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## Research

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## **Stabile SERS encoded silver silica nanocomposites for industrial labeling – the case of COVID-19 diagnosis**

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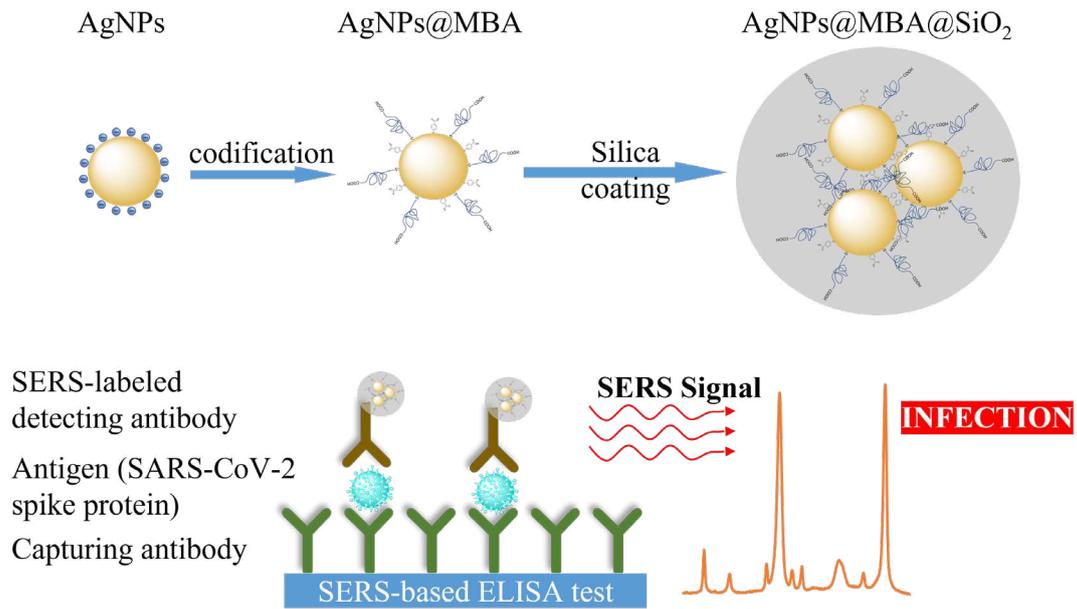
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### **ABSTRACT:**

Biosensors, especially those with a SERS readout, are required for an early and precise healthcare diagnosis. Unreproducible SERS platforms hampers the clinical translation of SERS. Here we report a synthetic procedure to obtain stabile, reproducible, and robust highly-SERS performing nanocomposites for labelling. We control the NPs agglomeration and codification which results in an increased number of hot spots, thus exhibiting reproducible and superior Raman enhancement. We studied fundamental aspects affecting the plasmonic thiol bond resulting in pH exhibiting a determining role. We validated their biosensing performance by designing a SERS-based sandwich immunoassay against COVID-19. The limits of detection for the recombinant SARS-CoV-2 protein is below 0.01 ng/μL. We offered herein one nanostructure with robust and homogeneous SERS signal which can be potentially applied for biodiagnosis.

**Keywords:** SERS-codified plasmonic silica nanocomposites, metallic-thiol bond, controlled agglomeration, COVID-19 SERS-based ELISA assay, industrial labelling/biosensing.



**Graphical abstract:** Scheme of the rational and controlled synthesis of robust and stable SERS encoded plasmonic-silica nanocomposites for enhanced, fast, sensitive, and selective labeling and biosensing.

## INTRODUCTION:

The design for precise diagnosis is critical to human health as for preventing pandemics or other biothreats. Nanosystems has been widely developed in sensor devices for diagnostics, *in vitro* and *in vivo* diagnosis(1). This diagnosis field takes benefits from the design and synthesis of nanomaterials, especially noble metal nanomaterials showing localized surface plasmon resonance (LSPR) properties. This LSPR phenomenon limits nanomaterials absorbing specific region of light, and makes the nanomaterials sensitive to the modifications of physical properties of nanomaterials and their environments showing plasmonic absorption shifts(2). Based on their optical properties, flexible functionalized nanomaterials have already been applied in sensing varieties of biomolecules, including biomarkers for cancer(3), enzyme(4), DNA(5), and other biological species(6).

