

# Estrogen receptor $\alpha$ phosphorylated at Ser216 confers inflammatory function to mouse microglia

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## Research

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## Abstract

Background Estrogen has been suggested to regulate anti-inflammatory signaling in brain microglia through estrogen receptor  $\alpha$  (ER $\alpha$ ), the only resident immune cells of the brain. The mechanism of how ER $\alpha$  regulates is not well understood. Previously, ER $\alpha$  is phosphorylated at Ser216 in mouse neutrophils, regulating their infiltration into the uterus. Therefore, ER $\alpha$  has now been examined as to its phosphorylation in microglia to regulate their inflammatory functions. Methods An antibody against an anti-phospho-S216 peptide of ER $\alpha$  (aP-S216) was used for double immunofluorescence staining to detect ER $\alpha$  in cultured microglia. A knock-in (KI) mouse line bearing the phosphorylation-blocked ER $\alpha$  mutation S216A (ER $\alpha$  KI) was generated to examine whether this phosphorylation regulate immune functions of microglia. Results Phosphorylated ER $\alpha$  at Ser216 was present in microglia but not astrocytes. Staining with an anti-Iba-1 antibody showed that microglia activation was augmented in substantial nigra of ER $\alpha$  KI brains. Lipopolysaccharide (LPS) treatments aggravated microglia activation in ER $\alpha$  KI brains, pro-inflammatory cytokines were increased while anti-inflammatory cytokines were decreased at mRNA and protein levels in whole brain extracts. These increases and decreases of cytokine proteins were also observed in LPS-treated microglia cultured from brains of ER $\alpha$  KI neonates. FACS analysis revealed that ER $\alpha$  KI mutation increased number of IL-6 producing microglia and apoptosis. ER $\alpha$  KI mice decreased motor connection ability in Rotarod tests. Conclusions Blocking of Ser216 phosphorylation aggravated microglia activation and inflammation of mouse brain, thus confirming that phosphorylated ER $\alpha$  exerts anti-inflammatory functions. ER $\alpha$  KI mice enable us to further investigate the mechanism by which phosphorylated ER $\alpha$  regulates brain immunity and inflammation.

## Background

Estrogen and estrogen receptor  $\alpha$  (ER $\alpha$ ) are known to regulate anti-inflammatory signaling in the brain [1] and are directly involving in the pathogenesis of neurodegeneration and other inflammation-related brain diseases [2]. Microglia are the resident macrophages in the brain, responsible for the control of neuroinflammation [3]. Since ER $\alpha$  is ubiquitously expressed in the brain, the signaling that specifically regulates ER $\alpha$  in microglia remains uninvestigated. Moreover, a proper animal model that enables us to perform targeted examinations to microglia in brain does not currently exist. Here we have now found that ER $\alpha$  is phosphorylated at Ser216 in microglia and generated an ER $\alpha$ S216A knock-in (*Esr1<sup>S216A</sup>*) mouse line to investigate whether this phosphorylation enables ER $\alpha$  to regulate inflammation of microglia. ER $\alpha$ 's Ser216 (Ser212 in human ER $\alpha$ ) is present within the DNA binding domain (DBD) and conserved a phosphorylation motif in 41 out of 46 total mouse nuclear receptors (Additional file 1: Figure S1) as well as in the corresponding human nuclear receptors [4]. These extremely high cross-species conservations strongly suggest critical regulatory functions this motif may plays for nuclear receptor actions. In fact, In addition to Ser216 of ER $\alpha$ , the corresponding residues of four other nuclear receptors have been reported to be phosphorylated in mouse tissues *in vivo* and to confer specific function to them, which include Thr38, Ser100, Ser154 and Thr167 of constitute active/androstane receptor (CAR), retinoid-related orphan receptors  $\alpha$  (ROR $\alpha$ ), farnesoid X receptor (FXR) and retinoid X receptor  $\alpha$  (RXR $\alpha$ ), respectively [4, 5, 6, 7, 8, 9]. For example, phosphorylation of Thr38 presses the constitutive activity of CAR, providing it with the response ability to its activator [6, 7]. RXR $\alpha$  was found to be phosphorylated at Thr167 in mouse adipose tissues in response to fasting. We generated RXR $\alpha$  T167A knock-in mouse and demonstrated that this phosphorylation regulates blood glucose levels by altering energy metabolism in adipose tissues [9]. These observations confirmed that phosphorylation of this conserved motif can be a common regulatory factor of nuclear receptors. Thus, phosphorylation of Ser216 presents an experimental basis to examine ER $\alpha$  in this context.

This manuscript analyzed inflammation of microglia of the ER $\alpha$  KI brain in comparison with that of ER $\alpha$  WT brains. Microglia in the brain and, subsequently, glia cells cultured from the brains of 2-day-old neonates were subjected to investigations by using immunohistochemistry, real-time PCR, ELISA, cytokine arrays, FACS and Western blots. With experimental observations obtained we will discuss the anti-inflammatory and anti-apoptotic functions of phosphorylated ER $\alpha$  in microglia. This ER $\alpha$  KI (*Esr1<sup>S216A</sup>*) mouse is the first KI at a potential phosphorylation motif that shows a physiological phenotype and can be used as an animal model for the study of physiological functions of phosphorylated ER $\alpha$  and their molecular mechanism as well as for drug discovery and development targeting microglia.

## Results

### ER $\alpha$ phosphorylated at Ser216 in microglia

Mixed glia cells were isolated from brains of 2-day-old neonates and cultured for 2 weeks to mature microglia prior to immunofluorescence staining. For microglia, these glia cells were double stained with an anti-ER $\alpha$  or P-S216 peptide antibody (in green) with an anti-Iba-1 antibody (in red) (Fig. 1a). These microglia stained appeared to be immature cells. Both ER $\alpha$  antibodies co-stained microglia. Staining of ER $\alpha$  by an anti-P-S216 peptide antibody was further confirmed with enriched mature microglia from glia cultures (Fig. 1b). For astrocytes, glia cells were co-stained by an anti-ER $\alpha$  or P-S216 peptide antibody (in green) with an anti-GFAP antibody (in red) (Fig. 1c). Anti-ER $\alpha$  antibody strongly stained astrocytes, while the P-S216 peptide antibody barely stained these cells. Obtained observations indicate that ER $\alpha$  is phosphorylated at serine 216 in microglia and were expressed in the nucleus as well as in the extra-nuclear region. ER $\alpha$  KO mice were further examined ER $\alpha$  in brain microglia. First, immunostaining was performed to show the absence of ER $\alpha$  in microglia (Fig. 1d). Whole extracts from enriched microglia were subjected to Western blot analysis. Both an anti-ER $\alpha$  and P-S216 antibodies detected ER $\alpha$  band only in the extracts from ER $\alpha$  WT microglia (Fig. 1e).

