

TonB-dependent receptor epitopes expressed in *M. bovis* BCG induced significant protection in the hamster model of leptospirosis

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Abstract

Leptospirosis is an emerging infectious disease caused by pathogenic *Leptospira* spp. A universal vaccine against leptospirosis is likely to require highly conserved epitopes from pathogenic leptospires, that are exposed on the bacterial surface, and that generate a protective and sterilizing immune response. Our group recently identified several genes predicted to encode TonB-dependent receptors (TBDR) in *Leptospira interrogans* using a reverse vaccinology approach. Three leptospiral TBDRs were previously described and partially characterized as ferric-citrate, hemin, and cobalamin transporters. In the current study, we designed a fusion protein composed of predicted surface-exposed epitopes from three conserved leptospiral TBDRs. Based on their three-dimensional structural models and the prediction of immunogenic regions, nine putative surface-exposed fragments were selected to compose a recombinant chimeric protein. A *Mycobacterium bovis* BCG strain expressing this chimeric antigen encoded in the pUP500/P_{pAN} mycobacterial expression vector was used to immunize Syrian hamsters. All animals (20/20) vaccinated with recombinant BCG survived infection with an endpoint dose of *L. interrogans* ($P < 0.001$). No animal survived in the negative control group. Immunization with our recombinant BCG elicited a humoral immune response against leptospiral TBDRs, as demonstrated by ELISA and immunoblot. No leptospiral DNA was detected by *lipL32* qPCR in the kidneys of vaccinated hamsters. Similarly, no growth was observed in macerated kidney cultures from the same animals, suggesting the induction of a sterilizing immune response. Design of new vaccine antigens based on the structure of outer membrane proteins is a promising approach to overcome the impact of leptospirosis by vaccination.

Key Points

- Predicted surface-exposed epitopes were identified in three leptospiral TBDRs;
- A *M. bovis* BCG strain expressing a chimeric protein (rTBDRchi) was constructed;
- Hamsters vaccinated with rBCG:TBDRchi were protected from lethal leptospirosis.

Introduction

Leptospirosis is a major zoonosis worldwide in terms of morbidity and mortality rates. The global incidence is estimated to be over one million cases every year, resulting in ~60,000 deaths (Costa et al. 2015). Rodents are the primary reservoir hosts of pathogenic *Leptospira* spp., while humans are incidental hosts. The disease also affects several wild and domestic animals (Ellis 2015; Haake and Levett 2015). Leptospirosis in humans can vary from mild symptoms, such as fever and myalgia to severe outcomes, with the development of acute kidney injury and pulmonary hemorrhage syndrome (Haake and Levett 2015). Leptospiral vaccines currently available are inactivated whole-cell preparations (bacterins) with widespread veterinary applications, but limited use in humans. Bacterins confer a short-term immune response that is protective only against the serovars included in the vaccine formulation (Adler 2015). Several recombinant proteins have been evaluated towards the development of new and improved leptospiral vaccines, with variable success (Felix et al. 2020). While some experimental recombinant vaccines were protective against lethal challenge, the majority failed to achieve heterologous and/or sterilizing protection, highlighting the need to evaluate novel vaccination strategies (Grassmann et al. 2017c; Felix et al. 2020). A new and effective vaccine against leptospirosis will likely include surface-exposed epitopes conserved among pathogenic *Leptospira* spp., allowing bacterial recognition by humoral immune response (Grassmann et al. 2017c; Grassmann et al. 2017a; Dellagostin et al. 2017). Our group previously used a reverse and structural vaccinology approach to identify leptospiral beta-barrel outer membrane proteins (β -OMP), including six TonB-dependent receptors (TBDR) (Grassmann et al. 2017a).

TBDRs are outer-membrane proteins that promote the uptake of essential nutrients such as iron, nickel, vitamins and carbohydrates (Noinaj et al. 2010). Since iron has limited bioavailability, its acquisition by bacteria is facilitated by the formation of complexes with other transport molecules, such as heme and siderophores, which are then internalized by specific TonB-dependent transport systems (Noinaj et al. 2010). Pathogenic and saprophytic *Leptospira* spp. require iron for *in vitro* growth (Faine 1959) and evolved mechanisms for iron sensing, acquisition and utilization (Louvel et al. 2006). Two leptospiral TBDRs, encoded by LIC10896 and LIC10964, were described as homologs to the ferric citrate transporter FecA and the PhuR hemin receptor (Nascimento et al. 2004; Louvel et al. 2006), respectively. Another TBDR, LIC12374, is annotated as a homolog of the cobalamin transporter BtuB (Nascimento et al. 2004; Louvel et al. 2006). Cobalamin (or vitamin B12) is a cofactor for enzymes related to a variety of essential functions, including those related to carbon metabolism and biosynthesis of nucleotides (Shelton et al. 2019). Similarly to iron, cobalamin is a small nutrient required for *Leptospira* spp. survival (Louvel et al. 2006). While saprophytic species rely on external sources, pathogenic leptospires encode a pathway for *de novo* production of cobalamin from L-glutamate (Fouts et al. 2016). Due to their important role in bacterial metabolism and nutrient acquisition, several TBDRs have been evaluated as vaccine candidates against many pathogens, resulting in successful protection (Alteri et al. 2009; Hu et al. 2012; Hubert et al. 2013).

Epitope mapping in vaccine candidates permits the identification of regions with increased probability to elicit an immune response, consequently improving the antigenicity and immunogenicity of vaccine formulations and avoiding production of undesirable or ineffective responses (Oyarzún and Kobe 2016; Parvizpour et al. 2020). The rational selection of conserved epitopes also contributes to cross-protection (Liu et al. 2017; Nosrati et al. 2019; Xiao et al. 2020), a particularly important aspect to consider for the development of leptospirosis vaccines, where the achievement of heterologous, long-term protection are major goals. Combined with protein structural information, epitope selection is a powerful tool to identify immunogenic surface-exposed fragments required for bacterial opsonization (Grassmann et al. 2017a). Opsonization of leptospires by specific immunoglobulins improved bacterial phagocytosis and clearance (Gomes-Solecki et al. 2017; Santecchia et al. 2020). Even though the immune response required for protection against leptospirosis is not yet fully understood, induction of a balanced cellular/humoral response is believed to be key for a protective immune response (Gomes-Solecki et al. 2017; Grassmann et al. 2017c; da Cunha et al. 2019).

