

# Cooperative Cathodes for Enhanced Hexavalent Chromium Reduction and Electricity Generation in Bioelectrochemical Reactor with Simultaneous Sludge Degradation

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## Research Article

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1           **Cooperative Cathodes for Enhanced Hexavalent Chromium**  
2           **Reduction and Electricity Generation in Bioelectrochemical Reactor**  
3           **with Simultaneous Sludge Degradation**

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15

16 **Abstract:** In a typical microbial electrochemical system (MES) with dual-chamber  
17 for Cr(VI) reduction, it was faced with the decline of Cr(VI) reduction efficiency after  
18 several cycles of operation, which limited the continuous and effective Cr(VI)  
19 reduction of MES. In this study, a novel bioelectrochemical reactor assembled with  
20 cooperative cathodes of chemical cathode and bio-cathode (BER<sub>CC</sub>) and with excess  
21 sludge as anodic substrate was designed to solve the problem. Comparative study of  
22 BER<sub>CC</sub> and BER with dual chemical cathodes (BER<sub>DC</sub>) revealed that BER<sub>CC</sub> broke  
23 through the limitation of Cr(VI) reduction decline for cycles of operation and  
24 improved performance of electricity generation. Both the Cr(VI) reduction rate and  
25 the power density were increased with the decrease of pH and the increase of TCOD  
26 and initial Cr(VI) concentration. Cooperative cathodes stimulated the growth of  
27 electrochemically active microorganisms in the anodic biofilm and produced  
28  $8.21 \pm 0.64$  mg Coulomb/(L·h) more electrons than dual chemical cathodes, which  
29 enhanced the electrons for electricity generation and Cr(VI) reduction by about 58.3%  
30 and  $56.1 \pm 5.6\%$  in BER<sub>CC</sub> than those in BER<sub>DC</sub>.

31

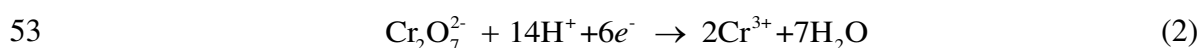
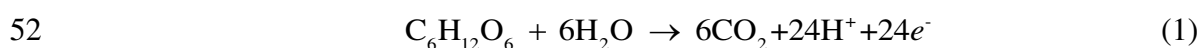
32 **Keywords:** Bioelectrochemical reactor, hexavalent chromium, cooperative cathodes,  
33 excess sludge, electron flux, electrochemically active microorganisms

34

## 35 1. Introduction

36 Microbial electrochemical system (MES) is an emerging platform technology,  
37 which combines electrochemical systems with microbial processes. This technology  
38 has been extensively studied and intensively developed for remediation of wastewater  
39 (Jadhav et al. 2017), soil (Chen et al. 2015), sediment (Wan et al. 2018), CO<sub>2</sub>  
40 (Rodrigues et al. 2019), etc. Remediation of hexavalent chromium [Cr(VI)]  
41 contaminated wastewater has been one of the widely researched fields for MES  
42 (Wang et al. 2008; Kim et al. 2017; Wang et al. 2017; Han et al. 2018; Li and Zhou  
43 2019), where the electrochemically active microorganisms (EAMs) use electrodes as  
44 electron acceptors/donors via extracellular electron transfer and close circuit (Logan  
45 et al. 2006; Shi et al. 2016; Rabaey et al. 2009). Microbial electrochemical Cr(VI)  
46 reduction mainly bases on cathodic reduction with abiotic/biotic cathode, reflecting  
47 the advantages of sustainability and low-cost (Sophia and Saikant 2016).

48  
49 In a typical MES with dual-chamber for Cr(VI) reduction, the anodic reaction  
50 equation (taking glucose as the example of anodic substrate) and the cathodic reaction  
51 equation were shown in Eq.(1) and Eq.(2), respectively (Wang et al. 2008).



54 At the early stages of the reaction process, the initial high concentrations of Cr(VI)  
55 and H<sup>+</sup> can positively affect the cathodic reaction and lead to a high Cr(VI) reduction  
56 rate. High concentration of anodic substrate at early stages also promotes the activity  
57 of EAMs and accelerates the substrate degradation. Thus, lots of electrons can be  
58 produced during organic matter degradation in the anodic chamber to support fast

59 cathodic Cr(VI) reduction, as shown in Eq.(1) and Eq.(2). As the reaction of MES  
60 progresses, however, the consumption of Cr(VI) and H<sup>+</sup> in the cathodic chamber  
61 results in the decline of motivation force from the cathodic reaction. After several  
62 Cr(VI) reduction cycles, the cathodic reduction efficiency gradually decreases, and the  
63 cathodic potential declines slowly with it, which also reduces the activity of EAMs in  
64 the anodic chamber and has a negative influence on the electricity generation finally  
65 (Xafenias et al. 2014). Similar changes were also shown in other studies. In a  
66 dual-chamber microbial fuel cell (MFC) with conductive polymer-mediated Cr(VI)  
67 reduction, Cr(VI) was completely removed for the initial two cycles, but less than  
68 80% of removal efficiency was generated for the 3<sup>rd</sup> cycle, and the removal efficiency  
69 dropped to 60% for the 4<sup>th</sup> cycle (Pang et al. 2013) . In another dual-chamber MFC  
70 with graphene/biofilm composites for Cr(VI) reduction, the removal efficiency was  
71 58±0.5% for the 1<sup>st</sup> cycle, while the value was decreased to 45.2±4% for the 2<sup>nd</sup> cycle,  
72 and the value dropped to 15.5±1.3% for the 3<sup>rd</sup> cycle (Song et al. 2016). In a  
73 four-chamber microbial desalination cell for Cr(VI) reduction and salinity removal,  
74 the decrease of Cr(VI) concentration led to the drop in current density, which was  
75 gradually decreased along each Cr(VI) reduction cycle (An et al. 2014). These studies  
76 showed that MES was faced with the decline of Cr(VI) reduction efficiency after  
77 cycles of operation, which limited the continuous and effective Cr(VI) reduction of  
78 MES.

79

80 Using high organic concentration of anodic substrate is one of the solutions that  
81 supply abundant electrons for continuous and effective Cr(VI) reduction in MES. The  
82 anodic substrate is the electron donor of MES for current generation and cathodic  
83 reaction. Sludge, as a typical substrate with high concentration of organic matters, has

84 been reported to supply electrons for electricity generation steadily in MES. Jiang et  
85 al. (2009) positioned a two-chamber MFC with potassium ferricyanide as catholyte,  
86 which degraded excess sludge with stable and high electricity generation during  
87 operation of 40 h. Meng et al. (2019) invented a solid phase bioelectrochemical  
88 system, providing a method for simultaneous sludge compost, bioelectricity  
89 generation, and desalination with a relatively stable operational period of nearly 151  
90 days (cell voltage > 0.6 V). Hence, MES with sludge as the anodic substrate can  
91 create the condition of sufficient electron donors for stable and high flow electron  
92 transfer. In our previous research, a bioelectrochemical reactor (BER) was constructed  
93 with sewage sludge as the anodic substrate and Cr(VI) as the catholyte, which paved  
94 the way for simultaneous treatment of sewage sludge and wastewater containing  
95 Cr(VI) (Yu et al. 2018).

96

97 Maintaining high activity of EAMs is the other solution that supplies electrons  
98 for continuous and effective Cr(VI) reduction in MES, which results in high  
99 performance of electricity generation. Due to the thermodynamic constraints,  
100 traditional MFC produced 1.14 V as determined by the NADH and pure oxygen redox  
101 potentials. Dual-cathode MFCs with two cathodes share the anode, incorporated an  
102 extra cathodic chamber flanked with the anodic chamber of dual-chamber MFCs and  
103 had higher electricity generation than that of dual-chamber MFCs (Jiang et al. 2011;  
104 Zhang and He 2012). A coupled MFC consisting of an oxic-biocathode MFC and an  
105 anoxic-biocathode MFC was implemented to remove carbon and nitrogen from  
106 synthetic wastewater simultaneously, which obtained a maximum COD removal rate  
107 of 98.8% and reached a power density of 14 W/m<sup>3</sup> net cathodic compartment and 7.2  
108 W/m<sup>3</sup> net cathodic compartment, respectively (Xie et al. 2011). From here we infer

109 that electricity generation can be kept in the state of stable and continuous Cr(VI)  
110 reduction rate by cooperative cathodes, where bio-cathode was coupled to traditional  
111 dual-chamber MES. Bio-cathode not only lowered the cost of construction and  
112 operation of reactors, but also increased the operational sustainability (He and  
113 Angenent 2006; Rabaey and Keller 2008). Hence, coupling of bio-cathode and  
114 chemical cathode can be regarded as the appropriate cooperative cathodes.  
115 Nevertheless, few studies were relative to cooperative cathodes for enhancing the  
116 performance of Cr(VI) reduction in MES.

117

118 To obtain abundant electrons for continuous and effective Cr(VI) reduction in  
119 MES, we designed a novel BER assembled with cooperative cathodes of chemical  
120 cathode and bio-cathode as shown in Fig. 1a. Under this configuration with excess  
121 sludge as electron donors and both chemical cathode and bio-cathode as electron  
122 acceptors, which can keep the high performance of BER. The performances of BER  
123 with cooperative cathodes (BER<sub>CC</sub>) and BER with dual chemical cathodes (BER<sub>DC</sub>)  
124 were compared, to substantiate the feasibility of keeping high efficiency by  
125 cooperative cathodes of BER<sub>CC</sub>. Afterwards, effects of the major impact factors [viz.,  
126 Cr(VI) concentration and pH of catholyte and total chemical oxygen demand (TCOD)  
127 of excess sludge] on the performance of BER<sub>CC</sub> were investigated. In addition, the  
128 distribution of electron flux and the structure of microbial community within BER<sub>DC</sub>  
129 and BER<sub>CC</sub> were analyzed. This study provides alternative technology for the  
130 continuous and efficient remediation of Cr(VI) contaminated wastewater and  
131 treatment of excess sludge.