### ER $\alpha$ S216A KI (*Esr1<sup>S216A</sup>*) mice

Utilizing ACN cassette, a single mutation of serine 216 to alanine was introduced in the *Esr1* gene (Fig. 2a). The mutation was verified by Southern hybridization (Fig. 2b) and PCR amplification (Fig. 2c). ER $\alpha$  mRNA and protein were equally expressed in the uterus of ER $\alpha$  WT and ER $\alpha$  KI mice (Figs. 2d and 2e) and the sequences of cDNAs confirmed the mutation (Fig. 2f). Moreover, co-immunostaining of glia cells with an anti-ER $\alpha$  and an anti-Iba-1 antibodies

demonstrated the presence of ER $\alpha$  in microglia prepared from ER $\alpha$  KI as observed as with ER $\alpha$  WT mice (Fig. 2g). ER $\alpha$  KI mice were fertile; no obvious reproductive abnormality was observed in either ER $\alpha$  KI females or males. However, both male and female ER $\alpha$  KI mice developed obesity. At 6-month-old mice ER $\alpha$  KI mice were about 40% over-weight compared to WT mice (Fig. 2h).

### Microglia in adult ER $\alpha$ KI brains

Although microglia are present throughout mouse brain, its distribution is much denser in specific regions such as olfactory telencephalon and substantia nigra (SN) reticulata [10]. Brain sections were prepared from saline- or LPS-treated adult ER $\alpha$  WT and KI males and stained with an anti-Iba-1 antibody (Fig. 3a). Intensities of Iba-1 staining are known to increase as microglia become more activated [11]. Those intensities were higher in saline-treated ER $\alpha$  KI compared with those in ER $\alpha$  WT. Moreover, increases in the staining intensity by LPS treatment were 20% higher in ER $\alpha$  KI than in ER $\alpha$  WT mice (Fig. 3b). In addition, microglia developed morphological changes such as hypertrophic cells with pseudopodia in ER $\alpha$  KI mice. Thus, blocking ER $\alpha$  phosphorylation at Ser216 aggravated microglia to increase both basal and LPS-induced activations. Brain extracts from adult ER $\alpha$  WT and KI mice treated with saline or LPS were subjected to RT-PCR analysis. LPS-induced mRNA levels of pro-inflammatory cytokines such as TNF- $\alpha$ , IL-1 $\alpha$  and IL-1 $\beta$  were higher in ER $\alpha$  KI brains compared to ER $\alpha$  WT brains (Fig. 4a). Conversely, LPS-induced expression of an anti-inflammatory IL-10 mRNA slightly diminished LPS in ER $\alpha$  KI brains. In addition, LPS induction of mRNAs for two enzymes, Cox-2 and iNOS, were higher in ER $\alpha$  KI brains; in particular, iNOS mRNA was induced only in KI brains. Subsequently, cytokine arrays were utilized to examine expressions of cytokines at their protein levels (Fig. 4b). Only a few cytokines were detected in the extracts of saline-treated brains and there was no difference in these expressions between ER $\alpha$  WT and KI brains. LPS treatment induced numerous cytokines and chemokines in both ER $\alpha$  WT and KI brains (Fig. 4c). Semi-quantitation of these expressions revealed that the degrees of these inductions were much higher in ER $\alpha$  KI than in WT brain (Fig. 4c). As observed with their mRNAs, protein levels of IL-6, IL-1 $\alpha$  and IL-1 $\beta$  also increased. However, no difference was observed in a TNF- $\alpha$  protein expression between WT and KI. In addition to cytokines, LPS-induced levels of chemokines and of a metalloprotease inhibitor TIMP-1 were higher in ER $\alpha$  KI than in WT brains. An integrin receptor ICAM-1 (also known as CD54) was also increased more in ER $\alpha$  KI brains after LPS treatment (Figs. 4b and 4c). These observations indicate that ER $\alpha$  KI microglia are more sensitive to LPS-elicited immune responses.

### Microglia in cultures

Mixed glia cells were prepared from brains of 2-day-old neonates, cultured for 2 weeks so that microglia matured, and treated with LPS, from which supernatants were collected for ELISA (Fig. 5a). The increases of pro-inflammatory factors IL-6 and PGE2 were about 50% higher in KI microglia after 24 hr treatment. Conversely, an anti-inflammatory cytokine IL-10 was greatly repressed in KI microglia after 48 hr treatment (Fig. 5a). Similar results were observed when examining intracellular levels of IL-6 by fluorescence-activated cell sorting (FACS). Briefly, mixed glia cells prepared from 2-day-old neonates were treated with PBS or LPS for 2 or 24 hrs. Male and female neonates were determined by different distances between anus and urinary opening. The mixed glial culture was fixed and permeabilized to allow for antibody entry. Mixed glia cultures were stained for the microglia marker, F4/80, and F4/80<sup>+</sup> microglia were analyzed for intracellular levels of IL-6. LPS-induced expression of IL-6 was approximately 35% higher in ER $\alpha$  KI than in ER $\alpha$  WT microglia (Fig. 5b). FACS was also utilized to analyze apoptosis in mixed glia cultures. Annexin V binds extracellular phosphatidylserine, which is actively exposed during apoptosis. After LPS treatment, we also observed about 30% increase in apoptosis in ER $\alpha$  KI microglia, as determined by Annexin V positivity (Fig. 5c). Noticeably, there are no sex differences in the responses. Taken together, these results suggested that ER $\alpha$  KI microglia became increasingly inflammatory as well as apoptotic after LPS treatment.

### Motor connection ability

Substantia nigra is an area of the brain that controls movement, microglia in which are associated with this function. Given the finding that inflammation was aggravated in microglia of ER $\alpha$  KI brains, Rotarod tests were employed to examine whether motor connection was affected. It was found that average latencies were 206  $\pm$  57 sec with 3-month old WT mice (Fig. 6). The corresponding KI mice significantly decreased this latency to 138  $\pm$  17 sec, impairing motor connection. These degrees of latency were continuously observed with 6-month old WT mice, 217  $\pm$  59 and 146  $\pm$  59 with WT and KI mice, respectively (Fig. 6). Thus, the microglia expression of ER $\alpha$  S216A mutant appeared to deteriorate motor connection of mice.