Surface enhanced Raman spectroscopy (SERS) also depends on plasmonic platforms. SERS signals can be collected from the molecules which are in close proximity of nanometallic surfaces with confined LSPR(7). In general, average enhancement factors for typical SERS substrates are amplified by  $10^6$  to  $10^8$  orders of magnitude comparing with their Raman signature(8). Moreover, this enhancement can be further increased up to  $10^{15}$  orders of magnitude to SERS signal by hot spots(9), which are highly localized regions of intense local field enhancement caused by the plasmonic coupling of the particles when they are very close to each other(8). Thus, hot spots have critical importance when designing a SERS nanostructure. Gold and silver are the most commonly applied materials for SERS substrates, as they offer high field enhancement in the visible to near infrared range due to their high density of electrons(10). SERS as a nondestructive technology provides chemical information in aqueous environments. Thus SERS has been an effective tool to realize qualitative and quantitative detection of biological species, including micro RNA analysis(11), enzyme(12), hydrogen peroxide(13), staphylococcal enterotoxin B(14) and other diseases biomarkers (15)(16) with gold nanowire, gold-silver alloy NPs and gold-MnO<sub>2</sub> core-shell, hollow gold nanospheres and gold nanostar and nanosphere separately. However, producing homogenous, sensitive and reproducible SERS platforms is the main difficulty which hampers SERS bioapplications(17), with many efforts made in the design and synthesis of uniform nanomaterials (18)(19)(20). The design and controllable synthesis of nanostructures are critical steps towards implementing SERS in medicine.

Here we present a controllable design and synthesis of a nanostructure confining encoded silver nano-agglomerates inside silica coating (AgNPs@MBA@SiO<sub>2</sub>). We systematically studied the fundamental aspects and optimized the thiol silver bonding for encoding silver nanoparticles. With the help of controlled agglomeration, we improved the percentage of hot spots which guaranteed the extremely high Raman enhancement. These encoded agglomerates were further encapsulated with a silica shell to protect them from oxidation, contaminations and increase the stability for a long period of time thanks to the unique properties of the SiO<sub>2</sub> layer (*e.g.*, surface chemistry, biocompatibility, optical transparency, and colloidal stability). With further biofunctionalization with antibody, we also demonstrated the performance of our devices for SARS-CoV-2 spike protein detection. The reference diagnosis for SARS-CoV-2 is based on reverse transcription polymerase chain reaction (RT-PCR)(21). Yet there are false positive or

negative reports, especially for the early stages. Biosensors as alternative or supplementary solutions have been developed based on plasmonic nanomaterials. Gold nanoparticles have been developed for colorimetric detection of SARS-CoV-2 with isolated RNA samples(22) and for IgM Antibodies against the SARS-CoV-2 virus detection based on a lateral flow device(23). A more sophisticated devices based on gold nanoislands was reported for SARS-CoV-2 detection targeting selected sequences(24). Compared with the biosensors mentioned, we selected the detection of SARS-CoV-2 spike protein, which is the major immunodominant protein(25). One of the promising advantages of our biosensor is that it can potentially screen SARS-CoV-2 without prior sample treatment. This whole designed procedure provides our nanostructure high Raman enhancement and robust intensity for sensitive sensing, and uniform synthesis for stability and repeatability. We offered an optimized, stable nanostructure which can be reliably applied in biosensing.

## EXPERIMENTAL SECTION

In the supporting information file, there is a more detail description of the materials and methods along with additional results.

### AgNPs@MBA@SiO<sub>2</sub> Synthesis

The detailed synthesis of AgNPs@MBA@SiO<sub>2</sub> can be seen in supporting information. In briefly, we performed a bottom up synthesis of AgNPs relying on the chemical reduction of metal salts and controlling the shape and size by citrate reduction of AgNO<sub>3</sub> (26). Then, we co-adsorbed and covalently bound onto the metallic AgNPs surfaces a SERS probe 4-mercaptobenzoic acid (MBA) and a polymeric stabilizer CTPEG12. CTPEG12 and MBA amounts were calculated based on the area of the NPs required by them *i.e.*, 1 molecule/nm<sup>2</sup> of the CTPEG12 and 3 molecules/nm<sup>2</sup> of MBA. Finally, we controlled their agglomeration into nanoagglomerates and we encapsulated them within a homogenous layer of silica to protect the readout (MBA) and the SERS enhancer (metallic AgNPs surface). Silica coating was performed by a modified Stöber method(27).

### Physicochemical characterization

Transmission electron microscopy (TEM): we checked the morphology of silica encapsulated MBA modified nanoagglomerates (AgNPs@MBA@SiO<sub>2</sub> and AuNPs@MBA@SiO<sub>2</sub> (*cf.* SI)) with a JEOL JEM 1010 TEM operating at an acceleration voltage of 80 kV with a tungsten filament. With the obtained images, we calculated the average size and polydispersity analysing at least 100 particles with the image process software “Image J”.

Dynamic light scattering (DLS) and zeta potential measurements were performed with Malvern Zetasizer Nano ZS. Aqueous samples were transferred into disposable polystyrene cuvette for size measurements and disposable folded capillary cells for zeta potential measurements. Each sample was measured 3 repeats.

The extinction spectrum of each synthetic intermediate was recorded with Ultrospec™ 2100 pro UV-Visible spectrophotometer using a Quartz cuvette. We collected the extinction spectra between 250 and 900 nm using Milli-Q water as a reference.