132

## 133 2. Materials and methods

### 134 2.1 Configuration of BER<sub>CC</sub> and BER<sub>DC</sub>

135 The bioelectrochemical reactor with cooperative cathodes (BER<sub>CC</sub>) consists of a  
136 cylindrical anodic chamber ( $\Phi$  80 mm×90 mm) and two cubic cathodic chambers (60  
137 mm×70 mm×90 mm each) (Fig. 1a). The effective volumes of the anodic chamber  
138 and cathodic chamber in the BER<sub>CC</sub> were 340 mL and 300 mL, respectively. Proton  
139 exchange membrane (PEM) (Nafion™117, DuPont Co., USA) separated the anodic  
140 chamber from the chemical cathodic chamber for preventing the Cr<sup>3+</sup> produced in the  
141 chemical cathodic chamber from entering the anodic chamber, and the cation  
142 exchange membrane (CEM) (AMI-7001, Ultrex, USA) separated the anodic chamber  
143 from the bio-cathodic chamber in consideration of cost. All of the anode and cathodes  
144 were made of graphite fibre brushes ( $\Phi$  70 mm×70 mm for the anode and  $\Phi$  60  
145 mm×70 mm for both cathodes) and titanium wires. The BER with dual chemical  
146 cathodes (BER<sub>DC</sub>) possesses the same configuration with BER<sub>CC</sub> as shown in Fig. 1b,  
147 which was applied as the control for experiments. Both chemical cathodic chambers  
148 were separated by PEMs from the anodic chamber. The BER<sub>CC</sub> at open circuit was  
149 used as the control to reflect the role of adsorption for Cr(VI) removal.

150

151 Fig.1

152

### 153 2.2 Inoculation and operation

154 Excess sludge was used as the inoculum and the anodic substrate of the anodes  
155 of BER<sub>CC</sub> and BER<sub>DC</sub>, which was collected from the Harbin municipal wastewater  
156 treatment plant. The total chemical oxygen demand (TCOD), soluble chemical



157 oxygen demand (SCOD) and moisture content of the excess sludge were  $22307 \pm 694$   
158 mg/L,  $124.5 \pm 0.6$  mg/L,  $97.1 \pm 0.5\%$ , respectively. For the start-up, both the inoculum  
159 (excess sludge) for the anode and catholyte (potassium dichromate of 0.1 mol/L) for  
160 the cathodes were both filled at the height of 70 mm and replaced every three days  
161 until stable voltage was obtained in the reactor (Aelterman et al. 2006).

162

163 After start-up, the catholyte for the chemical cathodes of BER<sub>CC</sub> and BER<sub>DC</sub> was  
164 changed to the aqueous solution prepared by dissolving K<sub>2</sub>Cr<sub>2</sub>O<sub>7</sub> in deionized water at  
165 a pH of 2 (Wang et al. 2008). The catholyte for the bio-cathode of BER<sub>CC</sub> included  
166 nutritional salts, inoculated bacteria, and dissolved oxygen. The nutritional salts were  
167 prepared as previously described (Rabaey et al. 2005). The inoculated bacteria were  
168 collected from the catholyte of the bio-cathode of an MES in a good operating state.  
169 Dissolved oxygen was continuously generated by the aerator at 200 mL/min (Fig. 1a).  
170 All the experiments were in the mode of sequencing batch at  $25 \pm 1^\circ\text{C}$ . All of the  
171 samples were analyzed in triplicate.

172

### 173 2.3 Analysis and calculation

174 The pH value and concentration of Cr(VI) of the catholyte and TCOD and SCOD  
175 of excess sludge were measured based on the Standard Methods (APHA 2015; Yu, et  
176 al. 2018). The precipitate produced during Cr(VI) reduction was observed by the  
177 scanning electron microscope energy dispersive X-ray analysis (SEM-EDX) and  
178 X-ray photoelectron spectroscopy (XPS). The voltage of BER<sub>CC</sub> and BER<sub>DC</sub> was  
179 measured with an external resistance of 1000  $\Omega$  by a voltage collector. Polarization  
180 curves were calculated according to the previous study (Logan and Regan 2006), and  
181 the effective volume of the anodic chamber was used to calculate the power density

182 (Jiang et al. 2009). The production of methane and hydrogen was measured by  
183 titration and off-gas analysis (TOGA) sensor (Pratt et al. 2003; Virdis et al., 2009).  
184 The qualification of electron fluxes was realized as previously studied (Yu et al.  
185 2018).

186

187 During the period of operation, a small amount of graphite fibres were collected  
188 from the anodes of BER<sub>CC</sub> and BER<sub>DC</sub> and the bio-cathode of BER<sub>CC</sub> for DNA  
189 extraction. Total genomic DNA was extracted using a DNA Isolation Kit. The V3-V4  
190 region of the bacterial 16S rRNA gene was amplified by the universal primers 341F  
191 (5'-CCCTACACGACGCTCTTCCGATCTG-3') and 805R  
192 (5'-GACTGGAGTTCCTTGGCACCCGAGAATTCCA-3') to analyze the structures  
193 of the bacterial communities. The sequences were computed with a similarity of 97%.  
194 Chao1 and Shannon index were used to estimate the levels of alpha diversity.

195

### 196 **3. Results and discussion**

#### 197 3.1 Performances of BER<sub>CC</sub> and BER<sub>DC</sub>

198 To reveal the advantages of cooperative cathodes on hexavalent chromium  
199 reduction, electricity generation, and sludge degradation, the performances of BERs  
200 with cooperative cathodes (BER<sub>CC</sub>) and chemical dual-cathodes (BER<sub>DC</sub>) were  
201 compared.

202

##### 203 3.1.1 Performance of hexavalent chromium reduction

204 As shown in Fig. 2a, Cr(VI) was almost reduced completely within 55.5 h and  
205 57.5 h in BER<sub>CC</sub> and BER<sub>DC</sub> for the 1<sup>st</sup> Cr(VI) reduction cycle, respectively. After it,  
206 the Cr(VI) reduction rates of BER<sub>DC</sub> were gradually decreased, and it took 102.5 h

207 and 66.0 h for BER<sub>DC</sub> to reduce Cr(VI) by 72.5±2.8% and 52.1±1.8%, respectively.  
208 The Cr(VI) reduction efficiency dropped to 39.9±1.3% for the 5<sup>th</sup> cycle. In another  
209 dual-chamber MFC, the Cr(VI) concentration was initially decreased rapidly and  
210 gradually slowed down after four cycles of operation (Xafenias et al. 2014). An et al.  
211 (2014) studied the microbial desalination cell (MDC) to reduce Cr(VI) at cathode, and  
212 the current densities of the MDCs were also gradually decreased from the 1<sup>st</sup> cycle to  
213 the 10<sup>th</sup> cycle. While the Cr(VI) reduction efficiency of BER<sub>CC</sub> reached approximately  
214 100% after 58.5 h and 60.5 h during the 2<sup>nd</sup> and the 3<sup>rd</sup> cycle, respectively. Based on  
215 the above results, Cr(VI) could be entirely reduced for seven cycles in BER<sub>CC</sub>,  
216 however, it took much longer time to reduce Cr(VI) for BER<sub>DC</sub> than that for BER<sub>CC</sub>  
217 (Fig. 2a). The variation of the Cr(VI) concentration in the BER<sub>CC</sub> at open circuit  
218 revealed that adsorption was non-significant in the BER<sub>CC</sub> (Fig. S1). Similar results  
219 were also found by Li et al. (2018). The reduction products (dark green precipitates)  
220 of Cr(VI) were found attached to the chemical cathode and sank to the bottom in  
221 BER<sub>CC</sub> (Fig. S2a and Fig. S2b). Both the main constituent elements of the precipitates  
222 at the bottom of the cathodic chamber and that attached to the surface of the chemical  
223 cathode were Cr (Fig. S2 and Table S1). The main component of the precipitate in the  
224 cathodic chamber was chromium oxide (Fig. S3).

225

226 Fig. 2

227

### 228 3.1.2 Performance of electricity generation and sludge degradation

229 During seven Cr(VI) reduction cycles, the voltage of BER<sub>CC</sub> was shown  
230 consistently above 0.672 V, and the maximum voltage reached about 0.813 V at the  
231 beginning of each cycle in Fig. 2b. While BER<sub>DC</sub> produced high voltage only at the

232 beginning stage of the 1<sup>st</sup> cycle, and the voltage was gradually decreased with the  
 233 Cr(VI) reduction proceeded. During the 2<sup>nd</sup> reduction cycle of BER<sub>DC</sub>, the voltage  
 234 was first increased slightly compared with that at the end of the 1<sup>st</sup> cycle, but it was  
 235 gradually declined with the proceeding of Cr(VI) reduction. The tendency of voltage  
 236 for the 5<sup>th</sup> cycle was similar to that for the previous cycles. After five cycles of a  
 237 dual-chamber MFC with alumina-nickel nanoparticles-dispersed carbon nanofiber  
 238 electrode, the performance of the MFC was decreased by less than 10% (Gupta et al.  
 239 2017). After 432 h of operation, the TCOD of sludge decreased by 73.6±4.4% from  
 240 27692±1662 mg/L to 7307±438 mg/L in the anodic chamber of BER<sub>CC</sub> and  
 241 52.1±3.1% from 27692±1662 mg/L mg/L to 13267±796 mg/L in the anodic chamber  
 242 of BER<sub>DC</sub>, respectively. Of the electrons produced in organic matter degradation, a  
 243 great amount were used for electricity generation, and this process could accelerate  
 244 the organic matter degradation of sludge in BES (Xafenias et al. 2014). BER<sub>CC</sub> can  
 245 consume electrons continuously produced from the degradation of sludge. Therefore,  
 246 that BER<sub>CC</sub> had a higher performance of electricity generation was the main reason  
 247 that the TCOD degradation of BER<sub>DC</sub> was higher than that of BER<sub>CC</sub> (ref. 3.1.2).