## Discussion

ER $\alpha$  is phosphorylated at Ser216 in microglia, only the immune cells in the brain. Microglia are the second immune cells which are found to express phosphorylated ER $\alpha$ . Neutrophils infiltrating the mouse uterus were first found to express phosphorylated ER $\alpha$  [5]. Studies with ER $\alpha$  S216A KI (*Esr1<sup>S216A</sup>*) mice show the anti-inflammatory and anti-apoptotic functions of phosphorylated ER $\alpha$  in microglia. Microglia in substantia nigra are activated in the brain of ER $\alpha$  S216A KI mice. Inflammatory activation of these microglia has been associated with neurodegenerative diseases such as Parkinson [12, 13]. ER $\alpha$  S216A KI mice and microglia can be excellent experimental models for us to further implicate phosphorylated ER $\alpha$  in inflammation-related neurodegenerative diseases and investigate their molecular and cellular mechanisms.

Immunological study reported that M1/M2 polarization of macrophages plays an important role in controlling the balance between promotion and suppression in inflammation. Microglia protect brain cells from various stresses and insults by accelerating pro-inflammation to eliminate damaged cells (M1 microglia function) and, subsequently, decorrelating it to recover the brain from injury (M2 microglia function) [14]. Phosphorylated ER $\alpha$  is found to regulate both M1 and M2 functions, as indicated by the fact that an early expression of pro-inflammatory factors (e.g. IL-6 and PGE2) is stimulated and subsequent

expression of anti-inflammatory factors (e.g. IL-10) is attenuated in LPS-treated ER $\alpha$  S216A KI mice. In other words, microglia in which ER $\alpha$  cannot be phosphorylated may have a strong function of M1 and may be less polarized to M2. Apoptosis is augmented in LPS-treated microglia from ER $\alpha$  S216A KI brains. In both the brain and cultured microglia, not all factors are regulated equally. However, phenotypes obtained by cultured microglia would reflect what should be observed in the brain. In addition, phosphorylated ER $\alpha$  may also have the potential to regulate microglia migration through the expression of cell adhesion molecules such as ICAM-1 and metalloprotease inhibitor TIMP-1. These patterns of regulations implicate phosphorylated ER $\alpha$  in the immune severance regulation to protect brains cells from injuries [15]. Rotarod tests found that ER $\alpha$  KI mice decreased motor ability. It remains to be seen if this correlate with decrease numbers of dopaminergic neurons in the areas of brain's substantia nigra. In addition to neurodegenerative diseases, recently, macrophage-elicited PGE2-EP2-NF- $\kappa$ B signaling in brains was linked to chronic inflammation, cerebral aneurisms and subarachnoid hemorrhage with NF- $\kappa$ B suggested to be a therapeutic target of aneurisms [16]. However, phosphorylated ER $\alpha$  may be a better target, possibly controlling inflammation but not the other signals in the brain.

The stimulated inflammation observed with ER $\alpha$  S216A KI brains closely resembles previous reports that microglia are activated by LPS in global ER $\alpha$  KO mice [1], confirming the role of ER $\alpha$  in the inflammation. It was reported that estrogen repressed LPS-induced inflammation and apparently suppressed in isolated rat microglia from normal rat [17], suggesting the possibility that what this ER $\alpha$ -mediated regulation was an estrogen action. However, our repeated experiments with cultured microglia did not find any effects of estrogen on their activities (unpublished). These differences between the two experiments are not reconciled now and whether estrogen directly regulates phosphorylated ER $\alpha$  in microglia remains further investigated in the future. Functions of ER $\alpha$  are primarily understood by estrogen actions with a gender-dependent manner predominately observed in females [18, 19, 20]. An alternative is the possibility that estrogen indirectly regulates phosphorylated ER $\alpha$  through astrocytes. A cross-talk between astrocytes and microglia is well known to regulate inflammation in the brain [21, 22]. Moreover, a recent report find that microglia play the determining role in sexual differentiation of the brain and estrogen may regulate microglia through astrocytes [2, 23]. Although it was not definitive, ER $\alpha$  appears to be not phosphorylated at Ser216 (Fig. 1), suggesting that an estrogen's direct target may be ER $\alpha$  in astrocytes.

Microglia are critically involved in the development of various neurodegenerative diseases including Alzheimer's, Parkinson's and Huntington's diseases [12]. In fact, LPS-induced activation of microglia in the substantia nigra was associated with development of Parkinson disease [13, 24], although mice are known not to develop this disease [25]. Estrogen signaling has been investigated as a therapeutic target for developing drugs against neurotoxicity and injury as well as neurodegenerative diseases [21]. Phosphorylated ER $\alpha$  in microglia can be a more direct target of drug development. If chemicals that specifically bind and activate or inactivate phosphorylated ER $\alpha$  are identified, they may be effective in microglia but not the other cells in which ER $\alpha$  is not phosphorylated. Our previous study showed that human ER $\alpha$  S212D mutant activated ERE-reporter gene as observed with ER $\alpha$  WT and ER $\alpha$  S212A in Huh-7 cells [26], suggesting that cell-based reporter assays can be established for high-throughput assays to identify ligands specific to phosphorylated ER $\alpha$ , using cells such as Huh-7 and/or mouse microglia-derived BV-2 cells. In addition to the brain, other tissues such as skin (Langerhans cells in) and liver (Kupffer cells) house resident macrophages. The presence of ER $\alpha$  was previously reported in these resident macrophages [27, 28], as well as bone marrow-derived and peritoneal macrophages [29]. ER $\alpha$  S216A KI mice can be utilized to examine if their inflammatory responses are altered. In fact, Kupffer cells isolated from ER $\alpha$  S216A KI mice expressed cytokines differently from those from normal mice (unpublished). Once confirmed, anti-inflammatory drugs that target skin and/or liver may also be developed.

## Conclusions

ER $\alpha$  S216A KI mice are an excellent animal model for us to further investigate the inflammatory regulation of phosphorylated ER $\alpha$  and its molecular mechanism in microglia and implicate microglia in inflammation-related brain diseases. Current findings with this mouse provide experimental bases for us to further extend functional study of phosphorylated ER $\alpha$  into various directions which include generation of microglia-specific aberration of Ser216 phosphorylation. Once ER $\alpha$  is confirmed in human microglia, therapeutic drugs which selectively target phosphorylated ER $\alpha$  over non-phosphorylated ER $\alpha$ . Together with our investigations using ER $\alpha$  S216A KI mice and RXR $\alpha$  T167A KI mice [9], we demonstrated the possibility that phosphorylation of the conserved motif within the DBD can be a common signal for nuclear receptors to diverge their functions.