Mycobacterium bovis Bacillus Calmette-Guérin (BCG) is an attenuated strain widely used as a vaccine against human tuberculosis (TB) (Dockrell and Smith 2017). In use for a hundred years, BCG vaccination is a highly cost-effective intervention against tuberculosis, and is recommended for all newborns in countries with a high burden of TB (Trunz et al. 2006; Lobo et al. 2021). BCG triggers a long-lasting immunity after a single dose, presenting several adjuvant

properties and a strong cellular response (Dockrell and Smith 2017). BCG-induced adaptive immune response include the expression of several interleukins, such as TNF- α and IFN- γ , and consequently, the proliferation and activation of macrophages (Dockrell and Smith 2017; Covián et al. 2019). Several studies have demonstrated the potential of recombinant BCG (rBCG) to enhance the immune response towards heterologous antigens (Gröschel et al. 2017; Soto et al. 2018), including those from *L. interrogans* (Seixas et al. 2007; Oliveira et al. 2019a; Dorneles et al. 2020), and represents a promising alternative for antigen delivery (Oliveira et al. 2017).

In this study, we identified highly conserved surface-exposed regions containing B and T CD4+ cell epitopes in three leptospiral TBDRs: LIC10896, LIC10964, and LIC12374. These fragments were combined on a multi-epitope fusion protein and evaluated using BCG as a vectorized vaccine carrier. Our preparation induced a protective immune response in the hamster model of acute leptospirosis, generating specific antibodies and sterilizing immunity.

Materials And Methods

Bacterial strains and growth conditions

Escherichia coli strains BL21 (DE3) Star and TOP10 (Invitrogen, São Paulo, SP, Brazil) were grown at 37°C in Lysogeny Broth (LB) medium, with 100 $\mu\text{g}\cdot\text{ml}^{-1}$ ampicillin when required. *M. bovis* BCG (Pasteur and recombinant strains) were cultivated at 37°C in Middlebrook 7H9 broth (Difco, BD, São Paulo, SP, Brazil) supplemented with 0.5% glycerol, 0.05% Tween 80 and 10% Oleic acid, Albumin, Dextrose Complex (OADC) or in 7H10 agar (Difco, BD, São Paulo, SP, Brazil) with 10% OADC. The BCG media was supplemented with kanamycin, 25 $\mu\text{g}\cdot\text{ml}^{-1}$, when needed. Low-passage virulent *L. interrogans* serogroup Icterohaemorrhagiae serovar (sv.) Copenhageni strain (st.) Fiocruz L1-130 (WDCM1012) was cultivated at 30°C in Ellinghausen–McCullough–Johnson–Harris (EMJH) medium supplemented with *Leptospira* EMJH enrichment (Difco, BD, São Paulo, SP, Brazil).

In silico structure predictions and epitope selection

Three proteins (LIC10896, LIC10964 and LIC12374) from *L. interrogans* sv. Copenhageni st. Fiocruz L1-130 (Taxid:267671), previously identified as leptospiral TonB-Dependent receptors, were selected for evaluation in this study. Three-dimensional (3D) structures were predicted by threading using the I-TASSER server (Yang and Zhang 2015). Models with the highest C-Score were selected for epitope prediction. The 3D protein structures were visualized using USCF Chimera 1.12 (Pettersen et al. 2004). Sequences for the CD4+ T cell epitopes, with high affinity to 14 different human major histocompatibility complex class II (MHC-II) alleles, were predicted using NetMHCII 2.2 (Nielsen and Lund 2009) at the default settings. For linear (continuous) B cell epitope prediction, amino acid sequences were analyzed using BepiPred 1.0 (Larsen et al. 2006). Only T cell epitopes with a strong binding affinity ($\text{IC}_{50} < 50 \text{ nM}$) and amino acid residues with a score higher than 0.35 in the B cell epitope analysis were considered. Epitopes were manually mapped onto the predicted 3D models of each β -OMP in USCF Chimera 1.12. These models were aligned with their best structural template obtained from the Orientations of Proteins in Membranes (OPM) database (Lomize et al. 2012). All surface-exposed fragments containing B and T CD4+ cell epitopes were used for antigen construction. Orthologs for each *L. interrogans* TBDR were identified by BlastP (NCBI) in 10 pathogenic (P1 clade) *Leptospira* spp. proteomes (Online Resource 1, Table S1). Protein sequences with >70% identity and 40% query coverage were considered orthologs. Identity between the *L. interrogans* polypeptides and their orthologs was determined after multiple sequence alignments (MSA) using MUSCLE (EMBL-EBI) (Edgar et al. 2004).