### Surface enhanced Raman spectroscopy

The SERS spectra was collected with a Renishaw's inVia Qontor Raman system equipped with a Leica confocal microscope.  $4.55 \times 10^{10}$  NPs/mL (calculated by UV-vis extinction) 200  $\mu$ l of AgNPs@MBA were irradiated with a NIR laser (785 nm) and imaged with an 5x objective (NA 0.12), providing a laser spot diameter of approximate 8  $\mu$ m. The spectra were collected with 1s exposure time and 100 mW laser power at the samples.

### **Labelling efficiency of AgNPs@MBA@SiO<sub>2</sub> onto different substrates.**

We labelled different materials with different compositions and patterns with 20  $\mu$ l of AgNPs@MBA@SiO<sub>2</sub> at a concentration of  $7 \times 10^{10}$  NPs/mL. The materials used are: semi-aniline leather, aniline leather, pigmented leather, polyester, silk, plastic (PVC), glass, brass, cotton, pigmented leather.

SERS characterization of all samples prepared here were conducted using Renishaw inVia Qontor Raman. 20X Leica objective was used with integration time 0.1 s and a power at the sample of 3 mW.

The surface of AgNPs@MBA@SiO<sub>2</sub> deposited materials were analyzed with a scanning electron microscope (SEM) from Phenom XL Desktop SEM with a Backscattered electron detector. The elemental analysis was conducted by energy-dispersive X-ray spectroscopy equipped with the SEM.

### **SERS-based sandwich immunoassay to diagnose COVID-19.**

AgNPs@MBA@SiO<sub>2</sub> were biofunctionalized with SARS-CoV-2 Spike antibody (AgNPs@MBA@SiO<sub>2</sub>@Ab) using the linker GPTMS. Then we validate their performance to detect SARS-CoV-2 spike recombinant proteins in a sandwich ELISA-like immunoassay.

We first coated the testing plates with a SARS-CoV-2 capturing antibody (50  $\mu$ l of 6  $\mu$ g/mL) over night at 4°C. SARS-CoV-2 spike proteins were diluted into a concentration series: 0.01, 0.02, 0.1, 0.2, 0.4, 0.6, 0.8, 1, 1.2, 1.6 ng/ $\mu$ L. We transferred 50  $\mu$ l of diluted spike protein (and PBS used as negative control) into each well of the testing plate. After 1h, we added 50  $\mu$ l  $9 \times 10^9$  NPs/mL AgNPs@MBA@SiO<sub>2</sub>@Ab. After 1h incubation at 37°C, we acquired the SERS signal with a 5x Leica objective, 1 s integration time, and 50 mW laser power. We present the data collected from 8 spectra from 8 random places of the testing plates.

### **Limit of detection (LOD)**

The limit of detection (LOD) was calculated by measuring the intensity at 1075  $\text{cm}^{-1}$  with the presence and the absence of antigen, with a 3:1 ratio threshold (Signal / Noise = 3).

## RESULTS AND DISCUSSION

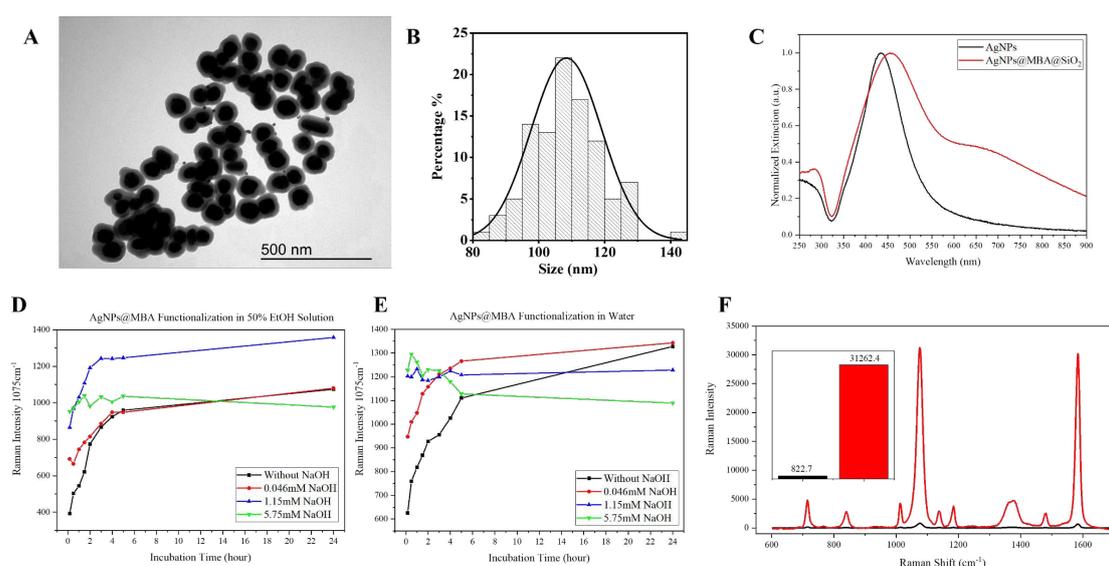
### AgNPs@MBA@SiO<sub>2</sub> synthesis and characterization