## Methods

### Materials

An antibody against an anti-phospho-S216 peptide of ER $\alpha$  ( $\alpha$ P-S216) was produced and evaluated by AnaSpec Inc (San Jose, CA). Iba-1 antibody and Antibody Diluent were purchased from WAKO; biotinylated goat anti-rabbit antibody and Vectastain ABC reagents from Vector Laboratory (Burlingame, CA); Mouse Cytokine Array Panel A and an anti-F4/80 antibody from R&D Systems, Inc.; an antibody against green fluorescent protein (HRP-conjugated) from Abcam; an anti-Iba-1 antibody from Gene Tex; an Alexa 594 anti-Rat antibody and DMEM/F12 media from Life Technologies; Fixation/Permeabilization Solution Kit (BD Biosciences, San Jose, CA). Trizol from Life Technologies; Direct-zol<sup>TM</sup> RNA kit from Zymo Research; RNeasy mini kit from Qiagen; MultiScribe Reverse Transcriptase from Applied Biosystems; an anti-GFAP antibody from STEMCELL Technologies; Precision Plus Protein Standards from Bio-Rad. All reagents are highest qualities commercially available.

Animals: Mice were maintained on a 12 hr light/12 hr dark cycle and fed with NIH-31 the Open Formula Autoclavable diet (Zeigler, PA) and water ad libitum. Ex3-ER $\alpha$  KO mice were generous gift from Dr. Korach' lab. All research has been reviewed and approved by an Institutional Animal Care and Use Committee of NIEHS/NIH. LPS solution (1 mg/ml) was kept at -20°C and thawed just before intraperitoneal injection. All experiments were performed in accordance with relevant guidelines and regulations.

## Generation ERα S216A KI (*Esr1*<sup>S216A</sup>) mice

A 5.1-kb left arm containing introns 2 and 3 and exon 3 and a 2.0-kb right arm containing intron 3 were amplified from genomic DNAs of C57B/6 were cloned into pCR-XL-TOPO (Thermo Fisher Scientific, MA). Codon serine 216 encoded by exon 3 in the left arm was changed to alanine by site directed mutagenesis. After digestion with restriction enzymes, these DNA arms were cloned into the targeting vector which carries two multi cloning sites, self-excising ACN cassette [30] and DT-A cassette. The ACN cassette contains a testis-specific promoter from the angiotensin-converting enzyme gene that drives the expression of the Cre-recombinase gene, and RNA pol II promoter was used to drive neomycin registrant (*neor*) gene as a selection marker, which allows to screen ES cells in the presence of G418. When chimeras those are born from these targeted ES-cells containing the ACN cassette are bred for germline transmission, somatic cells derived from the ES cells retained the cassette, but self-excision occurred in all ES-cell-derived sperm, and as a result, the unexpected consequence due to the presence of Neo gene in chromosome can be avoided all together. Linearized targeting vector was electroporated into C57B/6 ES cells. The G418-resistant ES clones were screened by Southern blot analysis of KpnI- and BglI- digested genomic DNA with 5' and 3' external probes. BglI-digestion generated 12- and 8.3-kb bands for KI and WT, respectively, detected by using the 3'-genomic probes. These Southern probes were amplified from ES cell genomic DNA: 5' probe, 130966/131442

(tgcagctgcttctactgcttgcttgaatcatcataagataataaagcaaacagtaaaagatctgcggttggtaggggagtgcaatactatgatgatgaaatggaaagtgatgggtaatagaataggaacaagaactggg 3' probe, 139971/140460

(ctgaagagatggctcagtggttaagagcatcactgttctctccagaggattctggttcaattcccagctctcacatagcagtaataactgtctgtaactccagttcaagaggacctggcaccctcacagacataca The positive ES clones were injected into C57BL/6 blastocysts. Male chimeras were bred to C57BL/6 females to establish germline transmission, and the resulting heterozygous mice were interbred to obtain homozygous wild-type and knock-in mice. Primers for genotyping by PCR used for 5' probe 5'-TGCAGCTGCTTCTACTGGCTTGA-3' and 5'-TAAGGAAGCTCCTGGTTCCCTGCT-3'; for 3' probe, 5'-CTGAAGAGATGGCTCAGTGGTTAA -3' and 5'-CTGTTACAGACTAGAGAGGGT-3'

## Primary cortical mixed glial culture

Primary cortical mixed glial cultures were prepared from brains of mice at postnatal day 1-3, as previously described [31, 32]. Cortices were isolated from brains from which meninges and blood vessels were removed using forceps. Cells were dispersed by dissociating tissues in DMEM/F12 media through trituration. Obtained cell suspension were plated on either 24-well plates or 96-well plates pre-coated in poly-D-lysine (20 µg/ml) at  $1 \times 10^5$  cells/well or  $5 \times 10^4$  cells/well. Cells were maintained in DMEM-F12 (1:1) media supplemented with 10% heat-inactivated fetal bovine serum, 2 mM L-glutamine, 1 mM sodium pyruvate, 100 µM non-essential amino acids, 50 U/ml penicillin, and 50 µg/ml streptomycin. Culture medium was changed every 3 days. To allow high yield of microglia in the culture, the cells were cultured for 2 weeks. The mixed glial cultures with a ratio of 20% microglia and 80% astrocytes were obtained [31].

## Microglia-enriched cultures

Mouse microglia-enriched cultures were prepared from primary mixed glial cultures as previously described (1, 2). Mixed glia cultures were plated on 150 cm<sup>3</sup> flasks pre-coated with poly-D-lysine (20 µg/ml) at  $5 \times 10^7$  cells/flask and maintained in DMEM-F12 media changed every three days for two weeks. Then, matured microglia were shaken off at 180 rpm for 40 min and re-plated on glass-bottom culture dishes (MatTek, Ashland, MA, USA) pre-coated with poly-D-lysine (20 µg/ml) at  $1 \times 10^6$  cells/well.

## Immunohistochemical staining

Mouse brain was first perfused with cold PBS to remove bloods, from which sections (35 µm thick) were prepared. Brain sections were treated with 1% hydrogen peroxide for 10 min, incubated for 20 min with blocking solution (BSA 1%/Triton X-100 0.4%/Normal Goat Serum 4% in PBS) and incubated overnight at 4°C with rabbit polyclonal antibody against Iba-1 (1:4000) in Antibody Diluent. Stained sections were washed in PBS three times each for 10 min and incubated for 2 hr with PBS containing 0.3% Triton X-100 and a biotinylated goat anti-rabbit antibody (1:227). After washing three times with PBS, these sections were incubated for 1 hr with the Vectastain ABC reagents diluted in PBS containing 0.3% Triton X-100. Finally, these treated sections were incubated with 3,3'-diaminobenzidine and urea-hydrogen peroxide tablets dissolved in water to visualize microglia. The nigral densities of the Iba-1 immunostaining were measured using ImageJ software. To quantify the Iba-1 staining of microglial cells in the substantia nigra, representative images of Iba-1-positive regions in the substantia nigra were captured at 40× magnification. A total of 100 microglia in each mouse were selected randomly, and the Iba-1 density was measured and normalized with size of area selected. One way ANOVA plus post hoc test with Bonferroni's multiple comparisons test was used to analyze the difference between saline injected WT microglia vs. LPS-injected WT or LPS-injected ERα KI.