Recombinant protein expression and purification

Selected surface-exposed fragments were assembled *in silico* using Vector NTI Advance® 11.5 (Invitrogen) without linkers, to construct the fusion proteins rTBDRchi, rLIC10896chi, rLIC10964chi and rLIC12374chi (Online Resource 2). *E. coli* codon optimized coding sequences (CDS) for each fusion protein were synthesized by Epoch Life Science (Texas, USA) and cloned into *E. coli* expression vector pAE (Ramos et al. 2004) using *Bam*HI and *Hind*III restriction sites. Recombinant plasmids were used to transform *E. coli* BL21 (DE3) C41 strain (Invitrogen, São Paulo, SP, Brazil) by heat-shock. When cultures reached mid-log phase (0.6-0.8 absorbance at OD_{600}), expression was induced by adding 0.5 mM IPTG (isopropyl- β -D-thiogalactopyranoside) and incubated at 37°C with agitation. After 4h, cells were harvested by centrifugation (7,000 $\times g$, 15 min, 4°C). Pellets were washed in phosphate-buffered saline (PBS) and resuspended in lysis buffer (20 mM sodium phosphate, 0.5 M NaCl and 20 mM Imidazole, pH 8.0) before sonication with six 30-s pulses on ice. After lysis, insoluble proteins were harvested by centrifugation (10,000 $\times g$, 1 h, 4°C), and resuspended in denaturing binding buffer (8M urea, 20 mM sodium phosphate, 0.5 M NaCl, and 20 mM Imidazole, pH 8.0) and incubated under constant agitation for 16 h. After a final centrifugation (10,000 $\times g$, 1 h, 4°C), recombinant proteins were purified using the AKTA Start automated chromatography system (GE Healthcare). Briefly, the supernatant was applied to a nickel-charged HisTrap FF column (GE Healthcare, São Paulo, SP, Brazil), and after washing with 20 column volumes, recombinant fusion proteins were eluted over a total of 20 ml using a 0-100% gradient of elution buffer (8M urea, 20 mM sodium phosphate, 0.5 M NaCl, and 0.5 M Imidazole, pH 8.0). Purified proteins were then dialyzed against PBS at 4°C for 24 h and stored at -80°C or 4°C.

Production of anti-rTBDRchi hyperimmune sera

Four-week-old female Wistar rats (*Rattus norvegicus*) were injected intraperitoneally with 50 μg rTBDRchi emulsified in complete Freund adjuvant for the first dose and incomplete Freund adjuvant for the remaining two doses. Two weeks after the second boost, animals were euthanized by exsanguination, and hyperimmune sera were obtained by centrifugation (3,500 $\times g$, 15 min, 4°C) and stored at -20°C.

Construction of recombinant *M. bovis* BCG and expression analysis

Using pAE/rTBDRchi as template, the rTBDRchi CDS was amplified by PCR using primers described in Online Resource 1 (Table S2). The PCR product was digested using *Xba*I and *Pst*I restriction enzymes (New England BioLabs) and cloned into the BCG expression vector pUP500/PpAN (Oliveira et al. 2019b), previously digested with *Spe*I and *Pst*I (New England BioLabs), generating pUP500/PpAN:rTBDRchi plasmid. Recombinant pUP500/PpAN:rTBDRchi was transformed into *M. bovis* BCG Pasteur cells by electroporation, as previously described (Parish and Stoker 1995). Cells were plated onto 7H10 agar (Difco, BD,

São Paulo, SP, Brazil) containing 25 µg.ml⁻¹ of kanamycin and incubated at 37°C for 21 days. Resistant colonies were inoculated in selective Middlebrook 7H9 broth (Difco) and cultivated at 37°C to evaluate recombinant expression by immunoblot. Briefly, whole-cell lysates were prepared from rBCG:TBDRchi grown to mid-logarithmic phase at 37°C (OD₆₀₀ = 0.8), when cells were collected by centrifugation (4,000 × g, 15 min) and resuspended in Tris-HCl (pH 8), adjusting the cell density to 10⁷ CFU.ml⁻¹. rBCG:TBDRchi whole-cell lysates were separated in 12% polyacrylamide gels and transferred to nitrocellulose membranes (Hybond ECL, GE Healthcare, Illinois, United States). Membranes were incubated overnight with rat anti-rTBDRchi sera at 1:100 dilution, followed by anti-rat HRP-conjugated secondary antibodies (Sigma Aldrich, Missouri, United States) diluted 1:5,000. Immunoblots were developed using Pierce ECL Western Blotting Substrate (Thermo-Fisher, Illinois, USA).

Vaccination and challenge of hamsters

rBCG:TBDRchi and BCG Pasteur (negative control) strains were cultivated *in vitro* and collected by centrifugation (4,000 × g, 15 min). Cells were resuspended in PBS, adjusting concentration to 10⁷ cells/ml (considering 1 OD₆₀₀ = 1 × 10⁸ cells/ml), and immediately used. Male Syrian hamsters (*Mesocricetus auratus*), 4–6 weeks old, were subcutaneously inoculated with 10⁶ cells of BCG Pasteur (n=13 per experiment) or recombinant BCG:TBDRchi (n=10 per experiment). An additional control group (n=4 per experiment) was immunized (intramuscular) with a leptospiral whole-cell bacterin (10⁸ heat-inactivated leptospires in 100 µl PBS). Two doses of each formulation were administered 21 days apart. Thirty days after the second boost, hamsters were infected intraperitoneally with 10³ leptospires (*L. interrogans* sv. Copenhageni st. Fiocruz L1-130), equivalent to five times the average 50% endpoint dose (ED50), as previously determined (Oliveira et al. 2019a). Hamsters were monitored daily for clinical signs of acute leptospirosis for 30 days after challenge. Criteria for humane euthanasia (CO₂ narcosis) was established as previously described (Coutinho et al. 2011), and includes: >10% weight loss, prostration, bristling, apathy, and lack of appetite. Blood samples were collected one day before the first immunization (day zero) and before challenge (day 51). A total of two independent experiments were performed. All animal procedures were conducted according to the rules and regulations of the Animal Experimentation Ethics Committee at UFPel.