248

249 The differences between the performances of BER<sub>CC</sub> and BER<sub>DC</sub> in electricity  
 250 generation and Cr(VI) reduction can be explained based on the electrochemical theory.  
 251 According to Eq.(1), the anodic potential declined with the degradation of sludge as  
 252 the Eq.(3),

$$E_{\text{ano}} = E_{\text{ano}}^0 - \frac{RT}{nF} \ln[\text{H}^+]^{24} \quad (3)$$

254 where  $E_{\text{ano}}$  is the cathode potential,  $E_{\text{ano}}^0$  stands for the standard cathode potential  
 255 (V),  $R$  is the ideal gas constant (8.314 J/mol K),  $T$  is the temperature (K),  $n$  is the

256 number of transferred electrons,  $F$  is the Faraday's constant,  $[H^+]$  is the concentrations  
 257 of  $H^+$ . As Cr(VI) was reduced by the chemical cathode of BER<sub>DC</sub>, the Cr(VI)  
 258 concentration gradually decreased. According to Eq.(2), the electrons that transferred  
 259 from the anode and participated in the chemical cathodic reaction were decreased, and  
 260 the Cr(VI) reduction rate was descending slowly with it. The cathodic potential  
 261 declined with the slowdown of Cr(VI) reduction rate as the Eq.(4),

$$262 \quad E_{\text{cat}} = E_{\text{cat}}^0 - \frac{RT}{nF} \ln \frac{[Cr^{3+}]^2}{[Cr_2O_7^{2-}][H^+]^{14}} \quad (4)$$

263 where  $E_{\text{cat}}$  is the cathode potential,  $E_{\text{cat}}^0$  is the standard cathode potential (V),  $[Cr^{3+}]$   
 264 and  $[Cr_2O_7^{2-}]$  are the concentrations of Cr(III) and Cr(VI), respectively. As indicated  
 265 in Eq.(5),

$$266 \quad E = E_{\text{cat}} - E_{\text{ano}} \quad (5)$$

267 The voltage of BER<sub>DC</sub> was decreased with the operation, and the reaction of  
 268 extracellular electron transfer at anode slowed down. At the beginning of the next  
 269 cycle, the Cr(VI) reduction rate and cathode potential were nearly recovered at the  
 270 beginning of the next cycle, however, the reaction rate of the extracellular electron  
 271 transfer could not recover completely. As a consequence, the voltage of BER<sub>DC</sub>  
 272 gradually declined. Relevant results revealed that the decrease in performance of MES  
 273 was attributed to the reduction of the catalytic activity of the electrode (Xafenias et al.  
 274 2014), which was consistent with the results of this study. On the condition of the  
 275 same electrons produced at the anode and transferred to the cathode(s), though the  
 276 electrons that reacted at chemical cathode decreased with the decline of Cr(VI)  
 277 concentration in BER<sub>CC</sub>, the electrons that reacted at the bio-cathode increased with it.  
 278 Therefore, the anodic reaction could be kept at a high rate, and the voltage could be  
 279 kept at a high level. A certain number of rod-shaped EAMs existed on the surface of

280 the anode, and the biofilm could be found on the bio-cathode (Fig. S1c and Fig. S1d).  
281 They indicated that the EAMs maintained high activity with the cooperative cathodes  
282 of chemical cathode and bio-cathode.

283

284 In summary, BER<sub>CC</sub> with cooperative cathodes not only broke through the  
285 limitation of Cr(VI) reduction decline during multiple cycles of operation, but also  
286 produced high voltage continuously and improved sludge degradation in contrast to  
287 BER<sub>DC</sub>.

288

### 289 3.2 Impact of TCOD, Cr(VI) concentration and pH on the performance of BER<sub>CC</sub>

290 The advantages of BER<sub>CC</sub> on performance were preliminarily studied (ref. 3.1).  
291 TCOD of excess sludge, and initial Cr(VI) concentration and pH of catholyte on the  
292 performance were investigated to reflect the operation characteristics of BER<sub>CC</sub> under  
293 different conditions.

294

#### 295 3.2.1 Impact of TCOD on the performance of BER<sub>CC</sub>

296 Excess sludge contains abundant organic matters, so that it can provide sufficient  
297 electron source for Cr(VI) reduction and electricity generation of BER<sub>CC</sub>. Therefore,  
298 the TCOD of the anodic substrate is not the limiting factor of the performance when  
299 BER<sub>CC</sub> is fueled by excess sludge. The variations of Cr(VI) reduction in BER<sub>CC</sub> at  
300 different TCOD (TCOD<sub>1</sub>=10769±321 mg/L, TCOD<sub>2</sub>=22307±694 mg/L, and TCOD<sub>3</sub>  
301 =29692±885 mg/L) of sludge were shown in Fig. 3a, which revealed that the more the  
302 TCOD in the anodic chamber, the higher the rate of Cr(VI) reduction in the chemical  
303 cathodic chamber. Fig. 3b showed the polarization curves and power density curves of  
304 BER<sub>CC</sub> at different TCOD of sludge. After calculation, the internal resistance of

305 BER<sub>CC</sub> with sludges of TCOD1, TCOD2 and TCOD3 were 151.3±5.7 Ω, 121.5±6.7 Ω,  
 306 140.2±7.4 Ω, respectively, since biofilms became thick gradually and the transfer  
 307 resistance increased at a high level of TCOD, which might have negative effects on  
 308 the performance of electricity generation at high TCOD level. Based on the  
 309 electrochemical theory, the actual cathodic potential that takes losses into  
 310 consideration was obtained as follows:

$$311 \quad E_{\text{cat}} = E_{\text{cat}}^0 - \frac{RT}{nF} \ln \frac{[\text{Cr}^{3+}]^2}{[\text{Cr}_2\text{O}_7^{2-}][\text{H}^+]^{14}} - (\eta_{\text{oct}} + \eta_{\text{ohm}} + \eta_c) \quad (6)$$

312 Where  $\eta_{\text{oct}}$ ,  $\eta_{\text{ohm}}$ , and  $\eta_c$  stand for the activate loss(V), ohm loss(V), and mass transfer  
 313 loss(V), respectively. The cathodic potential is determined by the concentrations of  
 314 Cr(VI) and Cr(III) and pH together [Eq.(6)]. As Fig. 3b shown, both the open-circuit  
 315 voltage and the maximum power density were increased gradually with the increase  
 316 of TCOD.

317

318 Fig. 3

319

### 320 3.2.2 Impact of initial Cr(VI) concentration on the performance of BER<sub>CC</sub>

321 On the condition of TCOD=29692±885 mg/L with the highest performance of  
 322 Cr(VI) reduction (ref. 3.2.1), the performances of Cr(VI) reduction in BER<sub>CC</sub> at  
 323 different initial Cr(VI) concentrations were shown in Fig. 4a. The rate of Cr(VI)  
 324 reduction was increased with the increase of initial Cr(VI) concentration, since high  
 325 initial Cr(VI) concentration was benefit for the improvement of reduction efficiency  
 326 [ref. Eq.(2)]. However, the reduction rate at an initial concentration of 200 mg/L was  
 327 less than twice that at 100 mg/L. The possible reason was that the reaction of Cr(VI)  
 328 reduction was inhibited, as H<sup>+</sup> was gradually consumed according to Eq.(2). As

329 calculation according to Fig. 4b, the internal resistance for 60 mg/L, 100 mg/L, 200  
330 mg/L were  $142.7 \pm 3.5 \Omega$ ,  $122.3 \pm 4.2 \Omega$ , and  $117.1 \pm 2.9 \Omega$  for 60 mg/L, 100 mg/L, 200  
331 mg/L, respectively. Li et al. studied a dual-chamber MFC with potassium dichromate  
332 as the electron acceptor. They found that the internal resistance decreased from  $300 \Omega$   
333 to  $100 \Omega$  when the Cr (VI) concentration was increased from 50 mg/L to 500 mg/L (Li  
334 et al. 2009). There is no significant difference between the anodic potentials of BER<sub>CC</sub>  
335 at different initial Cr(VI) concentrations, which were kept about -0.20 V (vs. SHE)  
336 during the reaction cycles. Cathodic potentials were increased following the growth of  
337 the initial Cr(VI) concentration. Wang et al. studied that the cathodic potential  
338 increased from 600 mV to 650 mV when the initial Cr (VI) concentration increased  
339 from 25 mg/L to 200 mg/L, because the increased Cr(VI) concentration would  
340 decrease the internal resistance of the MDC by enhancing the ionic strength and thus  
341 boost the voltage of the external resistance (Wang et al. 2008). Fig. 4b showed that the  
342 performances of Cr(VI) reduction rate and electricity generation were the highest  
343 ( $7.2 \pm 0.6 \text{ W/m}^3$ ) at the initial Cr(VI) concentration of 200 mg/L.

344

345 Fig. 4

346

### 347 3.2.3 Impact of pH on the performance of BER<sub>CC</sub>

348 At TCOD of  $29692 \pm 885 \text{ mg/L}$  and initial Cr(VI) concentration of 200 mg/L with  
349 the highest performance of Cr(VI) reduction (ref. 3.2.1 and 3.2.2), the performances  
350 of Cr(VI) reduction in BER<sub>CC</sub> at different initial pHs were shown in Fig. 5a. During  
351 the 1<sup>st</sup> Cr(VI) reduction cycle, the lower the pH value, the faster the reduction rate and  
352 the higher the reduction efficiency. The reduction rates and reduction efficiencies of  
353 BER at pH=1 and pH=2 were all higher than those at pH=4 [ref. Eq.(2)]. During the



354 2<sup>nd</sup> Cr(VI) reduction cycle, the reduction rate of BER<sub>CC</sub> was first higher at pH=1 than  
355 that of at pH=2. After a period of time, the reduction rate decreased slower at pH=1  
356 than that at pH=2. In combination, the Cr(VI) reduction rate of each BER<sub>CC</sub> was in the  
357 order of pH 1 > pH 4 > pH 2. With the decrease of pH value, the internal resistance  
358 was gradually declined as indicated in Fig. 5b. The maximum open-circuit voltage  
359 were 1.054±0.045 V, 0.991±0.034 V, 0.887±0.036 V, and the maximum power density  
360 were 5.2±0.4 W/m<sup>3</sup>, 7.2±0.4 W/m<sup>3</sup>, 9.6±0.5 W/m<sup>3</sup> for pH 1, pH 2, and pH 4,  
361 respectively. The results indicated that the electrons were utilized much at low pH  
362 with high voltage and power density obtained. An [et al. \(2014\)](#) studied an MDC with  
363 synthetic Cr(VI) containing wastewater as the catholyte. When the pH grew up, both  
364 the current density and desalination efficiency were decreased rapidly. Li [et al. \(2018\)](#)  
365 used dual-chamber MFCs to reduce Cr(VI) and generate bioelectricity simultaneously.  
366 They found that Cr(VI) reduction was fitted to the pseudo second-order model with  
367 the order of rate constant pH 2 > pH 3 > pH 4 > pH 5 > pH 1 > pH 6 > pH 7 .