We have synthesized a SERS encoded core-shell nanostructure comprising silver NPs agglomerates. Figure 1A shows the TEM image of AgNPs@MBA@SiO<sub>2</sub>, clearly demonstrating that each agglomerate was encapsulated within the silica protecting layer. The AgNPs were spherical with the anisotropy aspect ratio close to 1. Around 1% of the particles were non-spherical, *i.e.*, rod-shaped particles or quasi-flat triangles (calculated from approx. 100 NPs). Based on more than 100 NPs, we calculated the size of AgNPs@MBA@SiO<sub>2</sub> having each NP four different angles for measurement. The AgNPs core was approx. 60-70 nm and the SiO<sub>2</sub> layer was estimated around 20 nm. The complete size of AgNPs@MBA@SiO<sub>2</sub> was around 110 nm (figure 1B). More than 60% of the agglomerates were isolated dimers, trimers, tetramers, pentamers, hexamers, or mixtures thereof. The AgNPs@MBA@SiO<sub>2</sub> have a hydrodynamic diameter of 133.8 nm with PDI 0.130 and a zeta potential value of -24.7 mV typical for the silica coating (*cf.*, SI, figure SI-1). The hydrodynamic size agrees with the measurements obtained with the TEM. The NPs size, concentration and agglomeration degree can also be characterized with the UV-Vis spectrum(28). Figure 1C shows the normalized extinction spectra of the individual, citrate capped AgNPs and the nanoagglomerates, AgNPs@MBA@SiO<sub>2</sub>. The characteristic localized-surface plasmon resonances (LSPRs) of silver spherical nanoparticles is shown in the UV-Vis spectra of AgNPs which centered in the UV spectral range around 430 nm. As for AgNPs@MBA@SiO<sub>2</sub>, the new prominent shoulder absorption at around 700 nm, and the red-shift and broadening of this characteristic LSPR to higher wavelengths indicates the significant formation of nanoagglomerates.

Aromatic compounds are commonly used as SERS or Raman probes as they have high Raman cross section. Previous reported articles used ethanol as solvent during Raman probe modification based on the consideration of low solubility of hydrophobic Raman probes(29). However, to the best of our knowledge, there is still lack of detailed information on the fundamental factors affecting Raman probes NPs' modification. Therefore, we studied the formation of thiol-silver bond as a function of pH and solvent within this work. The SERS probe used here is an aromatic molecule MBA which has high Raman cross section(30) (31) and the polymeric stabilizer CTPEG12 is an aliphatic chain polymer with 12 carbon atoms and a carboxylic group at the end of the chain. Both MBA and CTPEG12 were bonded to AgNPs through their thiol group by forming covalent bonds.

To study the pH and solvent effect on MBA adsorption, we monitor the evolution of the characteristic ring breathing band of adsorbed MBA at 1075 cm<sup>-1</sup> over the time(32) and under different conditions (figure 1D and 1E). AgNPs@MBA were dispersed in 50% EtOH/water solution (figure 1D and *cf.*, SI, figure SI-2) or in aqueous solution (figure 1E and *cf.*, SI, figure SI-3) containing no or different amount of NaOH from 10 min to 24 h. Water and ethanol have different physical property (*e.g.*, pKa and dielectric constant), therefore, it was not surprising to measure different pH values for each of the solvents even with the same amount of NaOH(33) (*cf.*, SI, table SI-1).

Figure 1D and 1E shows the absorption and bonding kinetic results of MBA onto the AgNPs under the different conditions (pH and solvent) and over time. When comparing the first 10 min after the addition of Raman code in both solvents, the intensity at  $1075\text{ cm}^{-1}$  increased along with the system pH and amount of added NaOH. The codification reached a plateau intensity in a pH dependent manner being shorter under increased alkaline conditions. The adsorption of thiol onto silver metal surface follows physisorption to chemisorption where the S-H bond breaks and forms silver thiol covalent bond(34). Environmental pH will affect the deprotonation of thiol group, thus will affect the formation speed of silver thiol bond. As we observed in our system, an alkaline environment facilitates the codification of MBA onto silver metallic surfaces.



**Figure 1: Synthesis and characterization of the SERS-responsive nanoagglomerates. (A)** TEM image of AgNPs@MBA@SiO<sub>2</sub>. **(B)** size distribution of AgNPs@MBA@SiO<sub>2</sub> based on the quantification of 100 NPs from TEM images. **(C)** UV-Visible extinction spectra of AgNPs (black) and AgNPs@MBA@SiO<sub>2</sub> (red). **(D and E)** MBA adsorption kinetic under different conditions (with or without NaOH) measured by SERS intensity at  $1075\text{ cm}^{-1}$  **(D)** in 50% EtOH/water solution and **(E)** in aqueous solution. Black line: without NaOH; red line: with 0.046 mM NaOH; blue line: with 1.15mM NaOH; green line: with 5.75mM NaOH. **(F)** SERS spectra and SERS intensity at  $1075\text{cm}^{-1}$  (inset image) of non-agglomerated AgNPs@MBA@SiO<sub>2</sub> (in black), and nanoagglomerates, AgNPs@MBA@SiO<sub>2</sub> (in red).