Analysis of the leptospiral burden in hamster kidneys post-challenge

At the end of the experiment, kidneys were collected aseptically in order to evaluate the leptospiral burden post-challenge. Kidney samples were macerated in 10 ml EMJH medium supplemented with 10% *Leptospira* enrichment supplement. After one hour at 30°C, 500 µl of the macerate was inoculated into 10 ml of fresh EMJH and incubated at 30°C for up to 8 weeks. Cultures were monitored weekly by dark-field microscopy. DNA extraction from kidney samples, followed by qPCR was performed as previously described (Dorneles et al. 2020). Briefly, DNA was extracted from approximately 40 mg of kidney tissue using SV Genomic DNA Purification Kit (Promega, Brazil) and quantified using Qubit 2.0 fluorometer (Thermo Fisher). Quantitative real-time PCR (qPCR) detection of leptospiral DNA was performed in a LightCycler 96 system (Roche, Basel, Switzerland), using 200 ng of total DNA, 0.4 µM of each primer specific to *lipL32* (Online Resource 1, Table S2) and SYBR Green PCR Master Mix (Applied Biosystems, São Paulo, SP, Brazil). Reactions were performed in triplicate as previously described (Dorneles et al. 2020). A standard curve was generated from 10⁶ to 10 copies of *lipL32* and samples were considered negative when their qPCR quantitation cycle (Cq) were higher than that obtained for the lowest concentration of 10 copies. Absolute quantification analysis using LightCycler 96 software (Roche) was performed to determine the number of leptospire genome equivalents per reaction, and thereafter converted to copies per µg of total DNA.

Evaluation of the humoral immune response

Humoral immune response elicited after vaccination was evaluated by indirect ELISA (Enzyme-Linked Immunosorbent Assay), as previously described (da Cunha et al. 2019). First, 96 well microtiter plates (Polysorp, Nunc, São Paulo, Brazil) were coated with 1 µg of purified rTBDRchi per well at 4°C overnight, in 0.1 M carbonate-bicarbonate buffer, pH 9.6. Plates were washed three times with PBS-0.05% Tween 20 (PBST) and then blocked with 200 µl of 5% non-fat dry milk in PBS-T, for 1 h at 37°C. Hamsters' sera were diluted 1:50 in PBST and evaluated in triplicate. After incubation for 1h at 37°C, plates were washed and HRP-conjugated anti-Syrian hamster IgG (Sigma Aldrich, Missouri, United States) was added at 1:5,000 dilution for 1 h at 37°C. After a final wash, reactions were developed using o-phenylenediamine dihydrochloride (Sigma-Aldrich, Missouri, United States) and hydrogen peroxide, and stopped after 15 min with 4N H₂SO₄. Absorbance was read at 492 nm wavelength in a microplate reader (Mindray MR-96A, São Paulo, Brazil).

Immunoblots were performed as previously described, with small changes (Groshong et al. 2021). Briefly, samples containing 1 µg of each recombinant protein (rTBDRchi, rLIC10896, rLIC10964, and rLIC12374) or *L. interrogans* st. Fiocruz L1-130 whole-cell lysate (10⁸/lane), were analyzed by SDS-PAGE and transferred to nitrocellulose membranes (GE Healthcare, Illinois, United States) at 20 V for 25 min using a semi-dry transfer (Bio-Rad). Membranes were blocked for 1 h at room temperature with 5% non-fat dried milk diluted in PBS-T, and probed overnight at 4°C with a pool of hamster sera obtained after immunization, at a 1:200 dilution in blocking buffer. After washing 5 times with PBS-T, membranes were incubated for 1 h at RT with an HRP-conjugated goat anti-hamster IgG antibody (Sigma Aldrich, Missouri, United States) diluted in blocking buffer (1:6,000). Following six washes with PBS-T, immunoblots were developed using the Pierce ECL Western Blotting Substrate (Thermo-Fisher, Illinois, USA) detection reagent, and results were visualized using C-Digit Blot Scanner (LI-COR, Nebraska, USA).

Statistical analysis

Significant protection against lethal leptospirosis was evaluated using Fisher's exact test (two-tailed). Comparison of humoral immune response elicited by different experimental groups was performed using analysis of variance (one way-ANOVA with Tukey multiple comparison). For all analyses, *P*-values < 0.05 were considered significant. Graphical representations and statistical analysis were performed using GraphPad Prism 8.

Results

Structural modeling, sequence analysis and design of rTBDRchi

Our group previously described leptospiral TonB-dependent receptor (TBDR) orthologs based on a reverse and structural vaccinology approach (Grassmann et al. 2017a). In this study, we selected the three TBDRs for evaluation as vaccine candidates against leptospirosis, LIC10896, LIC10964, and LIC12374, for which important nutrient transport functions were previously suggested (Nascimento et al. 2004; Louvel et al. 2006). 3D models were generated using the standard threading assembly method by I-TASSER. All retrieved models with the best confidence score (C-score) were identified as structural analogs to TonB-dependent receptors with resolved structures, deposited on the Protein Data Bank (PDB) (Online Resource 1, Table S3). The I-TASSER server was also used to perform structural alignments between the leptospiral queries and their PDB templates using the TM-align program, with reported values varying from 0 to 1, where 1 indicates a perfect superposition of two structures (Zhang and Skolnick 2005). Predicted structures for the leptospiral TBDRs LIC10896, LIC10964, and LIC12374 presented TM-Align values varying from 0.724 to 0.842 (Online Resource 1, Figure S1).

Next, we analyzed each of the selected leptospiral proteins in order to predict T CD4+ and linear B cell epitopes along their sequences. Several T CD4+ and linear B-cell epitopes were identified along the full-length leptospiral TBDR sequences (Online Resource 3). Membrane-embed templates for each generated model were obtained from the Orientations of Proteins in Membranes (OPM) database (Lomize et al. 2012), and structural alignments were performed to identify the predicted surface-exposed fragments (Online Resource 1, Figure S1). Each identified T CD4+ and linear B cell epitope was mapped onto 3D structures allowing the selection of all predicted surface-exposed regions containing antigenic determinants. The three TBDRs analyzed contained nine surface-exposed loops with T and B cell epitopes (Figure 1a). Antigenic loops were selected and combined to construct a 41 kDa recombinant protein, named rTBDRchi (Figure 1b). Five fragments were selected from LIC10896, and two fragments from LIC10964 and LIC12374 (Figure 1, Table 1). *In silico* approaches for the T CD4+ cell epitope prediction focused on the identification of core binding regions composed of nine residues (9-mer), which largely determined binding affinity and specificity for human leukocyte antigens (HLA) presentation (Sanchez-trincado et al. 2017). A total of 24 strong binder 9-mers (IC50 <50 nM) were identified in the exposed loops. Thirteen different HLA alleles (out of 14 alleles analyzed) were predicted to bind to the rTBDRchi epitopes (Online Resource 1, Table S4). rTBDRchi also included promiscuous T cell epitopes, with predicted binding to several HLAs, from all selected leptospiral TBDRs.