368

369 Fig. 5

370

### 371 3.3 Quantification of the electron fluxes

372 Analysis of electron flux could reveal the advantages of cooperative cathodes in  
373 the production and utilization of electrons within BER<sub>CC</sub>. In the anodic chamber,  
374 electrons were produced from organic matter degradation. Electron fluxes were  
375 distributed among different electron sinks (viz., current generation, exoelectrogen  
376 growth, hydrogen gas production, methanogen growth, and methane production)  
377 based on the electron balances and estimated (exoelectrogen and methanogen) growth  
378 rates in the BER as the previous study reported ([Yu et al. 2018](#)). Most of the electrons

379 for current generation were transferred along the anode to the cathode and participated  
380 in the cathodic reaction with a small amount of loss.

381

382 During two Cr(VI) reduction cycles of BER<sub>DC</sub>, the average TCOD degradation  
383 rate was  $18.42 \pm 4.03$  mg C/(L·h) [viz., mg Coulombs/(L·h)] as shown in [Table 1](#),  
384 which contained  $7.23 \pm 1.94$  mg C/(L·h) for methane production,  $9.59 \pm 0.43$  mg C/(L·h)  
385 for biomass (exoelectrogen and methanogen) growth, and the portions for the average  
386 current production of 0.48 mA and Cr(VI) reduction rate of  $1.07 \pm 0.11$  mg C/(L·h). Of  
387 the electrons produced in the anodic chamber at a unit time as shown in [Fig. 6](#), the  
388 proportion of electrons for methanogens growth, hydrogen production, methane  
389 production, and current generation (transferred along the anode) were  $23.5 \pm 2.7\%$ ,  
390  $6.4 \pm 0.8\%$ , and  $33.3 \pm 3.5\%$ ,  $13.3 \pm 1.7\%$ , respectively.  $77.0 \pm 6.9\%$  of the electrons that  
391 transferred along the anode moved to chemical cathodes for Cr(VI) reduction. In  
392 contrast, the average TCOD degradation rate was  $8.21 \pm 0.64$  mg C/(L·h) more in  
393 BER<sub>CC</sub> than that in BER<sub>DC</sub>, and the electrons for the average current were increased  
394 by 0.28 mA, respectively ([Table 1](#)). These results were consistent with those in [3.1.2](#)  
395 and [3.1.3](#).

396

397 For each cycle in BER<sub>CC</sub>, the Cr(VI) concentration gradually decreased with the  
398 reduction of Cr(VI) by the chemical cathode, the electrons for chemical cathode were  
399 therefore reduced, and excess electrons transferred from the anode were used for  
400 bio-cathode. Cooperative cathodes supported the high activity of EAMs ([Fig. S1](#))  
401 instead of gradual reduction, so the Cr(VI) reduction could be continuously and  
402 efficiently maintained ([Fig. 2](#)) at the chemical cathode for the next cycle. During two  
403 operation cycles, quantification of the electron fluxes revealed that  $8.21 \pm 4.03$  mg

404 C/(L·h) more electrons were produced from organic matter degradation in BER<sub>CC</sub>  
405 than those in BER<sub>DC</sub>. Of them, electrons for Cr(VI) reduction was increased by  
406 56.1±5.6%, and some electrons supported the current generation increased by 0.28  
407 mA. The electrons for chemical cathodic reaction and bio-cathodic reaction (and loss)  
408 were increased by 51.5±4.6% and 41.1±3.7% in BER<sub>CC</sub> than those in BER<sub>DC</sub>,  
409 respectively (Fig. 6).

410

411 Table 1

412

413 Fig. 6

414

#### 415 3.4 Distribution of microbial community

416 Microbial community analysis was to further explore why more electrons were  
417 generated by sludge in the anodic chamber and utilized for electricity generation and  
418 Cr(VI) reduction within BER<sub>CC</sub> than BER<sub>DC</sub>.

419 By the metagenomics sequencing, optimal sequences of 40993, 25543, and  
420 35867 were obtained from the biofilms on the anode of BER<sub>CC</sub>, the bio-cathode of  
421 BER<sub>CC</sub>, and the anode of BER<sub>DC</sub>. These samples can also be divided into 4614, 4157,  
422 and 1580 OTUs by difference, respectively. The theoretical maximum OTUs assessed  
423 by Chao1 were 12168, 9217, 12384 for the anode of BER<sub>CC</sub>, bio-cathodes of BER<sub>CC</sub>,  
424 and the anode of BER<sub>DC</sub>, respectively. This result revealed that the bio-cathode of  
425 BER<sub>CC</sub> had the highest species richness. Shannon index provides information on  
426 species richness and distribution in the community. The order of the diversity  
427 distribution was the anode of BER<sub>CC</sub> (Shannon=6.633) > the bio-cathode of BER<sub>CC</sub>  
428 (Shannon=6.224) > the anode of BER<sub>DC</sub> (Shannon=4.172). On the other hand, the

429 results of Simpson index also revealed that the diversity of the biofilm on the anode of  
430 BER<sub>CC</sub> (Simpson=0.004) was higher than that on the bio-cathode of BER<sub>CC</sub>  
431 (Simpson=0.006) and that on the anode of BER<sub>DC</sub> (Simpson=0.042).

432

433 As shown in [Fig. 7](#), 16S rRNA microbial community structures of the biofilms  
434 on the anode of BER<sub>CC</sub>, the bio-cathode of BER<sub>CC</sub>, and the anode of BER<sub>DC</sub> were  
435 analyzed and classified based on the metagenomics sequencing technology. All of the  
436 three communities exhibited high diversity, where 36, 30, and 21 phyla were found in  
437 the biofilms on the anode of BER<sub>CC</sub>, the bio-cathode of BER<sub>CC</sub>, and the anode of  
438 BER<sub>DC</sub>, respectively ([Fig. 7a](#)). The most significant difference was due to the different  
439 proportions of Proteobacteria, Bacteroidetes, Firmicutes, Chloroflexi, Planctomycetes,  
440 and Actinobacteria in their communities. A sum of the five phyla accounted for 85.0%  
441 (anode of BER<sub>CC</sub>), 77.5% (bio-cathode of BER<sub>CC</sub>), and 98.6% (anode of BER<sub>DC</sub>),  
442 respectively. Chloroflexi had the lowest proportion (0.1%) in the community of  
443 biofilm on the bio-cathode of BER<sub>CC</sub> and the highest proportion (7.8%) in the  
444 community of biofilm on the anode of BER<sub>CC</sub>. Chloroflexi was enriched selectively in  
445 the biofilm on the anode of BER<sub>CC</sub>, which revealed that Chloroflexi might have the  
446 function of transferring electrons to the anode. In the community on the anodic  
447 biofilm of an MFC that operated at high temperature, Chloroflexi accounted for 12%,  
448 and some bacteria belonging to Firmicutes were also verified to generate electricity  
449 ([Wrighton et al. 2008](#)). Chloroflexi was also found predominant in the anodic biofilm  
450 of MFC with cellulose as a substrate ([Ishii et al. 2008](#)). Though most well-known  
451 EAMs (viz., *Geobacter sp.* and *Shewanella sp.*) belong to Proteobacteria,  
452 Proteobacteria were richer in the biofilms on the anode of BER<sub>CC</sub> (74.2%) than those  
453 on the bio-cathode of BER<sub>CC</sub> (49.7%) and those on the anode of BER<sub>DC</sub> (48.8%).

454 Bacteroidetes are wide in distribution scope and function, and Proteobacteria and  
455 Bacteroidetes have a close relationship with hydrolysis, acidity, and acetogenesis  
456 (Campanaro et al. 2016). Planctomycetes and Actinobacteria were abundant in the  
457 biofilms on the bio-cathode of BER<sub>CC</sub>, and Planctomycetes are mainly distributed in  
458 saltwater.

459

460 Fig. 7

461

462 The distribution of microbial community was shown in Fig. 7b, and 55, 61, and  
463 36 classes were found in the biofilms on the anode of BER<sub>CC</sub>, the bio-cathodes of  
464 BER<sub>CC</sub>, and the anode of BER<sub>DC</sub>, respectively, which were mainly distributed to 29  
465 classes. The biofilms on the anode of BER<sub>CC</sub> and BER<sub>DC</sub> had similar bacterial  
466 community. The biofilm on the anode of BER<sub>CC</sub> was concentrated in the classes of  
467  $\beta$ -proteobacteria (phylum of Proteobacteria),  $\delta$ -proteobacteria (phylum of  
468 Proteobacteria),  $\gamma$ -proteobacteria (phylum of Proteobacteria),  $\alpha$ -proteobacteria  
469 (phylum of Proteobacteria), Sphingobacteriia (phylum of Bacteroidetes), Clostridia  
470 (phylum of Firmicutes), and Anaerolineae (phylum of Chloroflexi). The community  
471 of the biofilm on the anode of BER<sub>DC</sub> contained  $\gamma$ -proteobacteria (phylum of  
472 Proteobacteria),  $\beta$ -proteobacteria (phylum of Proteobacteria),  $\alpha$ -proteobacteria  
473 (phylum of Proteobacteria), Sphingobacteriia (phylum of Bacteroidetes),  
474 Actinobacteria (phylum of Actinobacteria) and Bacilli (phylum of Firmicutes).  
475  $\gamma$ -proteobacteria play important roles in the degradation of fatty acid, and the  
476 digestion process (Campanaro et al. 2016). Clostridia is the functional classes during  
477 the process of hydrolysis, acidity, and acetogenesis (Campanaro et al. 2016).  
478  $\beta$ -proteobacteria are important in the utilization of sugar alcohol (Campanaro et al.