When comparing the plateau intensity after 24 h, the number of adsorbed MBA were similar when the codification was performed in 50% ethanol/water solution with 1.15 mM NaOH (pH 8) and in aqueous solution with 0.046 mM NaOH (pH 8.5). These conditions exhibited the maximum SERS intensity among all. Subsequent additions of NaOH and pH increase resulted in a MBA codification decrease in both solvents. Considering that the pKa of MBA and CTPEG12 is around 4 to 5(35), the deprotonated MBA and CTPEG12 are both negatively charged. This electrostatic repulsion on the surface and the competition between MAB and CTPEG12 will hamper MBA bonding from the bulk solution, since in general, compared with aromatic thiols, aliphatic thiols have better electrochemical and thermodynamic stability(36). Dissociation was also favored at higher pH(37), which could also be related to this dynamic equilibrium.

Ethanol has little effect on the modification and stability of metal-thiol bond. Although it was demonstrated that because of the reduction effect of ethanol on gold, the strength of metal-thiol contacts can be weakened(38), ethanol is still recommended for hydrophobic Raman probes.

The number of MBA molecules adsorbed on the metallic surface and the packing of the monolayer is discussed within this work. We show that they depend on factors like the metal-thiol bonding formation speed, which is affected by pH, the electrostatic repulsion on the metallic surface, and the competition between Raman probe and stabilizer. Controlling the pH in reaction is especially critical to achieve optimized codification of metallic surfaces with thiolated aromatic compounds.

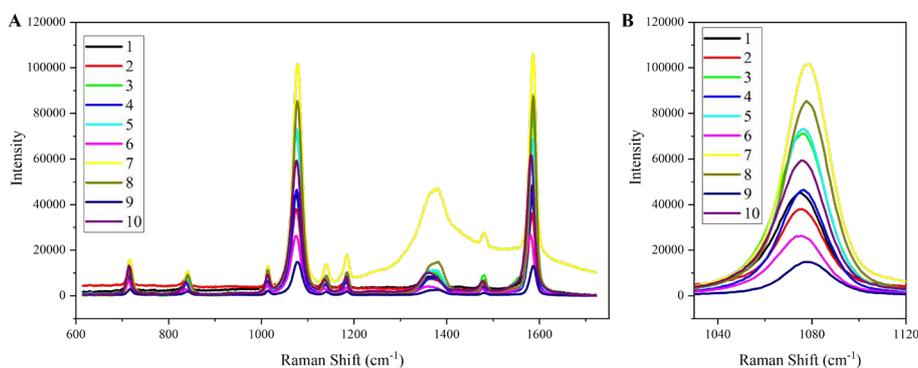
To confirm that we were controlling the degree of agglomeration to achieve optimal optical performance in the AgNPs@MBA@SiO<sub>2</sub>, we optically characterized the individual systems (AgNPs and AgNPs@MBA). UV-vis was performed under all codification conditions after 24 h addition of the Raman probe to monitor their LSPR informing on their degree of agglomeration and geometrical changes (*cf.*, SI, figure SI-4). As confirmed, both, AgNPs and AgNPs@MBA, showed the characteristic LSPR of isolated NPs at around 435 nm under all codification conditions. Furthermore, compared with the plasmonic absorption of AgNPs, AgNPs@MBA suffered a red shift, around 12 nm, indicating proper adsorption of MBA onto the metallic surface. None of the NPs showed absorption features in the NIR region (approx. 650 nm) attributed to plasmonic coupling of interacting NPs (*i.e.*, agglomeration) as shown for AgNPs@MBA@SiO<sub>2</sub> (figure 1C). Thus, confirming a control over the AgNPs@MBA synthesis to design enhanced SERS responsive materials.

By controlling the agglomeration of AgNPs@MBA and the MBA adsorption, we managed to increase the SERS efficiency up to nearly 40 times when comparing the SERS spectra (characteristic peak intensity at 1075 cm<sup>-1</sup>) of AgNPs@MBA@SiO<sub>2</sub> and non-agglomerated AgNPs@MBA@SiO<sub>2</sub> (figure 1F). This high SERS response was attributed to active SERS structures called “hot spots”, which are highly localized regions of intense local field enhancement caused by the plasmonic coupling of the NPs when they are very close to each other(8). In comparison to aggregation or uncontrolled agglomeration, where the NPs merged without hot spots(29), controlled agglomeration offers a useful tool to rationally design nanomaterials for SERS labeling.