## ELISA

Cells were harvested and centrifuged to collect cultured media at time points after LPS (Millipore) treatment. Cytokine and metabolite concentrations were measured by ELISA. IL-6, IL-10 and PGE2 ELISA kits purchased from R&D Systems. ELISA assays were performed according to the manufacturer's instructions.

Cytokine antibody array: Mouse brains were homogenized in 500  $\mu$ L of cold PBS containing protease inhibitor cocktail and 5  $\mu$ L of Triton-X100 and centrifuged at 10,000  $\times$ g for 5 min at 4°C. Obtained lysates (300  $\mu$ g) were subjected to cytokine protein array. Cytokine expressions were detected by a mouse cytokine antibody array, panel A kit according to the manufacturer's instructions. Obtained spots were measured as densities by ImageJ software and showed in a graph. Cytokine mouse antibody array were performed according to the manufacturer's instructions.

### Double immunofluorescence staining

Immunofluorescence staining was performed as previously described [33]. Mouse mixed glia or enriched microglia cells were cultured on 35 mm bottom glass dishes, fixed with 4% formalin and blocked with a goat normal serum in PBS buffer for 20 min. For the first staining, these dishes were incubated with given antibodies such as an anti-ER $\alpha$  and P-S216 antibody for 30 min at room temperature. For the second staining, stained dishes were washed with PBS buffer and incubated with marker antibodies such as anti-Iba-1 and GFAP antibodies for 30 min at room temperature. Subsequently, after washed with PBS, these dishes were incubated with a goat anti-rabbit IgG secondary antibody, Alexa Fluor 488 and a goat anti-mouse IgG secondary antibody, Alexa 594 (1:500) (Thermo Fisher) mixture at room temperature for 1 hr in the dark. These stained cells were washed with PBS buffer and mounted with mounting medium containing DAPI (VECTASHIELD<sup>®</sup>). Stained cells in glass bottom dishes were observed using Zeiss 710 confocal microscopy (Zeiss).

### Flow Cytometry

To assess cell death, mixed glia cultures were stained with an anti-F4/80 antibody (1:50) for 30 min on ice and Alexa 594 anti-rat antibody (1:500), followed by staining by Annexin-V as previously described [34] and analyzed using FlowJo software. To assess IL-6 expression, mixed glia cells were stained with an anti-F4/80 antibody (1:50) for 30 min on ice and Alexa 594 anti-rat antibody (1:500), followed by fixation/permeabilization using Fixation/Permeabilization kit (BD Pharmingen<sup>TM</sup>), washed in FACS buffer (PBS, 2% BSA, 0.1 mM EDTA, 0.1% sodium azide), collected by centrifugation and re-suspended in BD Perm/Wash<sup>TM</sup> buffer. Then these cells were incubated with anti-PE-IL-6 antibody for 20 min on ice in dark. After washing, stained cells were suspended in 200  $\mu$ L of FACS buffer for immediate measurements of fluorescence by Flow cytometer (LSRII, BD Bioscience). Mean fluorescence intensity (MFI) was quantified using the FACSscan flow cytometer. The instrument used the FlowJo software for the analysis of the data.

### Western blots

Mouse uteri or brains were homogenized in 50 mM Tris-HCl buffer saline (pH 7.6) containing 8 M urea and 1% SDS. After centrifugation, resulting supernatant was added to SDS sample buffer. Protein concentrations were determined by Bio-Rad protein assay (Bio-RAD, Hercules, CA). Proteins were separated on a SDS-PAGE and transferred onto PVDF membranes (GE Healthcare, Pittsburgh, PA). These membranes were blocked with 5% BSA or 5% skim milk in 50 mM Tris-HCl-buffered saline containing 0.1% Tween-20 (TBS-T), incubated with given primary antibodies, washed with TBS-T, incubated with HRP-conjugated secondary antibodies and visualized using WesternBright Sirius HRP substrate (Advansta, Menlo Park, CA).

### RT-PCR

Total RNAs were extracted from of enrich microglial cells using Trizol and a Direct-zol<sup>TM</sup> RNA kit. An RNeasy mini kit was used to extract RNAs from mouse brains according to these manufacturer's instructions cDNAs were synthesized using MultiScribe Reverse Transcriptase. Real-time PCR was performed using an ABI prism 7700 sequence detection systems (Applied Biosystems) with following TaqMan probes (Applied Biosystems) used: human and mouse glyceraldehyde-3-phosphate dehydrogenase (Hs99999905\_m1 and Mm99999915\_g1) for an internal control, IL-1 $\alpha$  (Mm00434228\_m1), IL-1 $\beta$  (Mm00434228\_m1), IL-6 (Mm00446190\_m1), IL-10 (Mm00439614\_m1) and iNOS (Mm00440485-m1). Primers used for Cox-2 were primer-L: CAAGACAGATCATAAGCGAGGA and -R: GGCGCAGTTTATGTTGTCTGT. Assays were performed with a 7900HT Fast Real-Time PCR System (Applied Biosystems).

### Rotarod test

Rotarod test was conducted by Rotamex-5 (Columbus instruments, Columbus, OH, USA). Groups of 7 ER $\alpha$  WT and of 8 ER $\alpha$  KI males were trained for 4 consecutive days before they were tested at their 3- and 6-month of ages. Initial rotation of rotarod was set at 1 rpm and incrementally accelerated 1 rpm every 12 seconds. Retention times (latencies) on rotarod to fall off from the rotarod was measured three times for each mouse and averaged.

### Statistical analysis

Statistical analyses were conducted with One- or Two-Way ANOVA plus post hoc test with Bonferroni's multiple comparisons test or Tukey-Kramer's multiple comparisons test. Values are presented as means  $\pm$  S.E. or  $\pm$  S.D.

## Abbreviations

Cox-2: Cyclooxygenase-2

ELISA: Enzyme linked immunosorbent assay

ER $\alpha$ : Estrogen receptor  $\alpha$

FACS: Fluorescence-activated cell sorting

ICAM-1: Intercellular adhesion molecule-1

MFI: Mean fluorescence intensity

IL: Interleukin

iNOS: inducible nitric oxide synthase

KI: Knock-in

KO: Knockout

LPS: Lipopolysaccharide

NF- $\kappa$ B: Nuclear factor-kappa B

PCR: Polymerase chain reaction

PGE2: Prostaglandin E2

RXR $\alpha$ : Retinoid X receptor  $\alpha$

TIMP-1: Tissue inhibitors of metalloproteinases-1

TNF $\alpha$ : Tumor necrosis factor  $\alpha$

WT: Wild type

## Declarations

### Ethics approval and consent to participate

Not applicable.