479 2016). The proportions of  $\delta$ -proteobacteria, Clostridia, and Anaerolineae in the  
480 biofilms on the anodes of BER<sub>CC</sub> and BER<sub>DC</sub> were 9.5~17, 38~54, and 85~212 times  
481 of that on the bio-cathode of BER<sub>CC</sub>, respectively, which reflected the selective  
482 distribution of EAMs on the anode.  $\delta$ -proteobacteria was abundant in the anodic  
483 biofilm of MES with high electricity generation (Lee et al. 2003; Bond et al. 2002),  
484 indicating that bacteria in relation to iron reduction and electricity generation might be  
485 present. Some bacteria belonging to Clostridia with the bioelectrochemical activity  
486 and the character of Fe(III) reduction was isolated from MFCs (Park et al. 2003).

487

488 At the level of genus (Fig. 7c), the predominant genera in the anodic biofilm of  
489 BER<sub>CC</sub> were *Rhodocyclaceae* with the proportion of 16.1%, *Comamonadaceae* with  
490 the proportion of 4.4%, and *Burkholderiales\_incertae\_sedis* with the proportion of  
491 2.2% (the class of  $\beta$ -proteobacteria), *Geobacteraceae* with the proportion of 3.7% and  
492 *Polyangiaceae* with the proportion of 2.7% (the class of  $\delta$ -proteobacteria),  
493 *Xanthomonadaceae* with the proportion of 3.5% (the class of  $\gamma$ -proteobacteria),  
494 *Saprospiraceae* with the proportion of 5.0% and *Chitinophagaceae* with the  
495 proportion of 4.1% (the class of Sphingobacteria), *Anaerolineaceae* with the  
496 proportion of 4.4% (the class of Anaerolineae), *Nitrospiraceae* with the proportion of  
497 3.4% (the class of Nitrospira), and *Planctomycetaceae* with the proportion of 3.2%  
498 (the class of Planctomycetacia). *Rhodocyclaceae* participates in the degradation of  
499 fatty acid, sulfate reduction in the process of hydrolysis and acidification, the  
500 metabolism of butyrate and propionate, and denitrification, which is the main bacteria  
501 of anodic biofilm without the impacts of aerobic environment (Martins et al. 2010;  
502 Sun et al. 2010). *Comamonadaceae* is abundant in the anodic biofilm that can oxidize  
503 the short-chain fatty acids and degrade cellulose (Rismaniyazdi et al. 2007), and it

504 could transfer electrons to the anode in MFC with organic matters and glucose as  
505 substrate (Chaudhuri and Lovly 2003). *Burkholderiales\_incertae\_sedis* has  
506 relationship with sugar alcohol utilization, degradation of fatty acid, sulfate recovery,  
507 butyrate metabolism, propionate metabolism, and denitrification. *Geobacteraceae*, as  
508 well-known EAMs, can transfer electrons to the anode directly. *Xanthomonas* of the  
509 class  $\gamma$ -proteobacteria is EAMs, which was found in MFCs under different operation  
510 conditions (Yu et al. 2015). *Saprospiraceae* had the function of hydrolysis in the BES  
511 with excess sludge as anodic fuel (Zhang et al. 2012). *Anaerolineaceae* and *Geobacter*  
512 were found the most abundant in the anodic biofilm of plant MFC (Lu et al. 2015).  
513 *Anaerolineaceae* could transform saccharides of small molecules into short-chain  
514 fatty acids and hydrogen (Lu et al. 2015). *Planctomycetaceae* was also found  
515 abundant in MEC (Cerrillo et al. 2016). In BER<sub>DC</sub>, the predominant genera included  
516 *Anaerolineaceae* (the class of Anaerolineae), *Saprospiraceae* (the class of  
517 Sphingobacteria), *Comamonadaceae* (the class of  $\beta$ -proteobacteria), *Rhodocyclaceae*  
518 (the class of  $\beta$ -proteobacteria), *Geobacter* (the class of  $\beta$ -proteobacteria) and  
519 *Chitinophagaceae* (the class of Sphingobacteria). The relative abundances of  
520 *Comamonadaceae*, *Geobacteraceae*, and *Xanthomonadaceae* were higher in the  
521 anodic biofilm of BER<sub>CC</sub> than those of BER<sub>DC</sub>, which implied that BER<sub>CC</sub> has a  
522 higher performance of electricity generation. This result was in accordance with that  
523 in 3.1.2. Many EAMs were found in the anodic biofilms of BERs as shown in Table 2.  
524 However, they were rarely present in the bio-cathode of BER<sub>CC</sub> except for  
525 *Comamonas denitrificans*. *Geobacter* was the most abundant EAM in the anodic  
526 biofilm of BERs, but the typical EAM of *Shewanella* was not detected.

527

528 

Table 2
---------

529

530 The genera in the biofilm of bio-cathode in BER<sub>CC</sub> were mainly  
531 *Xanthomonadaceae* (the class of  $\gamma$ -proteobacteria), *Moraxellaceae* (the class of  
532  $\gamma$ -proteobacteria), *Comamonadaceae* (the class of  $\beta$ -proteobacteria) *Alcaligenaceae*  
533 (the class of  $\beta$ -proteobacteria), *Bacillales\_Incertae\_Sedis\_XII* (the class of  
534  $\beta$ -proteobacteria) , *Sphingobacteriaceae* (the class of Sphingobacteria)  
535 *Chitinophagaceae* (the class of Sphingobacteria) and *Micrococcaceae* (the class of  
536 Actinobacteria). *Moraxellaceae* was found enriched in BES for the treatment of  
537 practical petroleum wastewater (Jain et al. 2016). *Comamonadaceae* and  
538 *Alcaligenaceae* were the predominant genera in the nitrification and denitrification  
539 community of MFC with dual-chamber and batch-type cathode (Sotres et al. 2016).  
540 More effective cooperative cathodes than dual chemical cathodes stimulated the  
541 growth of EAMs (Table 2) and produced more electrons (ref. 3.3), which enhanced  
542 the performances of Cr(VI) reduction and electricity generation in BER<sub>CC</sub> (ref. 3.1).

543

## 544 **Conclusions**

545 BER with cooperative cathodes of chemical cathode and bio-cathode (BER<sub>CC</sub>)  
546 for improvement of Cr(VI) reduction and electricity generation were realized. The  
547 specific conclusions were drawn as follows:

548 (1) The configuration of cooperative cathodes in BER<sub>CC</sub> benefited from  
549 preventing the decline of the performance of Cr(VI) reduction. Cr(VI) could be fully  
550 reduced, and the voltage was consistently above 0.672 V for at least seven cycles in  
551 BER<sub>CC</sub>. The TCOD of sludge decreased by 73.6±4.4%.

552 (2) Both the Cr(VI) reduction rate and the power density were increased with the



553 increase of TCOD and initial Cr(VI) concentration and the decrease of pH.

554 (3) Quantification of the electron fluxes revealed that  $8.21 \pm 4.03$  mg C/(L·h) more  
555 electrons were produced from sludge degradation and  $47.9 \pm 4.3\%$  more electrons  
556 participated in cathodic reactions in BER<sub>CC</sub> than those in BER<sub>DC</sub>.

557 (4) More EAMs were found in the anodic biofilm of BER<sub>CC</sub> than that of BER<sub>DC</sub>,  
558 of which *Geobacter* was the most abundant.

559 BER<sub>CC</sub> is an economical technology not only for energy production but also for  
560 treating Cr(VI) contaminated wastewater and degrading excess sludge simultaneously  
561 in electroplating industrial parks.

562

563

564

### Author's contribution

565 **Qingliang Zhao, Yimin Zhu:** Conceptualization, Supervision, Writing- Reviewing  
566 and Editing. **Hang Yu:** Conceptualization, Methodology, Software, Writing- Original  
567 draft preparation. **Yuhui Cao:** Data curation. **Tiantian Sun:** Software. **Weifeng Liu:**  
568 Writing- Reviewing and Editing. **Zhuyuan Liang:** Methodology, Software.

569

570

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576

577

### **Availability of data and materials**

578

The datasets used and/or analyzed during the current study are available from the

579

corresponding author on reasonable request.

580

581

### **Declarations**

582

#### **Ethics approval and consent to participate**

583

Not applicable.

584

#### **Consent for publication**

585

Not applicable.

586

#### **Competing interests**

587

The authors declare no competing interests.

588

589

590 **References**

- 591 Aelterman P, Rabaey K, Pham HT, Boon N, Verstraete W (2006) Continuous  
592 electricity generation at high voltages and currents using stacked microbial fuel  
593 cells. *Environ Sci Technol* 40: 3388-3394.
- 594 An ZY, Zhang HC, Wen QX, Chen ZQ, Du MA (2014) Desalination combined with  
595 hexavalent chromium reduction in a microbial desalination cell. *Desalination* 354:  
596 181-188.
- 597 APHA (2015) *Standard Methods for the Examination of Water and Wastewater*, 23rd  
598 ed. American Public Health Association, Standard methods for the examination  
599 of water and wastewater, 23rd edn. United Book Press, Gwynn Oak.
- 600 Bond DR, Holmes DE, Tender LM, Lovely DR (2002) Electrode-reducing  
601 microorganisms that harvest energy from marine sediments. *Science* 295: 483.
- 602 Campanaro S, Treu L, Kougias PG, Francisci DD, Valle G, Angelidaki I (2016)  
603 Metagenomic analysis and functional characterization of the biogas microbiome  
604 using high throughput shotgun sequencing and a novel binning strategy.  
605 *Biotechnol Biofuels* 9: 1-17.
- 606 Cerrillo M, Oliveras J, Viñas MM, Bonmatí A (2016) Comparative assessment of raw  
607 and digested pig slurry treatment in bioelectrochemical systems.  
608 *Bioelectrochemistry* 110: 69-78.
- 609 Chaudhuri SK, Lovley DR (2003) Electricity generation by direct oxidation of  
610 glucose in mediatorless microbial fuel cells. *Nat Biotechnol* 21: 1229-1232.
- 611 Chen Z, Zhu BK, Jia WF, Liang JH, Sun GX (2015) Can electrokinetic removal of  
612 metals from contaminated paddy soils be powered by microbial fuel cells?  
613 *Environ Technol Inno* 3: 63-67.
- 614 Gupta S, Yadav A, Verma Nishith (2017) Simultaneous Cr(VI) reduction and

615 bioelectricity generation using microbial fuel cell based on alumina-nickel  
616 nanoparticles-dispersed carbon nanofiber electrode. *Chem Eng J* 307(1):  
617 729-738.