Our optimized protocol for synthesizing a SERS encoded core-shell nanostructure can be further applied to gold nanomaterials to broaden the application field (*cf.*, SI, figure SI-5). We did observe differences in the SERS performance. AuNPs@MBA@SiO<sub>2</sub> nanoagglomerates enhanced 2 times (*cf.*, SI, figure SI-5F) whereas AgNPs@MBA@SiO<sub>2</sub> nanoagglomerates achieved almost 40 times higher signals (figure 1F) compared to their non-agglomerated counterparts.

## Stability of the AgNPs@MBA@SiO<sub>2</sub> SERS signal after labeling different types of substrates

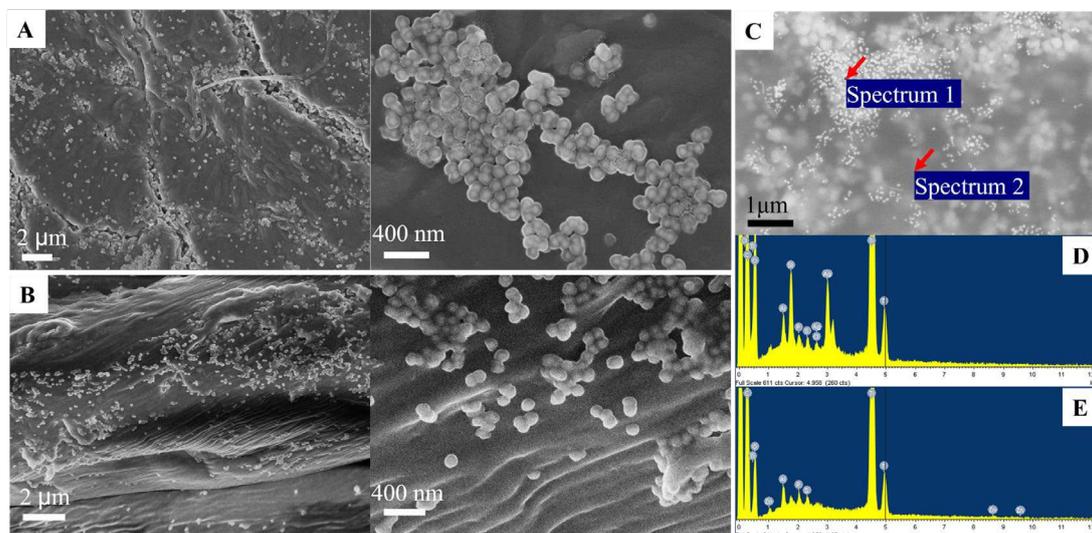
We analyzed the SERS signal robustness and stability of our AgNPs@MBA@SiO<sub>2</sub> by depositing them onto 10 different substrates made of different materials (*cf.*, SI, figure SI-6). Figure 2 shows corresponding SERS spectra and zoomed characteristic MBA peak at 1075 cm<sup>-1</sup> of AgNPs@MBA@SiO<sub>2</sub> demonstrating efficient labelling of all substrates. Except for glass which shows a broad band between 1000 to 1800 cm<sup>-1</sup>, the background signal originated from all substrates was neglectable. Regardless, both effects did not affect the characteristic bands of MBA at 1075 cm<sup>-1</sup> which is used for tracing and labelling. We did observe an uneven SERS intensity between all substrates that can be explained by differences in the distribution of AgNPs@MBA@SiO<sub>2</sub> within the different substrates.



**Figure 2: SERS characterization of different substrates wherein the AgNPs@MBA@SiO<sub>2</sub> has been placed. (A) SERS spectra and (B) zoomed spectra showing characteristic peak at 1075 cm<sup>-1</sup> of AgNPs@MBA@SiO<sub>2</sub> on different materials: 1, semi-aniline leather; 2, aniline leather; 3, pigmented leather; 4, polyester; 5, silk; 6, plastic (PVC); 7, glass; 8, brass; 9, cotton; and 10, dyed pigmented leather.**

We decided to examine the AgNPs@MBA@SiO<sub>2</sub> distribution within the different substrates and its impact on the NPs' geometry (figure 3). For example, in the case of cotton (figure 3B), the NPs are deposited onto the cotton fibers and hidden within the fibers, whereas in the leather (figure 3A), the NPs were more localized in the labeling spot (zoomed SEM areas of figure 3A and 3B). SEM images shows that the distribution can vary with surface properties of the substrates like roughness and porosity. Distribution of the NPs within the substrate can influence the SERS signal. Indeed, we observed that the SERS signal for the dye pigmented leather was higher than for cotton (figure 2). The leather is less rough and porous than cotton, thus hindering the diffusion of the AgNPs@MBA@SiO<sub>2</sub> through the substrate and enhancing the SERS signal. At any rate, both substrates demonstrated preservation of the NPs' geometry, thus SERS signal despite the different distribution. For the elemental analysis (figures 3D and 3E), we took two areas of the labeled spot on the leather, one with visible AgNPs@MBA@SiO<sub>2</sub> (spectrum 1) and another with no visible NPs (spectrum 2). Figures 3D confirms the presence of trace elements from the AgNPs@MBA@SiO<sub>2</sub>.

