub> CCl<sub>4</sub>, C<sub>2</sub>HCl<sub>3</sub> and C<sub>2</sub>Cl<sub>4</sub>.could reach 70.27%, 70.59%67.74% and  
487 81.82%, respectively. Besides, F test results exhibited that HRT and water depth were  
488 significantly related to the removal efficiency of reactor. The RLCs of plants were analyzed  
489 by DIVING-PAM-II, which prominent diversity exists in the adaptation abilities for different  
490 types of plants, but the physiological state of the plants was endangered by VHCs of polluted  
491 river water in all.

492 Moreover, the microbial community structure and diversity of reactor were regulated to  
493 more balance and adapt to the variation characteristics of complex polluted river water  
494 conditions. The microbial community structures of fillers were tested by high-throughput  
495 sequencing, the findings supported that the proportion on the reductive dechlorination  
496 bacterial of VHCs was improved. And there existed a slightly improvement on the relative  
497 abundance of *Rhodocyclaceae*, which might be related to it could promote VHCs degradation.  
498 In general, in situ ecological restoration had a good potential performance on pollutants  
499 removal in polluted river water. The results showed that technology provided a green and low  
500 consumption approach for the maintenance and purification on polluted river water.

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513 **Declaration of interests**

514

515  The authors declare that they have no known competing financial interests or personal  
516 relationships that could have appeared to influence the work reported in this paper.

517

518  The authors declare the following financial interests/personal relationships which may be  
519 considered as potential competing interests:

520

521 **Author contributions:**

522 Jia Wang: Validation, Visualization, Software, Data curation, Writing-Original Draft.

523 Shuangrong Wu: Supervision, Visualization, Writing-Review & Editing..

524 Qi Yang: Conceptualization, Methodology, Software, Investigation, Writing-Review &  
525 Editing.

526 Yonggang Gu: Resources, Formal analysis, Supervision.

527 Peijing Wang: Investigation, Data curation.

528 Zhaoxin Li: Writing-Review & Editing.

529 Lei Li: Software, Investigation.

530 Thank you.

531 Yours sincerely,

532 Yonggang Gu

533 **Ethical approval**

534 Not applicable.

535 **Consent to Participate**

536 I declare here that all authors approve publish this paper.

537 Thank you.

538 Yours sincerely,

539 Yonggang Gu

540     **Competing Tests**

541       The authors declare that they have no known competing financial interests or personal  
542       relationships that could have appeared to influence the work reported in this paper.

543       Thank you.

544

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552       Not applicable.

553

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557       Northern Area (2017ZX07103-001) and the National Science and Technology Major Project  
558       of China (2015ZX07406005).

559     **References**

560       Albers, C. N., Ellegaard-Jensen, L., Harder, C. B., Knudsen, B. E., Ekelund, F., Aamand, J., 2015.

561       Groundwater Chemistry Determines the Prokaryotic Community Structure of Waterworks  
562       Sand Filters[J]. *Environmental Science & Technology*, 49(2):839-46.

563       Anwar Sadat, M., Guan, Y., Zhang, D., Shao, G., Cheng, X., Yang, Y. 2020. The associations between  
564       river health and water resources management lead to the assessment of river state. *Ecological*

565       *Indicators*, 109.