618 Han HX, Chen S, Zhang N, Li Y, Sheng GP (2018). Visible-light-enhanced Cr(VI)  
619 reduction at Pd-decorated silicon nanowire photocathode in photoelectrocatalytic  
620 microbial fuel cell. *Sci Total Environ* 639: 1512-1519.

621 He Z, Angenent LT (2006) Application of bacterial biocathodes in microbial fuel cells.  
622 *Electroanal* 18: 2009-2015.

623 Humphries AC, Nott KP, Hall LD, Macaskie LE (2005) Reduction of Cr(VI) by  
624 immobilized cells of *desulfovibrio vulgaris* NCIMB 8303 and *Microbacterium* sp.  
625 NCIMB 13776. *Biotechnol Bioeng* 90: 589-596.

626 Ishii S, Shimoyama T, Hotta Y, Watanabe K (2008) Characterization of a filamentous  
627 biofilm community established in a cellulose-fed microbial fuel cell. *BMC*  
628 *Microbiol* 8: 6.

629 Jadhav DA, Ray SG, Ghangrekar MM (2017) Third generation in bio-electrochemical  
630 system research - A systematic review on mechanisms for recovery of valuable  
631 by-products from wastewater. *Renew Sust Energ Rev* 76: 1022-1031.

632 Jain P, Srikanth S, Kumar M, Sarma PM, Singh MP, Lal B (2016) Bio-electro  
633 catalytic treatment of petroleum produced water: Influence of cathode potential  
634 upliftment. *Bioresource Technol* 219: 652-658.

635 Jiang JQ, Zhao QL, Zhang JN, Zhang GD, Lee DJ (2009) Electricity generation from  
636 bio-treatment of sewage sludge with microbial fuel cell. *Bioresource Technol*  
637 100: 5808-5812.

638 Jin W, Tolba R, Wen J, Li K, Chen A (2013) Efficient extraction of lignin from black  
639 liquor via a novel membrane-assisted electrochemical approach. *Electrochim*

640 Acta 107: 611-618.

641 Kim CM, Lee CR, Song YE, Heo JH, Choi, SM, Lim DH, Cho J, Park C, Jang M,  
642 Kim JR (2017) Hexavalent chromium as a cathodic electron acceptor in a bipolar  
643 membrane microbial fuel cell with the simultaneous treatment of electroplating  
644 wastewater. Chem Eng J 328: 703-707.

645 Lee J, Phung NT, Chang IS, Kim BH, Sung HC (2003) Use of acetate for enrichment  
646 of electrochemically active microorganisms and their 16S rDNA analyses. FEMS  
647 Microbiol Lett 223: 185-191.

648 Li M, Zhou SQ (2019) Efficacy of Cu(II) as an electron-shuttle mediator for improved  
649 bioelectricity generation and Cr(VI) reduction in microbial fuel cells. Chem Eng  
650 J 273: 122-129.

651 Li M, Zhou SQ, Xu YT, Liu ZJ, Ma FZ, Zhi LL, Zhou X (2018) Simultaneous Cr(VI)  
652 reduction and bioelectricity generation in a dual chamber microbial fuel cell.  
653 Chem Eng J 334: 1621-1629.

654 Li Y, Lu AH, Ding HR, Jin S, Yan HH, Wang CQ, Zen CP, Wang X (2009) Cr(VI)  
655 reduction at rutile-catalyzed cathode in microbial fuel cells. Electrochem  
656 Commun 11: 1496-1499.

657 Logan BE, Hamelers B, Rozendal R, Schröder U, Keller J, Freguia S, Aelterman P,  
658 Verstraete W, Rabaey K (2006) Microbial fuel cells: Methodology and  
659 technology. Environ Sci Technol 40: 5181–5192.

660 Logan BE, Regan JM (2006) Electricity-producing bacterial communities in microbial  
661 fuel cells. Trends Microbiol 14: 512-518.

662 Lu L, Xing D, Ren ZJ (2015) Microbial community structure accompanied with  
663 electricity production in a constructed wetland plant microbial fuel cell.  
664 Bioresource Technol 195: 115-121.

665 Martins M, Faleiro ML, Chaves S, Tenreiro R, Costa MC (2010) Effect of uranium  
666 (VI) on two sulphate-reducing bacteria cultures from a uranium mine site. *Sci*  
667 *Total Environ* 408: 2621-2628.

668 Meng FY, Zhao QL, Zheng ZL, Wei L, Wang K, Jiang JQ, Ding J, Na XL (2019)  
669 Simultaneous sludge degradation, desalination and bioelectricity generation in  
670 two-phase microbial desalination cells. *Chem Eng J* 36: 1180-1188.

671 Pang YM, Xie DH, Wu BG, Lv ZS, Zeng XH, Wei CH, Feng CH (2013) Conductive  
672 polymer-mediated Cr(VI) reduction in a dual-chamber microbial fuel cell under  
673 neutral conditions. *Synth Met* 183(1): 57-62.

674 Park HS, Kim BH, Kim HS, Kim HJ, Kim GT, Kim M, Chang IS, Park YK, Chang HI  
675 (2003) A Novel Electrochemically Active and Fe(III)-reducing Bacterium  
676 Phylogenetically Related to *Clostridium butyricum* Isolated from a Microbial  
677 Fuel Cell. *FEMS Microbiol Lett* 223: 129-134.

678 Pratt S, Yuan ZG, Gapes D, Dorigo M, Zeng RJ, Keller J (2003) Development of a  
679 novel titration and off-gas analysis (TOGA) sensor for study of biological  
680 processes in wastewater treatment systems. *Biotechnol Bioeng* 81: 482-495.

681 Rabaey K, Angenent L, Schröder U, Keller J (2009) *Bioelectrochemical Systems:*  
682 *From Extracellular Electron Transfer to Biotechnological Application*, IWA  
683 Publishing: London, UK.

684 Rabaey K, Keller J (2008) Microbial fuel cell cathodes: from bottleneck to prime  
685 opportunity? *Water Sci Technol* 57: 655-659.

686 Rabaey K, Ossieur W, Verhaege M, Verstraete W (2005) Continuous microbial fuel  
687 cells convert carbohydrates to electricity. *Water Sci Technol* 52: 515-523.

688 Rismaniyazdi H, Christy AD, Dehority BA, Morrison M, Yu Z, Tuovinen OH (2007)  
689 Electricity generation from cellulose by rumen microorganisms in microbial fuel

690 cells. *Biotechnol Bioeng* 97: 1398-1407.

691 Rodrigues RM, Guan X, Iñiguez JA, Estabrook DA, Chapman JO, Huang SY, Sletten  
692 EM, Liu C (2019) Perfluorocarbon nanoemulsion promotes the delivery of  
693 reducing equivalents for electricity-driven microbial CO<sub>2</sub> reduction. *Nat Catal* 2:  
694 407-414.

695 Shi L, Dong H, Reguera G, Beyenal H, Lu A, Liu J, Yu HQ, Fredrickson JK (2016)  
696 Extracellular electron transfer mechanisms between microorganisms and  
697 minerals. *Nat Rev Microb* 14: 651-662.

698 Song TS, Jin Y, Bao J, Kang D, Xie J (2016) Graphene/biofilm composites for  
699 enhancement of hexavalent chromium reduction and electricity production in a  
700 biocathode microbial fuel cell. *J Hazard Mater* 317: 73-80.

701 Sophia AC, Saikant S (2016) Reduction of chromium(VI) with energy recovery using  
702 microbial fuel cell technology. *J Water Process Eng* 11: 39-45.

703 Sotres A, Cerrillo M, Viñas M, Bonmatí A (2016) Nitrogen removal in a  
704 two-chambered microbial fuel cell: establishment of a nitrifying-denitrifying  
705 microbial community on an intermittent aerated cathode. *Chem Eng J* 284:  
706 905-916.

707 Sun WJ, Sierra-Alvarez R, Milner L, Field JA (2010) Anaerobic oxidation of arsenite  
708 linked to chlorate reduction. *Appl Environ Microbiol* 76: 6804-6811.

709 Viridis B, Rabaey K, Yuan Z, Rozendal RA, Keller J (2009) Electron Fluxes in a  
710 Microbial Fuel Cell Performing Carbon and Nitrogen Removal. *Environ Sci*  
711 *Technol* 43: 5144-5149.

712 Wan H, Yi XY, Liu XP, Feng CH, Deng Z, Wei CH (2018) Time-dependent bacterial  
713 community and electrochemical characterizations of cathodic biofilms in the  
714 surfactant-amended sediment-based bioelectrochemical reactor with enhanced

715 2,3,4,5-tetrachlorobiphenyl dechlorination. *Environ Pollut* 236: 343-354.

716 Wang G, Huang L, Zhang Y (2008) Cathodic reduction of hexavalent chromium  
717 [Cr(VI)] coupled with electricity generation in microbial fuel cells. *Biotechnol*  
718 *Lett* 30: 1959-1966.

719 Wang GY, Zhang BG, Li S, Yang M, Yin CC (2017) Simultaneous microbial reduction  
720 of vanadium (V) and chromium (VI) by *Shewanella loihica* PV-4. *Bioresource*  
721 *Technol* 227: 353-358.