AgNPs@MBA@SiO<sub>2</sub> exhibit robust SERS signals; obtained in 0.1 s and with insignificant interference by other components of the substrate. This provides our SERS-responsive NPs with numerous potential applications where high optical signal is needed. The application field ranges from industrial applications where a robust, labelling signal



**Figure 3: Scanning Electron Microscopy (SEM) and elemental analysis (EDX) of the materials labelled with AgNPs@MBA@SiO<sub>2</sub>.** SEM images of leather (A) or cotton (B) labelled with AgNPs@MBA@SiO<sub>2</sub> showing their distribution within the substrate depending on their characteristics (C) SEM images of (A) showing areas with (spectrum 1) and without (spectrum 2) visible NPs that were selected for elemental analysis. (D and E) Their corresponding spectra. The peaks from left to right in (D) correspond to: Cl, C, Ti, O, Al, Si, P, S, Ag/Cl, Ag, Ti, T, and in (E) to: C, O, Ti, O, Zn, Al, P, S, Ti, T.

is required or tracing to clinical biosensing and environmental sciences.

### SERS-based sandwich immunoassay for COVID-19 rapid and *in situ* diagnosis

Within this work, we demonstrate the capabilities of our AgNPs@MBA@SiO<sub>2</sub> to enhance the sensitivity of ELISA tests. We took as example the immunogenic SARS-CoV-2 spike protein, responsible for COVID-19 pandemics.

Herewith, we prepared our SERS-based ELISA-like sandwich immunoassay consisting of a substrate functionalized with the capturing antibody, responsible to immobilize the antigen for further recognition by an anti-antigen antibody providing a readout and the diagnostic result. In our case, the readout is the SERS signal provided by the AgNPs@MBA@SiO<sub>2</sub> which recognizes the antigen upon their surface functionalization with the antibody anti-SARS-CoV-2-spike protein, *i.e.*, AgNPs@MBA@SiO<sub>2</sub>@Ab.

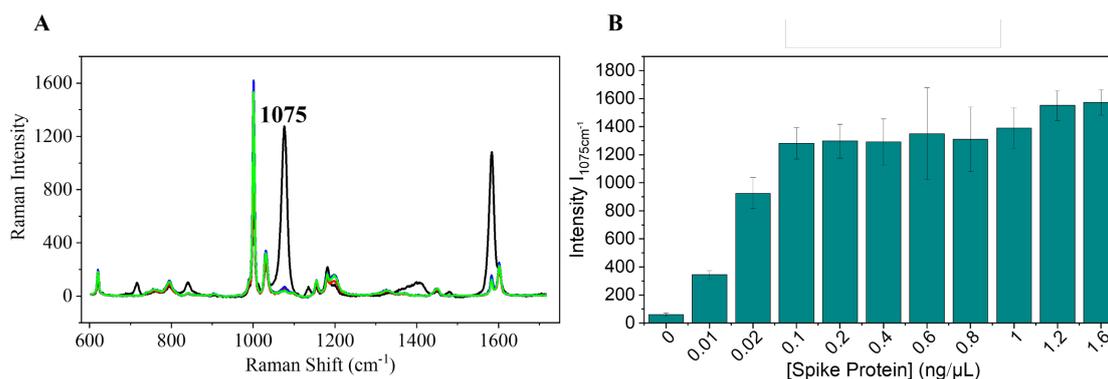
For the surface functionalization of the AgNPs@MBA@SiO<sub>2</sub> with the antibody, we first attached a linker (GPTMS) through a reaction called epoxy-silanization(39). It enriches the NPs' surface with epoxy groups which are highly active towards amino groups. Thus, forming covalent bonds between antibodies and GPTMS-NPs. The obtained AgNPs@MBA@SiO<sub>2</sub>@Ab recognize the immunogenic SARS-CoV-2-spike protein. To confirm NPs' surface bioconjugation, we incubate the AgNPs@MBA@SiO<sub>2</sub>@Ab and the AgNPs@MBA@SiO<sub>2</sub> with a fluorescently labeled secondary antibody. Only

AgNPs@MBA@SiO<sub>2</sub>@Ab provide a bright fluorescent signal originated from the antibody recognition and confirming successful bioconjugation (*cf.*, SI, figure SI-7).