- 566 Brestic, M., Zivcak, M. 2013. PSII Fluorescence Techniques for Measurement of Drought and High
- 567 Temperature Stress Signal in Crop Plants: Protocols and Applications. in: *Molecular Stress*
- 568 *Physiology of Plants*, pp. 87-131.
- 569 Cappelletti, M., Presentato, A., Piacenza, E., Firrincieli, A., Turner, R.J., Zannoni, D. 2020.
- 570 Biotechnology of Rhodococcus for the production of valuable compounds. *Appl Microbiol*
- 571 *Biotechnol*, 104(20), 8567-8594.
- 572 Chouari, R., Le Paslier, D., Daegelen, P., Ginestet, P., Weissenbach, J., Sghir, A. 2005. Novel
- 573 predominant archaeal and bacterial groups revealed by molecular analysis of an anaerobic
- 574 sludge digester. *Environmental Microbiology*, 7(8), 1104-1115.
- 575 Dong, S., Feng, C., Cui, W., Chen, N., Liu, Y., Liu, T., 2018. Anaerobic Bioremediation Performance
- 576 and Indigenous Microbial Communities in Treatment of Trichloroethylene/Nitrate-
- 577 Contaminated Groundwater[J]. *Environmental Engineering Science*, 35(4):311-322
- 578 Fatahi-Bafghi, M. 2019. Antibiotic resistance genes in the Actinobacteria phylum. *Eur J Clin Microbiol*
- 579 *Infect Dis*, 38(9), 1599-1624.
- 580 Giripunje, M. D., Fulke, A. B., Meshram, P. U., 2015. Remediation Techniques for Heavy-Metals
- 581 Contamination in Lakes: A Mini-Review. *CLEAN - Soil, Air, Water*, 43(9), 1350-1354.
- 582 Guo, C., Cui, Y., Dong, B., Luo, Y., Liu, F., Zhao, S., Wu, H. 2017. Test study of the optimal design for
- 583 hydraulic performance and treatment performance of free water surface flow constructed
- 584 wetland. *Bioresour Technol*, 238, 461-471.
- 585 Guo, Z., Kang, Y., Hu, Z., Liang, S., Xie, H., Ngo, H.H., Zhang, J. 2020. Removal pathways of
- 586 benzofluoranthene in a constructed wetland amended with metallic ions embedded carbon.
- 587 *Bioresour Technol*, 311, 123481.

- 588 Hassan, S.S., Anjum, K., Abbas, S.Q., Akhter, N., Shagufta, B.I., Shah, S.A., Tasneem, U. 2017.
- 589 Emerging biopharmaceuticals from marine actinobacteria. *Environ Toxicol Pharmacol*, 49, 34-
- 590 47.
- 591 Hirsch, R. M., Moyer, D., Archfield, S. A., 2010. Weighted Regressions on Time, Discharge, and
- 592 Season (WRTDS), with an Application to Chesapeake Bay River Inputs. *Journal of The*
- 593 *American Water Resources Association*, 46(5), 857-880.
- 594 Hou, X., Liu, G., Jiang, G., 2018. Metabolism of typical halogenated organic pollutants in plants.
- 595 *Scientia Sinica Chimica*, 48(10), 1236-1246.
- 596 Huang, X., Yang, X., Zhu, J., Yu, J. 2020. Microbial interspecific interaction and nitrogen metabolism
- 597 pathway for the treatment of municipal wastewater by iron carbon based constructed wetland.
- 598 *Bioresour Technol*, 315, 123814.
- 599 Jagnow, G., Haider, K., Ellwardt, P. C..1977. Anaerobic dechlorination and degradation of
- 600 hexachlorocyclohexane isomers by anaerobic and facultative anaerobic bacteria[J]. Archives
- 601 of Microbiology, 115(3):285-292.
- 602 Jiang, Y., Li, Y., Zhang, Y., Zhang, X. 2017. Effects of HRT on the efficiency of denitrification and
- 603 carbon source release in constructed wetland filled with bark. *Water Sci Technol*, 75(12),
- 604 2908-2915.
- 605 Kobayashi, Y., Ralph, T.J., Sharma, P., Mitrovic, S.M.J.M., Research, F. 2020. Influence of historical
- 606 inundation frequency on soil microbes (Cyanobacteria, Proteobacteria, Actinobacteria) in
- 607 semi-arid floodplain wetlands. 71.
- 608 Krall, J.P.J.P.P. 2010. Relationship between photosystem II activity and CO<sub>2</sub> fixation in leaves. 86(1),
- 609 180-187.