### Consent for publication

Not applicable.

### Availability of data and materials

Not applicable.

### Competing interests

The authors declare that they have no competing interests.

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### Author Contributions

All authors conceived and designed the experiments. SS and MN performed most of the experiments, analyzed and interpreted data and wrote the manuscript. SG and MR designed and produced *Esr1*<sup>S216A</sup> mouse. SS, S-HC, KY, HH, RM and JM performed the experiments and analyzed the data. All authors wrote or contributed to the writing of the manuscript. All authors read and approved the final manuscript.

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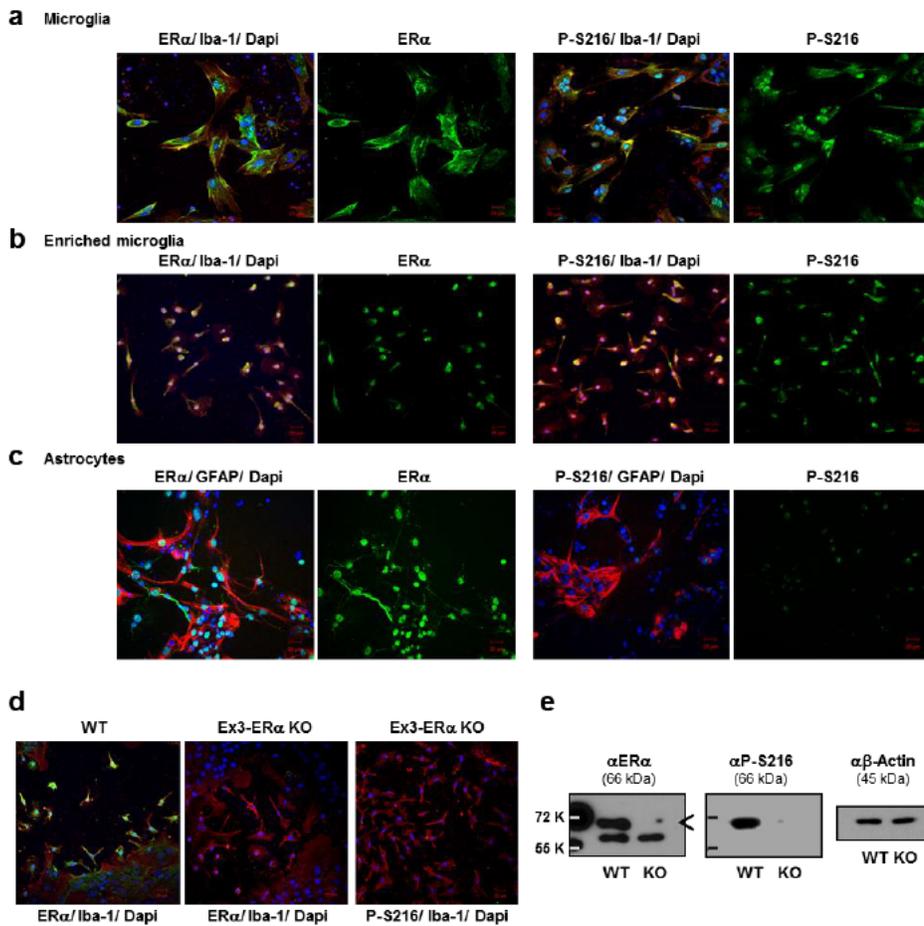
Sawako Shindo, Kiyoshi Nagata

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## Figures



**Figure 1**

ERα phosphorylated at Ser216 in mouse brain microglia. (a) Microglia. Mixed glia cells were prepared, cultured for 4-days and double-stained with either an anti-ERα (in green) or an anti-P-S216 (in green) antibody and an anti-Iba-1 (in red) antibody for microglia marker and visualized as described in the Methods section. Nuclei were stained by DAPI in blue. (b) Enriched microglia. Microglia was enriched from glia cells as described in the Methods section for double staining. (c) Astrocytes. Mixed glia cells were double-stained with either an anti-ERα (in green) or an anti-P-S216 (in green) antibody and an anti-GFAP (in red) antibody for astrocyte marker and were visualized as described in the Methods section. Nuclei were stained by DAPI in blue. (d) Glia cells obtained from 2-days-old neonates of Ex3-ERα KO and wild type (WT) mice were cultured and double-stained by an anti-ERα or an anti-P-Ser-216 (P-S216) (in green) and an anti-Iba-1 (in red) antibodies and visualized as described in the Methods section. Nuclei were stained by DAPI in blue. (e) Whole lysates prepared from enriched microglia were subjected to Western blots by an anti-ERα or an anti-P-S216 antibody. An anti-β-Actin antibody was utilized to verify equal amounts of proteins in an each well.