Once we engineered the detecting antibody with a SERS readout, we prepared the ELISA plate by coating the substrate with an anti-SARS-CoV-2 spike protein capturing antibody. Upon addition of the spike protein and rinsing the unbounded molecules, we add our AgNPs@MBA@SiO<sub>2</sub>@Ab. The NPs recognize the antigen protein *via* a second recognition site and are immobilized onto the ELISA plate substrate. This recognition provides a positive SERS signal (enhancement of the 1075 cm<sup>-1</sup> band) thus confirming virus detection (figure 4). The signal was fast (1 s) and specific for the viral protein. Neither the ELISA plate (*cf.*, SI, figure SI-7) nor the absence of antigenic SARS-CoV-2 spike protein (figure 4A) and/or the absence of capturing antibody (figure 4A) provided a SERS positive signal. In the absence of capturing antibody, we increased the concentration of antigen one order of magnitude to detect possible unspecific interactions of the antigen with the plate that could be responsible for false positive results but we did not detect signal. Only when the viral antigen was immobilized, the AgNPs@MBA@SiO<sub>2</sub>@Ab provided a strong SERS enhancement of the specific band at 1075 cm<sup>-1</sup>. The enhancement was 20 times higher than in the control samples.

We further conducted SERS-based ELISA tests with a concentration series of the antigen ranging from 0 to 1.6 ng/μL (figure 4B). The intensity ratio between 0.01 and 0 ng/μL at 1075 cm<sup>-1</sup> was close to 6, revealing that the LOD of AgNPs@MBA@SiO<sub>2</sub>@Ab for antigenic SARS-CoV-2 spike protein detection should be less than 0.01 ng/μL.

We want to mention that when the antigen concentration is higher than 0.1 ng/μL, the response is less sensitive to small changes in analyst's concentration (figure 4B). However, the specificity is maintained since we obtained positive signals all over the antigen concentration range. A reduction in the sensitivity, can be neglected in the cases where a qualitative response is required, like for example for the fast diagnosis of viral infections.



**Figure 4: Functional validation of our SERS-based ELISA test. (A)** Averaged SERS spectra collected from AgNPs@MBA@SiO<sub>2</sub>@Ab in the presence of 0.1 ng/μL antigenic SARS-CoV-2 spike protein (black) and in the absence of antigen (red), capturing antibody (blue) or both (green). In the absence of capturing antibody, the concentration of antigen was increased to 1 ng/μL. **(B)** SERS intensity at 1075 cm<sup>-1</sup> provided by the AgNPs@MBA@SiO<sub>2</sub>@Ab incubated with different amount of antigenic SARS-CoV-2 spike protein (from 0 to 1.6 ng/μL). The results correspond to an average of 8 spectra recorded from 8 random places in the ELISA plate.

Our SERS encoded nanocomposites, AgNPs@MBA@SiO<sub>2</sub>@Ab, offer a robust and stable labelling platform with high sensitivity and selectivity that can be applied for example in sandwich-based diagnostic assays.

## CONCLUSION

We have rationally designed a stable synthetic procedure for SERS encoded plasmonic-silica nanocomposites. We have studied fundamental factors affecting the codification of plasmonic NPs with SERS probe for an enhanced labelling efficiency. In general, weak basic environment facilitates the codification of Raman probe onto metallic surface. Continuous increase of the surrounding pH will favour the dissociation of the metallic-thiol covalent bond. Thus, a precise control over the pH during the codification is critical for an optimized modification of thiolated compound onto metallic surfaces. We also found that ethanol has little effect on the formation of silver thiol bond. The final packing quality of the Raman probe monolayer is affected by the metal-thiol bonding formation speed, the electrostatic repulsion on the metallic surface, and the competition between Raman probe and stabilizer. A precise control over the NPs agglomeration, increases the percentage of hot spots which results in a SERS signal increase of nearly 40 times. This encoded nanoagglomerates were encapsulated and protected by a silica layer which offers the possibility to be multifunctionalized. The nanoagglomerates provide a robust and stable SERS signal with no interference regardless the substrate used. The range of applicability of these nanoagglomerates goes from industrial labeling to environmental science and clinical biosensing, where a fast, *in situ*, sensitive and specific readout is required. We validated the performance of our nanoagglomerates for the diagnosis of COVID-19. We built up a SERS-based ELISA assay for the fast, sensitive, and specific detection of SARS-CoV-2 antigens.

### **Ethics approval and consent to participate**

Not applicable

### **Consent for publication**

All authors have provided their consent to publish this work.

### **Availability of data and materials**

Along with this manuscript, we provide a supporting information file comprising a detailed description of the methodology and further results.

### **Competing interests**

The authors declare that they have no competing interests.

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### **Authors' contributions**

CX acquired the data and made substantial contribution to the analysis and interpretation of data. CX have drafted the work.

VI contributed to the analysis of the data.

PRG designed and conceptualized the idea and made substantial contribution to the analysis and interpretation of data. PRG have wrote the work.

CX, VI, and PRG have approved the submitted version. All have agreed both to be personally accountable for the author's own contributions and to ensure that questions related to the accuracy or integrity of any part of the work, even ones in which the author was not personally involved, are appropriately investigated, resolved, and the resolution documented in the literature.

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### **AUTHORS' INFORMATION (OPTIONAL)**

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