- 610 Ladislas, S., El-Mufleh, A., Gérante, C., Chazarenc, F., Andrès, Y., Béchet, B., 2011. Potential of  
611 Aquatic Macrophytes as Bioindicators of Heavy Metal Pollution in Urban Stormwater Runoff.  
612 *Water, Air, & Soil Pollution*, 223(2), 877-888.
- 613 Li, D., Zheng, B., Liu, Y., Chu, Z., He, Y., Huang, M. 2018. Use of multiple water surface flow  
614 constructed wetlands for non-point source water pollution control. *Appl Microbiol Biotechnol*,  
615 102(13), 5355-5368.
- 616 Li, X., Li, Y., Li, Y., Wu, J. 2020a. Myriophyllum elatinoides growth and rhizosphere bacterial  
617 community structure under different nitrogen concentrations in swine wastewater. *Bioresour*  
618 *Technol*, 301, 122776.
- 619 Li, X., Wu, S., Yang, C., Zeng, G. 2020b. Microalgal and duckweed based constructed wetlands for  
620 swine wastewater treatment: A review. *Bioresour Technol*, 318, 123858.
- 621 Li, X., Zhang, L., Yang, Z., Wang, P., Yan, Y., Ran, J. 2020c. Adsorption materials for volatile organic  
622 compounds (VOCs) and the key factors for VOCs adsorption process: A review. *Separation*  
623 *and Purification Technology*, 235.
- 624 Liu, J.J., Dong, B., Guo, C.Q., Liu, F.P., Brown, L., Li, Q. 2016. Variations of effective volume and  
625 removal rate under different water levels of constructed wetland. *Ecological Engineering*, 95,  
626 652-664.
- 627 Long, Y., Wu, S., Xiao, Y., Cui, P., Zhou, H. 2018. VOCs reduction and inhibition mechanisms of using  
628 active carbon filler in bituminous materials. *Journal of Cleaner Production*, 181, 784-793.
- 629 Lu, G., Wang, B., Zhang, C., Li, S., Wen, J., Lu, G., Zhu, C., Zhou, Y., 2018. Heavy metals  
630 contamination and accumulation in submerged macrophytes in an urban river in China. *Int J*  
631 *Phytoremediation*, 20(8), 839-846.

- 632 Maaike, V. A., Van, O., Gera, H., Hundscheid, M., Runia, W. T., Hordijk, C. A., Wietse, D. B., 2015.
- 633           Legacy effects of anaerobic soil disinfestation on soil bacterial community composition and
- 634           production of pathogen-suppressing volatiles[J]. *Frontiers in Microbiology*, 6:701-716.
- 635 Man, Y., Wang, J., Tam, N.F., Wan, X., Huang, W., Zheng, Y., Tang, J., Tao, R., Yang, Y. 2020.
- 636           Responses of rhizosphere and bulk substrate microbiome to wastewater-borne sulfonamides in
- 637           constructed wetlands with different plant species. *Sci Total Environ*, 706, 135955.
- 638 Mandal, B., Das, S.C., Mandal, L.N.J.P., Soil. 1992. Effect of growth and subsequent decomposition of
- 639           cyanobacteria on the transformation of phosphorus in submerged soils. 143(2), 289-297.
- 640 Mathon, B., Coquery, M., Miege, C., Vandycke, A., Choubert, J.M. 2019. Influence of water depth and
- 641           season on the photodegradation of micropollutants in a free-water surface constructed wetland
- 642           receiving treated wastewater. *Chemosphere*, 235, 260-270.
- 643 Milobedzka, A., Muszynski, A. 2015. Population dynamics of filamentous bacteria identified in Polish
- 644           full-scale wastewater treatment plants with nutrients removal. *Water Sci Technol*, 71(5), 675-
- 645           84.
- 646 Mun, H., Townley, H.E. 2021. Nanoencapsulation of Plant Volatile Organic Compounds to Improve
- 647           Their Biological Activities. *Planta Med*, 87(3), 236-251.
- 648 Navarro-Ortega, A., Tauler, R., Lacorte, S., Barceló, D. 2010. Occurrence and transport of PAHs,
- 649           pesticides and alkylphenols in sediment samples along the Ebro River Basin. *Journal of*
- 650           *Hydrology*, 383(1-2), 5-17.
- 651 Navel, S., Mermillod-Blondin, F., Montuelle, B., Chauvet, E., Marmonier, P. 2012. Sedimentary
- 652           context controls the influence of ecosystem engineering by bioturbators on microbial
- 653           processes in river sediments. *Oikos*, 121(7), 1134-1144.