722 Wrighton KC, Agbo P, Warnecke F, Weber KA, Brodie EL, DeSantis TZ, Hugenholtz  
723 P, Andersen GL, Coates JD (2008) A novel ecological role of the Firmicutes  
724 identified in thermophilic microbial fuel cells. *ISME J* 2: 1146-1156.

725 Xafenias N, Zhang Y, Banks CJ (2014) Evaluating hexavalent chromium reduction  
726 and electricity production in microbial fuel cells with alkaline cathodes. *Int J*  
727 *Environ Sci Technol* 12: 2435-2446.

728 Xie S, Liang P, Chen Y, Xia X, Huang X (2011) Simultaneous carbon and nitrogen  
729 removal using an oxic/anoxic-biocathode microbial fuel cells coupled system.  
730 *Bioresource Technol* 102(1): 348-354.

731 Yu H, Zhao QL, Dong QS, Jiang JQ, Wang K, Zhang YS (2018) Electronic and  
732 metagenomic insights into the performance of bioelectrochemical reactor  
733 simultaneously treating sewage sludge and Cr(VI)-laden wastewater. *Chem Eng*  
734 *J* 341: 495-504.

735 Yu J, Park Y, Kim B, Lee T (2015) Power densities and microbial communities of  
736 brewery wastewater-fed microbial fuel cells according to the initial substrates.  
737 *Bioproc Biosyst Eng* 38: 85-92.

738 Zhang F, He Z (2012) Simultaneous nitrification and denitrification with electricity  
739 generation in dual-cathode microbial fuel cells. *J Chem Technol Biotechnol*



740 87(1): 153-159.

741 Zhang GD, Zhao QL, Jiao Y, Wang K, Lee DJ, Ren NQ (2012) Efficient electricity

742 generation from sewage sludge using biocathode microbial fuel cell. *Water Res*

743 46: 43-52.

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## Table and Figure captions

745 **Table 1** Summary of the experimental results at the anode and cathodes during two  
746 operation cycles of BER<sub>CC</sub> and BER<sub>DC</sub> ( $\Delta$ TCOD - the TCOD consumption  
747 rate (mg/L-C/h);  $\Delta$ CH<sub>4</sub> - the methane production rate (mg/L-C/h);  $\Delta$ H<sub>2</sub> - the  
748 hydrogen gas production rate (mg/L-C/h);  $\Delta$ X - the biomass growth rate  
749 (mg/L-C/h);  $\Delta$ X<sub>E</sub> - the growth of exoelectrogens (mg/L-C/h);  $\Delta$ X<sub>CH<sub>4</sub></sub> - the  
750 growth of methanogens).

751 **Table 2** Relative abundance of well-known EAMs in the bacterial communities from  
752 anodes of BER<sub>CC</sub> and BER<sub>DC</sub>.

753 **Fig. 1** The configurations of BER<sub>CC</sub> (a) and BER<sub>DC</sub> (b).

754 **Fig. 2** Variations of Cr(VI) concentration (a), voltage (b), and TCOD (c) in BER<sub>CC</sub>  
755 and BER<sub>DC</sub>.

756 **Fig. 3** The reduction of Cr(VI) (a) and the polarization curves and power density  
757 curves (b) of BER<sub>CC</sub> at different TCODs of excess sludge.

758 **Fig. 4** The reduction of Cr(VI) (a) and the polarization curves and power density  
759 curves (b) of BER<sub>CC</sub> at different Cr(VI) concentrations.

760 **Fig. 5** The reduction of Cr(VI) (a) and the polarization curves and power density  
761 curves (b) of BER<sub>CC</sub> at different pHs of catholyte.

762 **Fig. 6** Distribution of electron fluxes among different electron sinks and the electron  
763 fluxes per unit time at the anode and cathode of BER<sub>DC</sub> and BER<sub>CC</sub>  
764 (Anode-CH<sub>4</sub> - estimated electrons used for methane production at anode;  
765 Anode-H<sub>2</sub> - estimated electrons used for hydrogen production at anode;  
766 Anode-I - electrons transferred from the anode to the cathode to generate  
767 current at anode; Anode-X<sub>CH<sub>4</sub></sub> - estimated electrons consumed for methanogen  
768 growth at anode; Anode-X<sub>E</sub> - estimated electrons consumed for exoelectrogen

769 growth at anode; Bio-cathode and loss - estimated electrons transferred to  
770 bio-cathode or loss in transfer; Chemical cathode - estimated electrons  
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773 of BER<sub>CC</sub> and the anode of BER<sub>DC</sub> at the phyla (a), class (b) and genus level  
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 780 growth of exoelectrogens (mg/L-C/h);  $\Delta$ X<sub>CH<sub>4</sub></sub> - the growth of methanogens).

	BER <sub>DC</sub>	BER <sub>CC</sub>
Cycle (h)	55.5~102.5	57.5~58.5
Current (mA)	0.48	0.66
$\Delta$ TCOD (mg C/L·h)	18.42±4.03	26.63±3.94
$\Delta$ CH <sub>4</sub> (mg C/L·h)	7.23±1.94	6.65±1.23
$\Delta$ Cr(VI) (mg C/L·h)	1.07±0.11	1.67±0.17
$\Delta$ X (mg C/L·h)	9.59±0.43	8.59±2.23
$\Delta$ X <sub>CH<sub>4</sub></sub> (mg C/L·h)	5.18±1.23	3.18±1.23
$\Delta$ X <sub>E</sub> (mg C/L·h)	4.41±0.67	5.31±0.72

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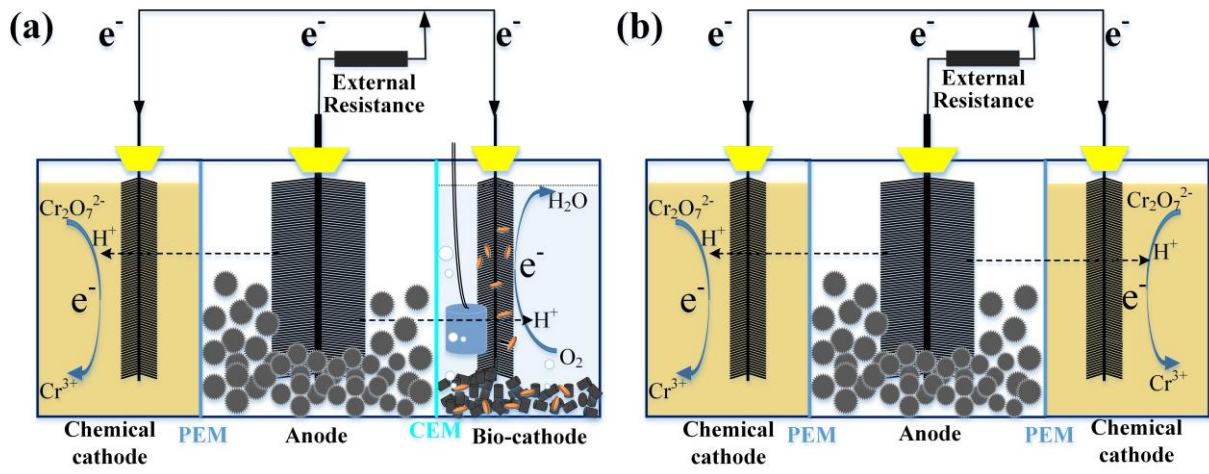
792 **Table 2** Relative abundance of well-known EAMs in the bacterial communities from  
 793 anodes of BER<sub>CC</sub> and BER<sub>DC</sub>.

EAM	Relative abundance (%)		
	Anode of BER <sub>CC</sub>	Anode of BER <sub>DC</sub>	Bio-cathode of BER <sub>CC</sub>
<i>Geobacter spp.</i>	3.7	2.2	-
<i>Arcobacter butzlerii</i>	0.04	0.01	1.22
<i>Desulfovibrio denitrificans</i>	0.01	-	-
<i>Comamonas denitrificans</i>	0.13	0.02	-
<i>Desulfuromonas acetoxidans</i>	0.02	-	-
<i>Klebsiella pneumoniae</i>	0.01	-	-
<i>Rhodoferax ferrireducens</i>	0.03	-	-

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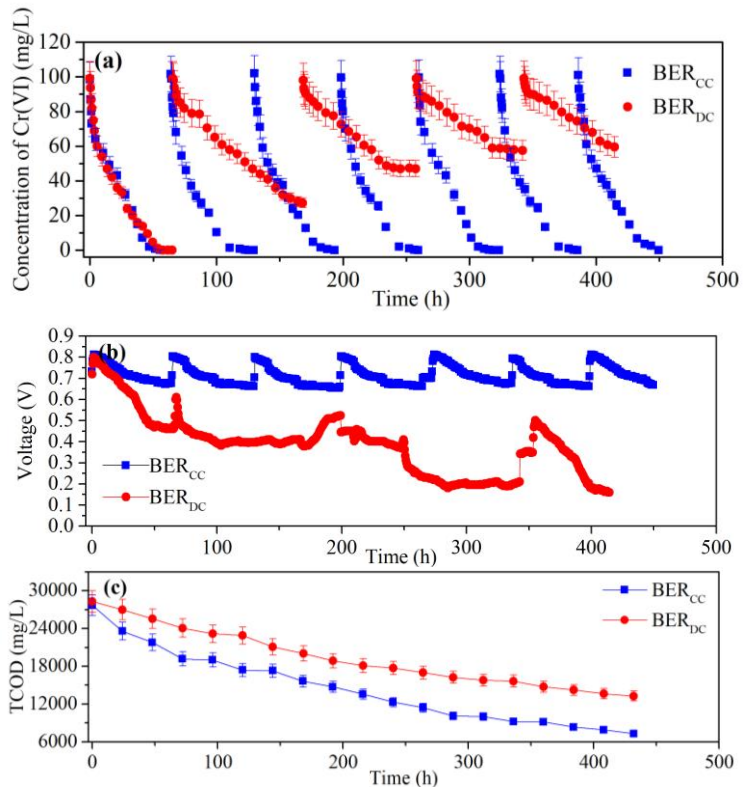
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**Fig. 1** The configurations of BER<sub>CC</sub> (a) and BER<sub>DC</sub> (b).