Table 1

Sequences of the selected fragments predicted to contain surface-exposed regions, and B and T cell epitopes. Protein sequences were submitted to BepiPrep NetMHCII for prediction of linear B cell (bold) and T CD4+ cell (underlined) epitopes, respectively. All surface-exposed loops containing identified epitopes v selected to compose the fusion protein and are listed.

Protein	Fragment	Sequence
LIC10896	3	YNRSYNYREEANARFQASNPISIYLKDSNMLRPLNQNNLKIYGEEVLWGNLNLAYEPKIGQQFFIKTLYSVQSDKIVREG DGANY IDNFNFKSTN
	4	WSQGGTDL ANGYRRLGNNPDGTR
	5	REIAQRN FTGSDR DIYPIGVEVIYNPLAYANGNRKIYER
	6	SWNGFN TSYGCKT NSEER LLLVANICDAT
	9	DSKIYGLISTGQVDPLSTYAA YSPTTLNRPLQGQSD
LIC10964	3	DLLDNP ENPGATVSQKLLHEKSHFHSFFIFSAGNLELDFSYQR
	8	IQNFIYAASIAQIDLD SGLPKYEYKQGN
LIC12374	2	DQNFSYKNDHGT VVLNTLDD TIDRRKNAS
	6	FPSEEPWYRRQDPLSGDIK

rTBDRchi fragments are conserved among pathogenic *Leptospira* spp.

The selected fragments were evaluated for amino acid conservation between *L. interrogans* and the orthologs from ten additional pathogenic (P1 clade) *Leptospira* spp. (Vincent et al. 2019). By performing a MSA, all sequences showed high conservation levels with identities varying from 73.7 to 100% for different rTBDRchi fragments (Online Resource 4). We were not able to identify a LIC10896 ortholog in *L. kmetzi*, although fragments from LIC10964 and LIC12374 from *L. interrogans*, were highly conserved (84.21 – 93.10%) with their orthologs in *L. kmetzi*. As expected, due to their close phylogenetic relationship, *L. kirschneri* and *L. noguchii* showed the highest conservation with the *L. interrogans* TBDRs, with 7/9 fragments presenting more than 95% identity.

BCG-vectored rTBDRchi protects hamsters against *L. interrogans* challenge

The rTBDRchi CDS was synthesized and cloned into pUP500/P_{DAN} vectors (Oliveira et al. 2019b), allowing its heterologous expression in *M. bovis* BCG, as demonstrated by immunoblot (Online Resource 1, Figure S2). Subcutaneous immunization of hamsters with 10⁶ live rBCG:TBDRchi mycobacteria, elicited 100% protection against homologous *L. interrogans* challenge in two independent experiments (20/20 animals, *P* < 0.001) with animals surviving for 30 days post-challenge (Table 2, Figure 2). In contrast, all animals inoculated with wild-type BCG Pasteur (26/26) developed endpoint criteria, with clinical signs and weight-loss observed for all animals 13-15 days post-challenge (Figure 2). *Leptospira* burden in kidneys was evaluated by culture and *lipL32* qPCR using tissues from animals immunized with rBCG-TBDRchi or from the control groups (Table 2). Cultures from macerated kidneys from the negative control group (BCG Pasteur) were positive for leptospires following two weeks of incubation. No growth was detected after eight weeks in the cultures from the rBCG:TBDRchi and bacterin groups. Similarly, *lipL32* qPCR analysis was negative for leptospiral DNA in the kidneys of animals vaccinated with recombinant BCG (Table 2). The leptospiral burden in the negative control group ranged from 1.08x10¹ – 6.97x10⁴ genome equivalents per µg of total extracted DNA, with an average of 5.03 x 10³ (Table 2, Online Resource 1 – Table S5).

Table 2

Protection against lethal leptospirosis elicited by immunization with rBCG:TBDRchi.

Syrian hamsters were immunized with 10^6 cells of BCG Pasteur wild-type or recombinant BCG:TBDRchi. Thirty days after the second dose, animals were injected intraperitoneally with 5x median endpoint dose (ED50) of *L. interrogans* serovar Copenhageni st. Fiocruz L1-130. Data from two independent experiments are presented.

Vaccine	% Protection ^a	P value ^b	Leptospire in kidneys ^c	
			% Positive Culture	% Positive qPCR
rBCG:TBDRchi	100 (20/20)	< 0.001	0 (0/20)	0 (0/20)
BCG Pasteur	0 (0/26)	<i>ns</i>	100 (26/26)	100 (26/26)
Bacterin	100 (8/8)	< 0.001	0 (0/8)	0 (0/8)

^a in parenthesis: survivors/total hamsters from two independent experiments.
^b P values were calculated by Fisher's exact test (two-tailed) for each experiment.
^c in parenthesis: number of positive kidneys/total samples.

rBCG:TBDRchi elicits a humoral immune response in hamsters

The immunogenicity of rBCG:TBDRchi was evaluated by indirect ELISA using rTBDRchi as the antigen. Hamsters vaccinated with rBCG:TBDRchi produced a significant anti-rTBDRchi IgG response ($P < 0.05$), detected 30 days after the second dose (Figure 3a), demonstrating the potential of rBCG:TBDRchi to promote a humoral immune response. In contrast, hamsters injected with BCG-Pasteur were negative for rTBDRchi IgG in the same ELISA. The presence of antibodies against each leptospiral TBDR in immunized hamsters was assessed by Western blotting (WB). Sera obtained from hamsters after immunization with rBCG:TBDRchi (1:200) detected two proteins in *L. interrogans* lysates, with apparent molecular weight of ~100 and ~85 kDa, likely corresponding to LIC10896 (99 kDa) and LIC10964 (87 kDa) (Figure 3b).