- 654 Nilsson, C., Polvi, L.E., Gardeström, J., Hasselquist, E.M., Lind, L., Sarneel, J.M. 2015. Riparian and  
655 in-stream restoration of boreal streams and rivers: success or failure. *Ecohydrology*, 8(5), 753-  
656 764.
- 657 Nubel U., Bateson M. M., Vandieken V., Wieland A., Kuhl K., Ward D. M., 2002. Microscopic  
658 Examination of Distribution and Phenotypic Properties of Phylogenetically Diverse  
659 Chloroflexaceae-Related Bacteria in Hot Spring Microbial Mats[J]. *Applied & Environmental*  
660 *Microbiology*, 68(9):4593-603.
- 661 Özgencil, İ.K., Beklioğlu, M., Özkan, K., Tavşanoğlu, Ç., Fattorini, N., 2020. Changes in functional  
662 composition and diversity of waterbirds: The roles of water level and submerged macrophytes.  
663 *Freshwater Biology*, 65(11), 1-13.
- 664 Pan, B., Yuan, J., Zhang, X., Wang, Z., Chen, J., Lu, J., Yang, W., Li, Z., Zhao, N., Xu, M. 2016. A  
665 review of ecological restoration techniques in fluvial rivers. *International Journal of Sediment*  
666 *Research*, 31(2), 110-119.
- 667 Frankenbach, S., SeroDio, J., 2017. One pulse, one light curve: Fast characterization of the light  
668 response of microphytobenthos biofilms using chlorophyll fluorescence. *Limnology and*  
669 *Oceanography: Methods*.
- 670 Rugner, H., Schwientek, M., Milacic, R., Zuliani, T., Vidmar, J., Paunovic, M., Laschou, S., Kalogianni,  
671 E., Skoulikidis, N.T., Diamantini, E., Majone, B., Bellin, A., Chiogna, G., Martinez, E., Lopez  
672 de Alda, M., Diaz-Cruz, M.S., Grathwohl, P. 2019. Particle bound pollutants in rivers: Results  
673 from suspended sediment sampling in Globaqua River Basins. *Sci Total Environ*, 647, 645-  
674 652.
- 675 San Bautista, A., Calatayud, A., Nebauer, S.G., Pascual, B., Maroto, J.V., López-Galarza, S. 2011.

- 676 Effects of simple and double grafting melon plants on mineral absorption, photosynthesis,  
677 biomass and yield. *Scientia Horticulturae*, 130(3), 575-580.
- 678 Song, C., Liu, X., Song, Y., Liu, R., Gao, H., Han, L., Peng, J. 2017. Key blackening and stinking  
679 pollutants in Dongsha River of Beijing: Spatial distribution and source identification. *Journal*  
680 *of Environmental Management*, 200, 335-346.
- 681 Su, T.-M., Yang, S.-C., Shih, S.-S., Lee, H.-Y. 2009. Optimal design for hydraulic efficiency  
682 performance of free-water-surface constructed wetlands. *Ecological Engineering*, 35(8), 1200-  
683 1207.
- 684 Takahashi, S., Badger, M.R. 2011. Photoprotection in plants: a new light on photosystem II damage.  
685 *Trends Plant Sci*, 16(1), 53-60.
- 686 Tang, S., Liao, Y., Xu, Y., Dang, Z., Zhu, X., Ji, G. 2020. Microbial coupling mechanisms of nitrogen  
687 removal in constructed wetlands: A review. *Bioresour Technol*, 314, 123759.
- 688 Wang, H., Wang, J., Bo, G., Wu, S., Luo, L. 2020. Degradation of pollutants in polluted river water  
689 using Ti/IrO<sub>2</sub>-Ta<sub>2</sub>O<sub>5</sub> coating electrode and evaluation of electrode characteristics. *Journal of*  
690 *Cleaner Production*, 273.
- 691 Wang, J., Gu, Y., Wang, H., Li, Z. 2021. Investigation on the treatment effect of slope wetland on  
692 pollutants under different hydraulic retention times. *Environ Sci Pollut Res Int*, 28(8), 9107-  
693 9119.
- 694 Wang, Q., Xie, H., Ngo, H.H., Guo, W., Zhang, J., Liu, C., Liang, S., Hu, Z., Yang, Z., Zhao, C. 2016.  
695 Microbial abundance and community in subsurface flow constructed wetland microcosms:  
696 role of plant presence. *Environ Sci Pollut Res Int*, 23(5), 4036-45.
- 697 Wang, X., Tian, Y., Liu, H., Zhao, X., Peng, S. 2019. Optimizing the performance of organics and