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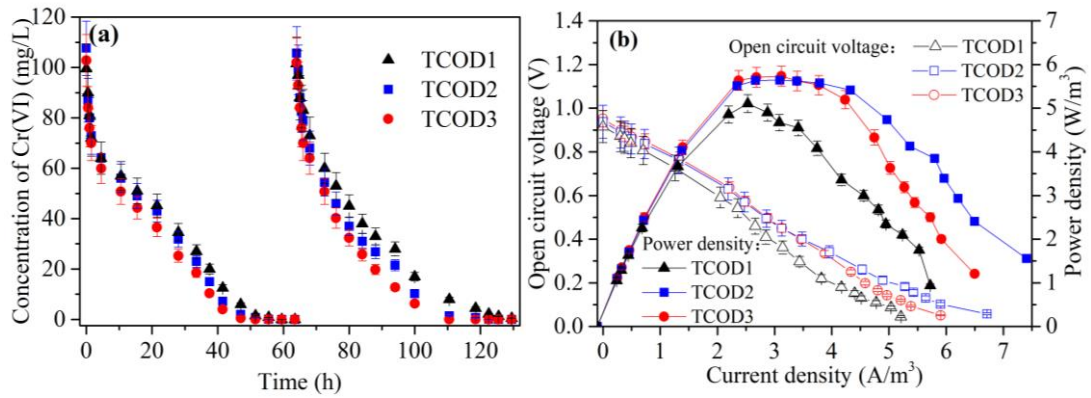
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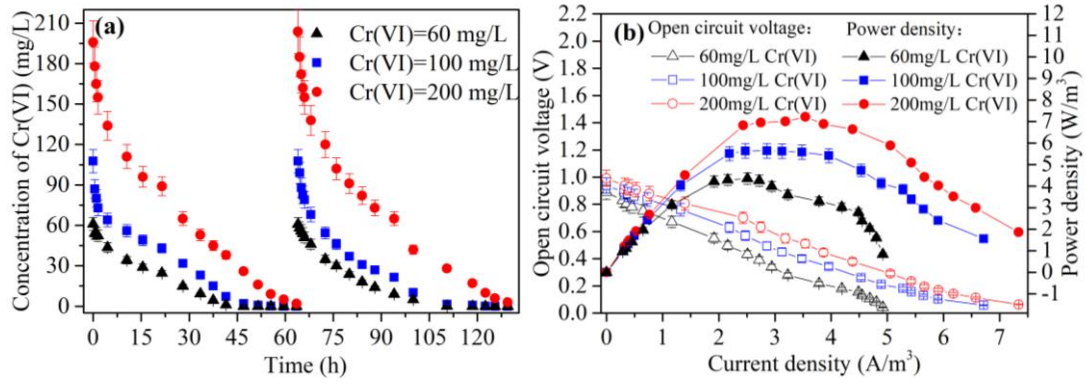
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curves (b) of BER<sub>CC</sub> at different TCODs of excess sludge.

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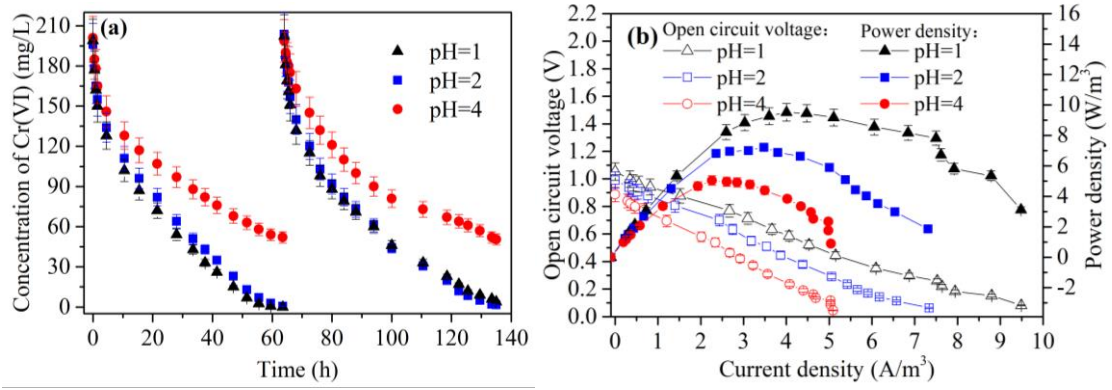
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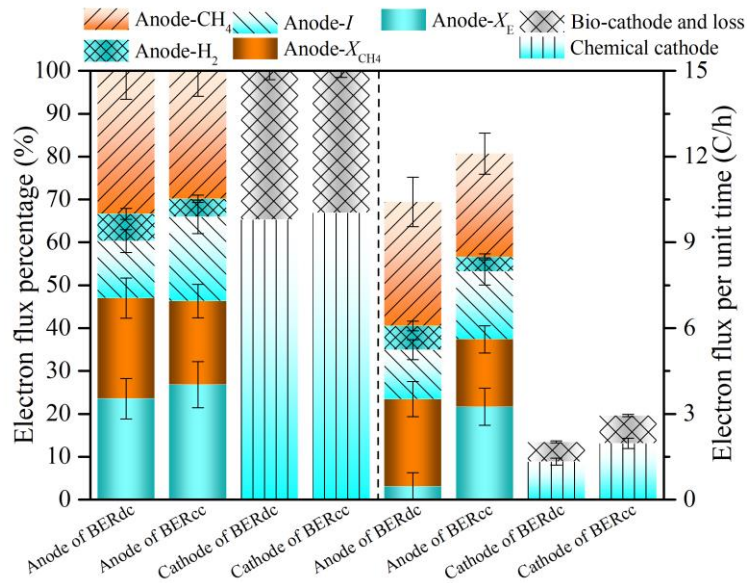
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826 electrons used for hydrogen production at anode; Anode-I - electrons transferred from

827 the anode to the cathode to generate current at anode; Anode-X<sub>CH<sub>4</sub></sub> - estimated

828 electrons consumed for methanogen growth at anode; Anode-X<sub>E</sub> - estimated electrons

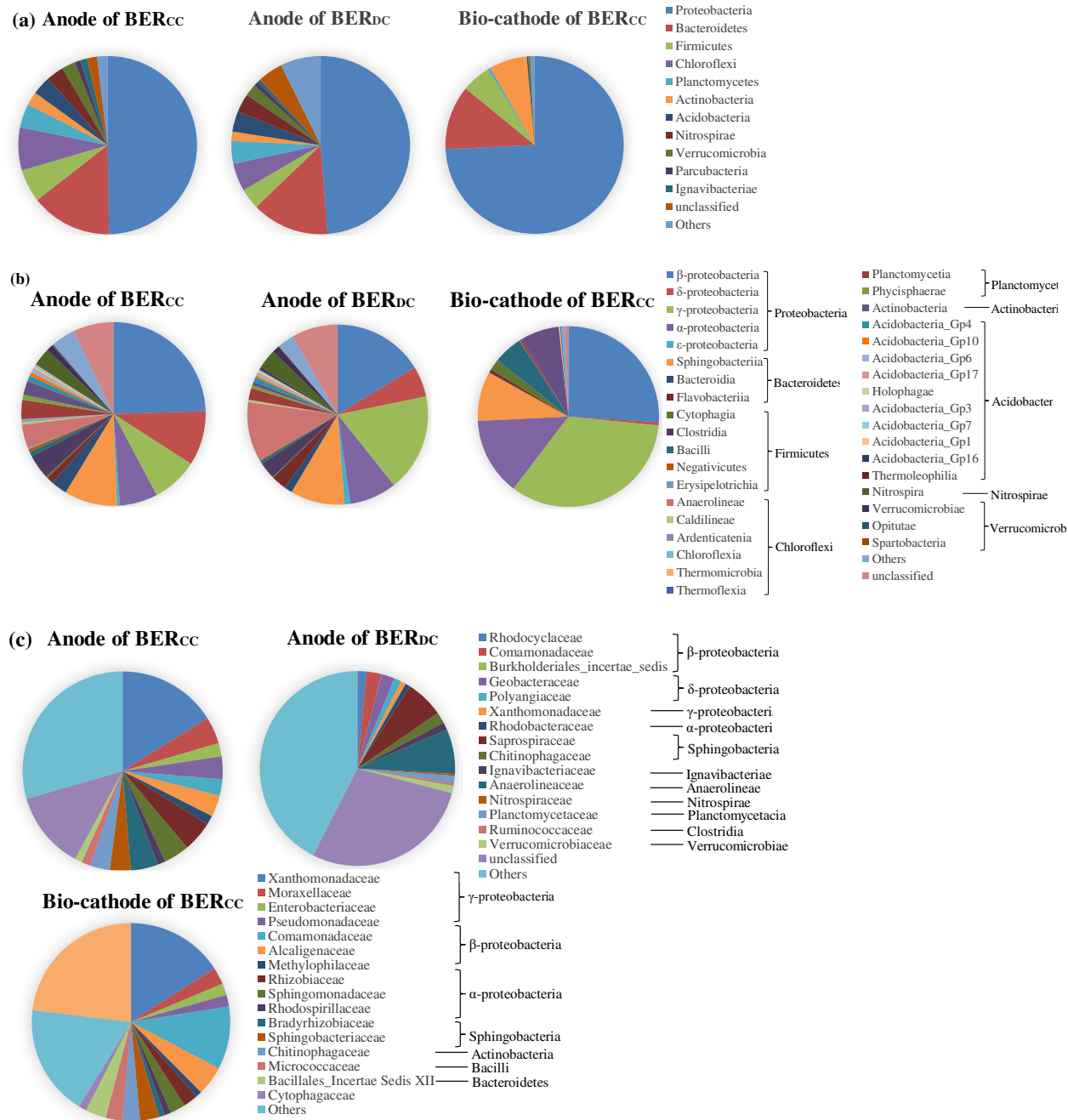
829 consumed for exoelectrogen growth at anode; Bio-cathode and loss - estimated

830 electrons transferred to bio-cathode or loss in transfer; Chemical cathode - estimated

831 electrons transferred to chemical cathode).

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