Discussion

Predicted surface-exposed fragments from three leptospiral TonB-dependent receptors were used in this work to construct the chimeric protein rTBDRchi. *L. interrogans* proteins LIC10896, LIC10964 and LIC12374 are putative TBDRs homologs (Nascimento et al. 2004; Louvel et al. 2006; Grassmann et al. 2017b). These observations were further supported in the current study, the 3D models of LIC10896, LIC10964 and LIC12374 presented high similarity to previously described TBDRs from other bacteria (Online Resource 1, Table S3). Among the three structural models, LIC12374 is the model with the highest RMSD when aligned to its closest structural analog in PDB, the cobalamin transporter BtuB from *E. coli* (Chimento et al. 2003). Previously reported as a leptospiral FecA ortholog, LIC10896 was modeled with high structural similarity to the FpvA pyoverdine-Fe transporter from *Pseudomonas aeruginosa* (Cobessi et al. 2005). Similarly, LIC10964 was previously associated with hemin metabolism in *Leptospira* spp., and our 3D model was highly similar to the *N. meningitidis* zinc transporter, ZnuD (Calmettes et al. 2015). Identification of specific functions for TBDRs through structural modelling is difficult and should be determined experimentally. Independently of the specific substrate for a TBDR, the 22 transmembrane β -strand barrel conformation is highly conserved among known TBDR structures, supporting our approach for the identification of surface-exposed regions.

While LIC10896 and LIC12374 have orthologs conserved in all pathogenic and saprophyte species, LIC10964 is present only in pathogenic species, as previously noted (Grassmann et al. 2017a). Host-adapted leptospire, cultivated within dialysis membrane chambers (DMCs), expressed higher levels of *LIC10964* compared to those cultivated *in vitro* (Caimano et al. 2014), suggesting a potential role during infection, in agreement with its predicted function as hemin transporter. The high level of sequence conservation of TBDRs across pathogenic leptospire makes this class of proteins an attractive target for development of cross-protective leptospirosis vaccines. In agreement with this notion, the TBDR TbpA from *Haemophilus parasuis* was able to induce a cross-protective immune response against three different serovars in a guinea pig model of Glasser's disease, with protection varying between 60 and 80% (Huang et al. 2013). Similarly, hyperimmune sera generated in mice and guinea pigs against *N. meningitidis* outer membrane vesicles overexpressing the TBDR ZnuD, presented a neutralizing effect on 14 different *N. meningitidis* strains *in vitro* (Hubert et al. 2013). Although we did not evaluate rBCG:TBDRchi in a heterologous challenge, all of the epitopes in the recombinant vaccine antigen were highly conserved in other pathogenic *Leptospira* spp. (Online Resource 3), enhancing the possibility of heterologous protection after immunization, a requisite for a universal leptospiral vaccine (Grassmann et al. 2017c).

Multi-epitope constructs based on different proteins have been reported for leptospirosis diagnosis and vaccine development. (Lin et al. 2016)) evaluated a recombinant protein containing four repeats of T and B cell epitopes in the leptospiral proteins OmpL1, LipL32 and LipL21. That chimeric protein also stimulated significant protection (4/5) in a guinea pig model of leptospirosis and induced a cross-reactive antibody response against various *L. interrogans* serogroups. (Fernandes et al. 2017), designed a chimeric protein based on sequences from five leptospiral proteins, however, this construction failed to induce significant protection in hamsters, where 50% survived, despite the presence of epitopes from LigA, a well-known protective antigen (Yan et al. 2009; Coutinho et al. 2011). To the best of our knowledge, this study pioneered the use of protein structural information on leptospiral β -OMPs to construct a recombinant molecule for use in an experimental vaccine against leptospirosis. In the current study, the humoral response generated by rBCG:TBDRchi was predominantly directed against two (LIC10896 and LIC10964) of the three proteins used in the antigen construction (Figure 3b). However, this does not rule out additional antibodies against LIC12374 that were below the limit of detection for the Western blot and might have contributed to the protection observed, given the presumable importance of LIC12374 epitopes in rTBDRchi. Different antigen presentation strategies for the selected epitopes should be assessed in the future,

towards the induction of strong antibody responses against all epitopes in rTBDRchi. Our approach for designing rTBDRchi antigen proved to be successful, given the protection we observed in the hamster model of acute leptospirosis.

Recombinant BCG vaccines have been shown to elicit both cellular and humoral immunity (Oliveira et al. 2017). In the current study, immunization with rBGG:TBDRchi induced a robust humoral immune responses against highly conserved surface-exposed epitopes from leptospiral TBDRs, as demonstrated by ELISA and Western blot. In contrast, in our previous study, we were unable to detect significant levels of antibodies by ELISA, in sera from animals immunized with a chimeric antigen containing fragments from the leptospiral lipoproteins LipL32, LigAni and LemA (Oliveira et al. 2019a; Dorneles et al. 2020). The rationale of using epitope-rich fragments may have contributed to the induction of humoral response obtained by rBGG:TBDRchi. As an extracellular bacterium, clearance of leptospires by the immune system seems to be related to bacterial opsonization followed by phagocytosis (Gomes-Solecki et al. 2017; Santecchia et al. 2020). Different studies have shown that opsonization is a requirement for efficient bactericidal activity of macrophages in leptospirosis (Johnson and Muschel 1966; Banfi et al. 1982; Wang et al. 1984; Santecchia et al. 2020). Studies on phagocytosis without opsonization showed that pathogenic leptospires have several mechanisms to survive and escape killing by phagocytic cells (Zhang et al. 2012; Hu et al. 2013; Zhao et al. 2013; Toma et al. 2014). It is accepted that opsonization with immune serum enhances *Leptospira* uptake and killing by human macrophages; the role of phagocytic cells during infection was recently reviewed (Santecchia et al. 2020). These observations support the importance of the humoral response promoted by rBCG:TBDRchi immunization in the clearance of virulent leptospires during early stages of infection.