- 698 nutrient removal in constructed wetland-microbial fuel cell systems. *Sci Total Environ*, 653,  
699 860-871.
- 700 Wang, X., Wang, S., Peng, G., S. w. katz, D., Ling, H. 2017. Ecological restoration for river ecosystems:  
701 comparing the huangpu river in shanghai and the hudson river in new york. *Ecosystem Health  
702 and Sustainability*, 1(7), 1-14.
- 703 White, A.J., Critchley, C.J.P.R. 1999. Rapid light curves: A new fluorescence method to assess the state  
704 of the photosynthetic apparatus. 59(1), 63-72.
- 705 Wolinska, A., FraC, M., Oszust, K., Szafranek-Nakonieczna, A., Zielenkiewicz, U., Stępniewska, Z.,  
706 2017. Microbial biodiversity of meadows under different modes of land use: catabolic and  
707 genetic fingerprinting. *World Journal of Microbiology & Biotechnology*, 33(8), 154-169.
- 708 Xiong, C., Tam, N.F., Dai, Y., Zhang, X., Li, R., Zheng, Y., Wang, L., Yang, Y. 2020. Enhanced  
709 performance of pilot-scale hybrid constructed wetlands with A/O reactor in raw domestic  
710 sewage treatment. *J Environ Manage*, 258, 110026.
- 711 Yaghoubian, Y., Siadat, S.A., Moradi Telavat, M.R., Pirdashti, H. 2016. Quantify the response of  
712 purslane plant growth, photosynthesis pigments and photosystem II photochemistry to  
713 cadmium concentration gradients in the soil. *Russian Journal of Plant Physiology*, 63(1), 77-  
714 84.
- 715 Yang, J., Li, Q., An, Y., Zhang, M., Du, J., Chen, C., Zhao, R., Zhao, D., An, S. 2020. The improvement  
716 of pollutant removal efficiency in saturated vertical flow constructed wetlands by tubifex  
717 tubifex. *Bioresour Technol*, 318, 124202.
- 718 Yuan, K., Chen, B., Qing, Q., Zou, S., Wang, X., Luan, T. 2017. Polycyclic aromatic hydrocarbons  
719 (PAHs) enrich their degrading genera and genes in human-impacted aquatic environments.

- 720                   *Environ Pollut*, 230, 936-944.
- 721       Zhang, C.B., Wang, J., Liu, W.L., Zhu, S.X., Liu, D., Chang, S.X., Chang, J., Ge, Y. 2010. Effects of  
722                   plant diversity on nutrient retention and enzyme activities in a full-scale constructed wetland.
- 723                   *Bioresour Technol*, 101(6), 1686-92.
- 724       Zhang, L., Liu, H., Peng, Y., Zhang, Y., Sun, Q. 2020a. Characteristics and significance of dissolved  
725                   organic matter in river sediments of extremely water-deficient basins: A Beiyun River case  
726                   study. *Journal of Cleaner Production*, 277.
- 727       Zhang, P., Zhang, Z., Li, B., Zhang, H., Hu, J., Zhao, J. 2020b. Photosynthetic rate prediction model of  
728                   newborn leaves verified by core fluorescence parameters. *Sci Rep*, 10(1), 3013.
- 729       Zheng, Y., Yang, D., Dzakpasu, M., Yang, Q., Liu, Y., Zhang, H., Zhang, L., Wang, X.C., Zhao, Y. 2020.  
730                   Effects of plants competition on critical bacteria selection and pollutants dynamics in a long-  
731                   term polyculture constructed wetland. *Bioresource Technology*, 316.
- 732       Zhang, X., Qin, H., Wang, H., Wan, A., Liu, G., 2018. Effects of water level fluctuations on root  
733                   architectural and morphological traits of plants in lakeshore areas of three subtropical  
734                   floodplain lakes in China. *Environ Sci Pollut Res Int*, 25(34), 34583-34594.
- 735       Ziv-El M., Delgado A. G., Yao Y., Kang D.W., Nelson K. G., Halden R.U., Krajmalnik-Brown R., 2011.  
736                   Development and characterization of DehaloR<sup>2</sup>, a novel anaerobic microbial consortium  
737                   performing rapid dechlorination of TCE to ethene[J]. *Applied Microbiology & Biotechnology*,  
738                   92(5):1063-1071.
- 739       Zou, L., Luo, Y., Hooper, M., Hu, E. 2006. Removal of VOCs by photocatalysis process using  
740                   adsorption enhanced TiO<sub>2</sub>-SiO<sub>2</sub> catalyst. *Chemical Engineering and Processing: Process  
741                   Intensification*, 45(11), 959-964.