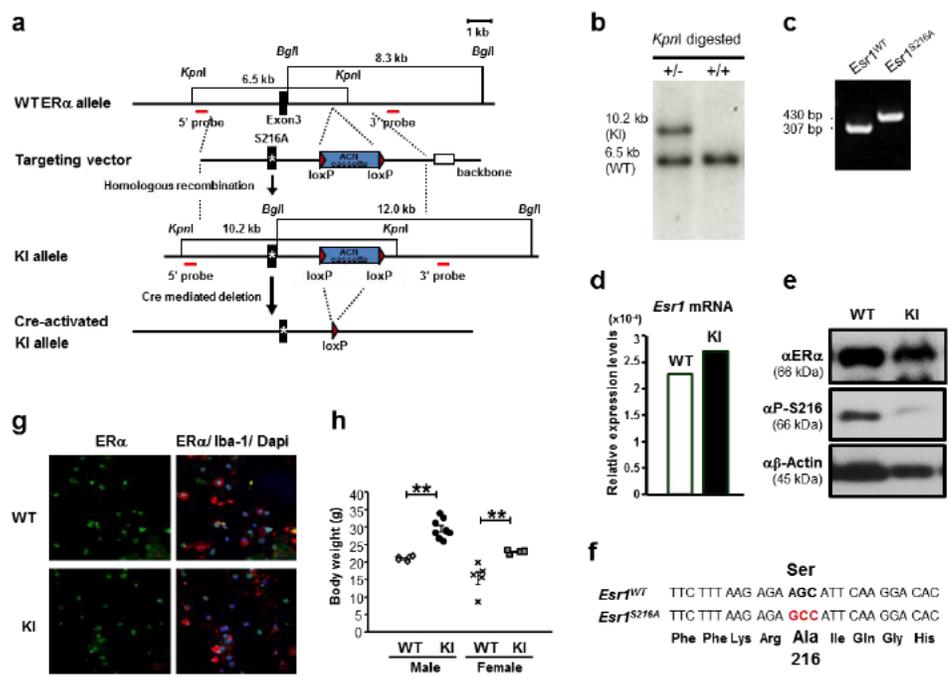


Figure 2

Generation and phenotypes of ER $\alpha$  S216A KI ( $Esr1^{S216A}$ ) mice. (a) Map of strategy and Knock-in process. (b) Southern blot analyses were performed to identify the appropriately targeted allele. Genomic DNAs from mouse ear biopsies were digested with *KpnI* and *BglI*. *KpnI*-digestion generated 10.2- and 6.5-kb bands for knock-in (KI) and wild type (WT), respectively, detected by using the 5'-genomic probe. (c) Genotyping by PCR. Genomic DNA was isolated from mouse ear biopsies. PCR primers were used to amplify across the region in which the one remaining loxP site was inserted. This gave a band of 430 bp for a knock-in allele and 307 bp for a wild-type allele. Self-excision of the ACN cassette in the mutant allele was confirmed. (d) Expression of ER $\alpha$  mRNA. RNAs were prepared from WT and KI uteri and subjected to real time PCR analysis. These are averages from duplicates of two independent samples. (e) Conformation of ER $\alpha$  protein expression in WT and KI uteri. Whole extracts (15  $\mu$ g proteins) prepared from WT and KI uteri were subjected to Western blots by an anti-ER $\alpha$ , an anti-P-Ser-216 (P-S216) or an anti- $\beta$ -Actin antibody. (f) Sequence conformation of the mutation. cDNAs were synthesized from WT and KI uteri and sequenced. (g) ER $\alpha$  expression in ER $\alpha$  KI microglia. Accumulated microglia in the glia cells, which were prepared from brains of 2 days-old neonates of ER $\alpha$  KI males, were subjected to fluorescence staining with an anti-ER $\alpha$  (in green) antibody or double staining with an anti-Iba-1 (in red) antibody. DAPI stained nuclei in blue. (h) Obese phenotype of ER $\alpha$  S216A KI mice. Six-month-old mice were weighed; 5 males and 5 females of WT and 8 males and 4 females of KI mice. One Way ANOVA plus post hoc test with Tukey-Kramer's multiple comparisons test (Version 5.0, Stat view-j) was used for statistical analysis. Values are presented as means  $\pm$  S.E.. \*\*,  $p < 0.01$ .

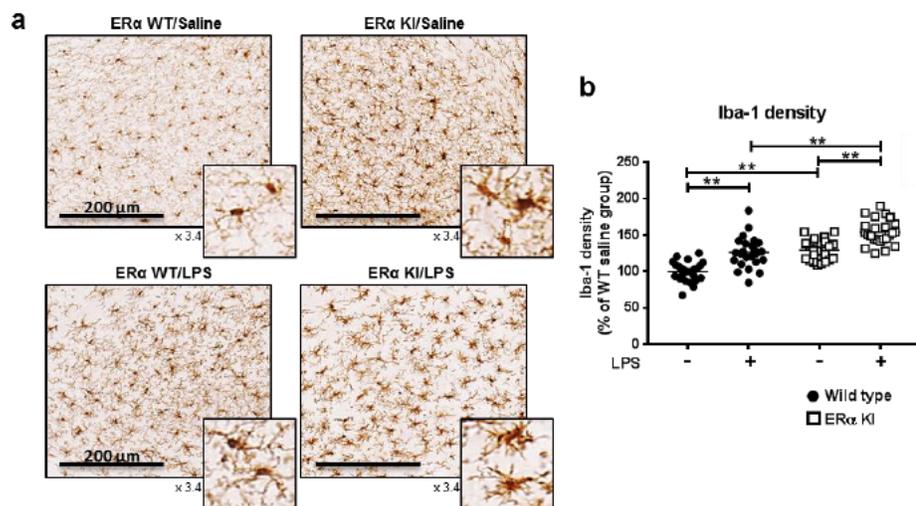
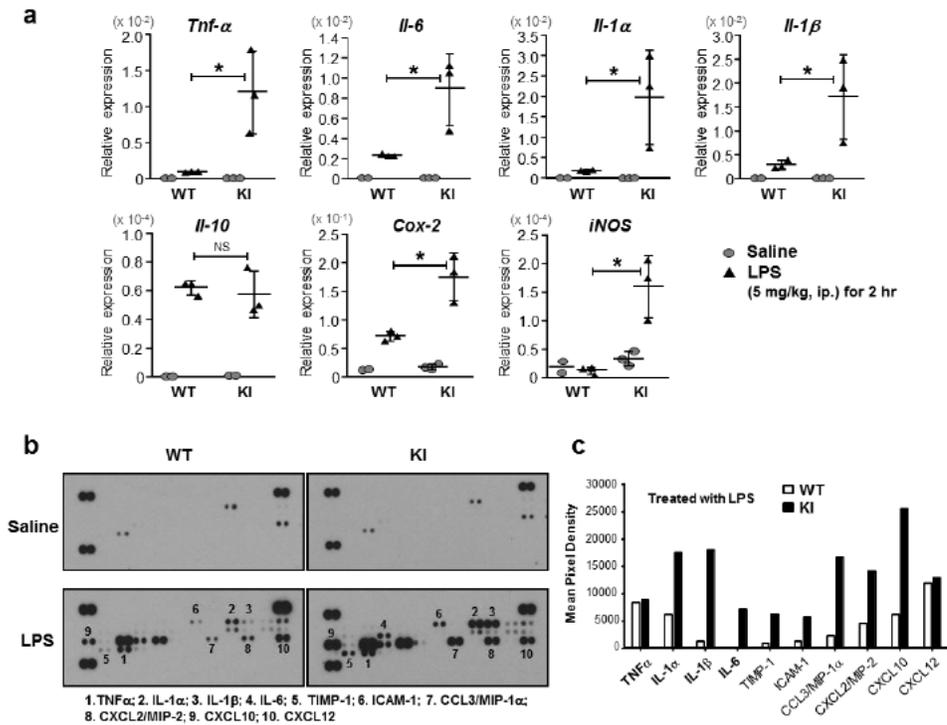
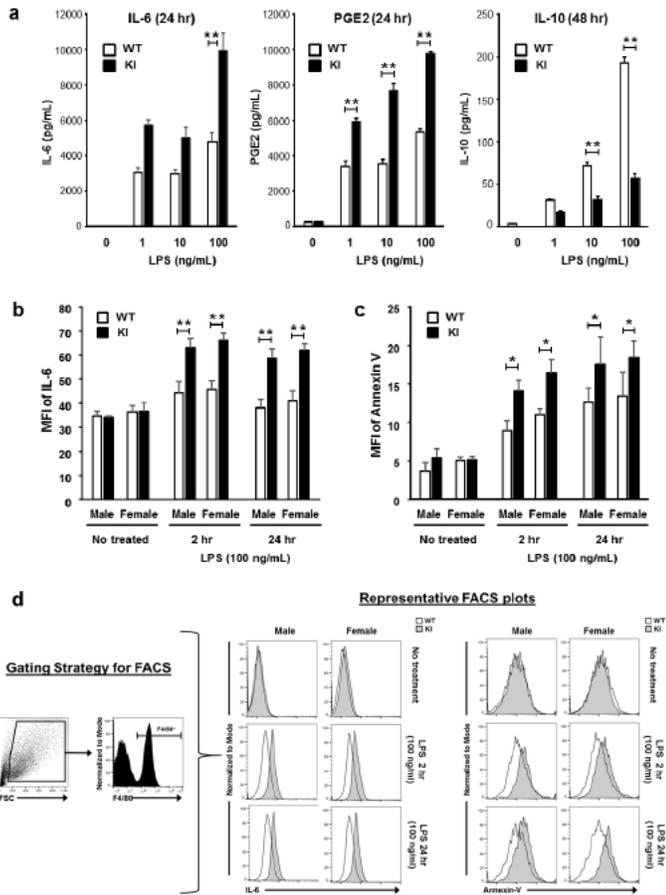