The apparent absence of leptospires in the kidneys of rBCG:TBDRchi-immunized hamsters agrees with previous studies using BCG vectorized vaccines against leptospirosis (Oliveira et al. 2019a; Dorneles et al. 2020). In contrast, several subunit vaccines against leptospirosis have failed to induce sterilizing immunity (Felix et al. 2020). It is believed that a robust and effective cell-mediated response may contribute to limit the initial tissue colonization by *Leptospira* spp. and contribute towards bacterial clearance (Zuerner 2015; Santecchia et al. 2019). A BCG-induced trained immunity has been shown to decrease host susceptibility against different pathogens (Covián et al. 2019). Trained immunity is a proposed term to describe the enhancement of a non-specific immune response generated against unrelated infections after previous exposure to microbial fragments (Netea et al. 2011). This phenomenon has been observed after inoculation of mycobacterial components such as muramyl di-peptide, complete Freund's adjuvant and live BCG (Netea et al. 2011; Kleinnijenhuis et al. 2012). BCG vaccination increases IFN- γ production and other monocyte-derived cytokines, such as tumor necrosis factors (TNF) and IL-1 β , through an NOD2 pathway (Kleinnijenhuis et al. 2012). Interestingly, the effects of trained immunity on leptospirosis were previously described using CL429, a TLR2 and NOD2 pathways synthetic agonist (Santecchia et al. 2019). CL429-trained macrophages produced significantly more pro-inflammatory cytokines in response to pathogenic *Leptospira* than naïve macrophages, independently of B and T cells. The same trained macrophages also produced increased amounts of nitric oxide (Santecchia et al. 2019), a potent antimicrobial compound previously related to leptospiral clearance (Prêtre et al. 2011). Unfortunately, the lack of immunological reagents to characterize cellular immunity in the hamster model for acute leptospirosis limited in-depth analyses of cellular responses elicited by rBCG:TBDRchi immunization.

In conclusion, using a structural vaccinology approach, we designed a chimeric antigen that included predicted surface-exposed B and T cell epitopes from *L. interrogans* TBDRs. We used a BCG delivery system targeting robust immune response, resulting in production of a humoral immune response specific to the selected epitopes in leptospires. This vaccine formulation induced a protective and sterilizing immune response in the hamster model of lethal leptospirosis following a homologous challenge with *L. interrogans*.

Declarations

Author contributions: EB, JD, AG, AM, TO and OD contributed to conceive and design the study. EB, JD, AH, AM performed research throughout the work. JD, AH, AM, ASN performed animal experimentations. EB, AH, JD analyzed the data. EB, AG, AM wrote the paper. All authors contributed for manuscript review.

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Conflicts of interest: The authors declare no conflict of interest.

Data availability: *Leptospira interrogans* Fiocruz L1-130 strain is available as described in the Fiocruz/CLEP collection (WDCM 1012). Raw data from epitope mapping are available as Online Resource (ESM_3). Any additional data and materials used in this article are available upon request.

Ethics approval: All procedures involving manipulations of animals were approved by the Ethics Committee in Animal Experimentation (CEEA) of Universidade Federal de Pelotas (UFPEL) under protocol number 4646-2015. CEEA-UFPEL is accredited by the Brazilian National Council for Animal Experimentation Control (CONCEA).

References

1. Adler B (2015) Vaccines against leptospirosis. *Curr Top Microbiol Immunol* 387:251–272. doi:10.1007/978-3-662-45059-8_10
2. Alteri CJ, Hagan EC, Sivick KE, Smith SN, Mobley HLT (2009) Mucosal Immunization with Iron Receptor Antigens Protects against Urinary Tract Infection. *PLoS Pathogen* 5:e1000586. doi:10.1371/journal.ppat.1000586
3. Banfi E, Cinco M, Bellini M, Soranzo MR (1982) The role of antibodies and serum complement in the interaction between macrophages and leptospires. *J Gen Microbiol* 128:813–816. doi:10.1099/00221287-128-4-813