Figure 3

Microglia in substantia nigra of adult brains. (a) Saline or LPS (5 mg/kg, ip., 2 hr)-treated brains. Brains were removed from 6-month-old ER $\alpha$  WT and KI males, from which frozen sections were prepared for immunostaining with an anti Iba-1 antibody. (b) Three brains were used for each group. Four sections from each brain were stained for analysis. Stained microglia were scanned to semi-quantitate microglia activation. Values are presented as means  $\pm$  S.E., by taking values of ER $\alpha$  WT as 100%. Statistical analysis was conducted with One Way ANOVA plus post hoc test with Bonferroni's multiple comparisons test (Version 7.00, GraphPad, San Diego, CA). \*\*,  $p < 0.01$ .



**Figure 4** Expression of inflammatory factors in adult brains. (a) qRT-PCR. Adult WT or KI male mice were treated with intraperitoneally injected LPS (5 mg/kg, ip.) or saline for each group for 2 hr, from the brains of which RNAs were prepared for subsequent analysis. Assay was duplicated for each RNA sample. With 6 RT-PCR data for either WT or KI, Two Way ANOVA plus post hoc test with Bonferroni's multiple comparisons test (Version 5.0, Stat view-j) was used for statistical analysis. Values are presented as means  $\pm$  S.D.,  $n = 3$ . (b) Cytokine antibody arrays. From the same brains used in (a), whole extracts were prepared, 300  $\mu$ g proteins of which were subjected to analysis. (c) Mean Pixel Densities were obtained from each spot using Image J soft.



**Figure 5**

Expression of inflammatory factors in cultured microglia. (a) ELISA. Mixed glia cells were prepared from 5 neonates and cultured for 2 weeks and then treated with 1, 10 or 100 ng/mL of LPS or PBS for an additional 24 or 48 hr. Media were recovered and subjected to ELISA for IL-6, PGE2 and IL-10. WT and KI denote ER $\alpha$  wild type and KI mice, respectively. ONE Way ANOVA plus post hoc test with Tukey-Kramer's multiple comparisons test (Version 5.0, Stat view-j) was used for statistical analysis. Values are presented as means  $\pm$  S.D.. \*\*,  $p < 0.01$ ,  $n=4$ . (b) FACS. Mixed glia cells were separately prepared from male and female neonates, cultured for 2 weeks and then treated with 100 ng/mL of LPS or PBS for an additional 2 or 24 hr. IL-6 production was determined by intracellular staining, followed by flow cytometry analysis. Assay was triplicated with three independent. ONE Way ANOVA plus post hoc test with Tukey-Kramer's multiple comparisons test (Version 5.0, Stat view-j) was used for statistical analysis. Values are presented as means  $\pm$  S.D.. \*\*,  $p < 0.01$ . (c) Cell death was assessed by Annexin V staining, following by flow cytometry analysis. Assay was performed in triplicate eight times with three independent samples. These were separately prepared from male and female neonates. Two Way ANOVA plus post hoc test with Tukey-Kramer's multiple comparisons test (GraphPad Prism) was used for statistical analysis. Values are presented as means  $\pm$  S.D.. \*,  $p < 0.05$ . (d) The gating strategy and the representative FACS plots for Fig. 5b and 5c.

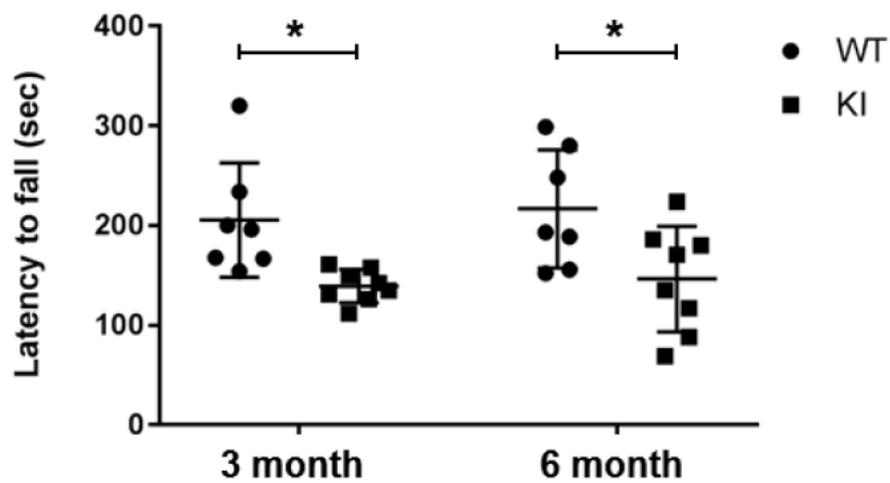


Figure 6

Latency to fall in rotarod test. Three-month-old and 6-month-old male mice were tested as described in the Experimental Procedure section. These values are represented as mean  $\pm$  S.D. of 7-8 mice for each group. Data were analyzed by Two Way ANOVA plus post hoc test with Bonferroni's multiple comparisons test (version 7.02, GraphPad Prism) and are shown as scatter plots. \*,  $p < 0.05$ .

## Supplementary Files

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