4. Caimano MJ, Sivasankaran SK, Allard A, Hurley D, Hokamp K, Grassmann A, Hinton JCD, Nally JE (2014) A Model System for Studying the Transcriptomic and Physiological Changes Associated with Mammalian Host-Adaptation by *Leptospira interrogans* Serovar Copenhageni. *PLoS Pathog* 10:e1004004. doi:10.1371/journal.ppat.1004004
5. Calmettes C, Ing C, Buckwalter CM, Bakkouri M, el, Lai CC, Moraes TF, Pogoutse A, Gray-owen SD (2015) The molecular mechanism of Zinc acquisition by the neisserial outer-membrane transporter ZnuD. *Nat Commun* 6:1–11. doi:10.1038/ncomms8996
6. Chimento DP, Kadner RJ, Wiener MC (2003) The *Escherichia coli* Outer Membrane Cobalamin Transporter BtuB: Structural Analysis of Calcium and Substrate Binding, and Identification of Orthologous Transporters by Sequence/Structure Conservation. *J Mol Biol* 332:999–1014. doi:https://doi.org/10.1016/j.jmb.2003.07.005
7. Cobessi D, Celia H, Folschweiller N, Schalk IJ, Abdallah MA, Pattus F (2005) The Crystal Structure of the Pyoverdine Outer Membrane Receptor FpvA from *Pseudomonas aeruginosa* at 3.6 Å Resolution. *J Mol Biol* 347:121–134. doi:10.1016/j.jmb.2005.01.021
8. Costa F, Hagan JE, Calcagno J, Kane M, Torgerson P, Martinez-silveira MS, Stein C, Abela-ridder B, Ko AI (2015) Global Morbidity and Mortality of Leptospirosis: A Systematic Review. *PLoS Negl Trop Dis* 9:e0003898. doi:10.1371/journal.pntd.0003898
9. Coutinho ML, Choy HA, Kelley MM, Matsunaga J, Babbitt JT, Lewis MS, Aleixo JAG, Haake DA (2011) A LigA Three-Domain Region Protects Hamsters from Lethal Infection by *Leptospira interrogans*. *PLoS Negl Trop Dis* 5:1–10. doi:10.1371/journal.pntd.0001422
10. Covián C, Fernández-Fierro A, Retamal-Díaz A, Díaz FE, Vasquez AE, Lay MK, Riedel CA, González PA, Bueno SM, Kalergis AM (2019) BCG-Induced Cross-Protection and Development of Trained Immunity: Implication for Vaccine Design. *Front Immunol* 10:1–14. doi:10.3389/fimmu.2019.02806
11. da Cunha CEP, Bettin EB, Bakry AFAAY, Seixas Neto ACP, Amaral MG, Dellagostin OA (2019) Evaluation of different strategies to promote a protective immune response against leptospirosis using a recombinant LigA and LigB chimera. *Vaccine* 37:1844–1852. doi:10.1016/j.vaccine.2019.02.010
12. Dellagostin OA, Grassmann AA, Rizzi C, Schuch RA, Jorge S, Oliveira TL, McBride AJA, Hartwig DD (2017) Reverse Vaccinology: An Approach for Identifying Leptospiral Vaccine Candidates. *Int J Mol Sci* 18:158. doi:10.3390/ijms18010158
13. Dockrell HM, Smith SG (2017) What have we learnt about BCG vaccination in the last 20 years? *Front Immunol* 8. doi:10.3389/fimmu.2017.01134
14. Dorneles J, Bonemann A, Clair A, Seixas P, Rizzi C, Burlamarque É, Silva A, Caetano C, Castro D, Gevehr C, Larré T, Antonio O (2020) Protection against leptospirosis conferred by *Mycobacterium bovis* BCG expressing antigens from *Leptospira interrogans*. *Vaccine* 38:8136–8144. doi:10.1016/j.vaccine.2020.10.086
15. Edgar RC, Drive RM, Valley M (2004) MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res* 32:1792–1797. doi:10.1093/nar/gkh340
16. Ellis WA (2015) Animal leptospirosis. *Curr Top Microbiol Immunol* 387:99–137. doi:10.1007/978-3-662-45059-8_6
17. Faine S (1959) Iron as a Growth Requirement for Pathogenic *Leptospira*. *J Gen Microbiol* 20:246–251. doi:10.1099/00221287-20-2-246
18. Felix CR, Siedler BS, Barbosa LN, Timm GR, McFadden J, McBride AJA (2020) An overview of human leptospirosis vaccine design and future perspectives. *Expert Opin Drug Discov* 15:179–188. doi:10.1080/17460441.2020.1694508
19. Fernandes LGV, Teixeira AF, Filho AFS, Souza GO, Vasconcellos SA, Heinemann MB, Romero EC, Nascimento ALTO (2017) Immune response and protective profile elicited by a multi-epitope chimeric protein derived from *Leptospira interrogans*. *International Journal of Infectious Diseases* 57:61–69. doi:10.1016/j.ijid.2017.01.032
20. Fouts DE, Matthias MA, Adhikarla H, Adler B, Amorim-Santos L, Berg DE, Bulach D, Buschiazzi A, Chang YF, Galloway RL, Haake DA, Haft DH, Hartskeerl R, Ko AI, Levett PN, Matsunaga J, Mechaly AE, Monk JM, Nascimento ALT, Nelson KE, Palsson B, Peacock SJ, Picardeau M, Ricaldi JN, Thaipandungpanit J, Wunder EA, Yang XF, Zhang JJ, Vinetz JM, Derrick E, Michael A, Fouts DE, Matthias MA, Adhikarla H, Adler B, Amorim- L (2016) What Makes a Bacterial Species Pathogenic?:Comparative Genomic Analysis of the Genus *Leptospira*. *PLoS Negl Trop Dis* 10:1–57. doi:10.1371/journal.pntd.0004403
21. Gomes-Solecki M, Santecchia I, Werts C (2017) Animal models of leptospirosis: Of mice and hamsters. *Front Immunol* 8. doi:10.3389/fimmu.2017.00058
22. Grassmann AA, Kremer FS, dos Santos JC, Souza JD, Pinto L, McBride S, Gomes-solecki AAJA, John M, McBride A (2017a) AAJA Discovery of Novel Leptospirosis Vaccine Candidates Using Reverse and Structural Vaccinology. *Frontiers in Immunology* 8:463. doi:10.3389/fimmu.2017.00463
23. Grassmann AA, Kremer FS, dos Santos JC, Souza JD, Pinto L, da McBride S, Gomes-solecki AJAAJA, John M, McBride A, Santos AJAAJA, Souza JC, Pinto JD, da McBride L (2017b) AJAAJA Discovery of Novel Leptospirosis Vaccine Candidates Using Reverse and Structural Vaccinology. *Frontiers in Immunology* 8:463. doi:10.3389/fimmu.2017.00463
24. Grassmann AA, Souza JD, John A, McBride AJA, John A, McBride AJA (2017c) A universal vaccine against leptospirosis: Are we going in the right direction? *Front Immunol* 8:1–11. doi:10.3389/fimmu.2017.00256
25. Gröschel MI, Sayes F, Shin SJ, Frigui W, Pawlik A, Orgeur M, Canetti R, Honoré N, Simeone R, van der Werf TS, Bitter W, Cho SN, Majlessi L, Brosch R (2017) Recombinant BCG Expressing ESX-1 of *Mycobacterium marinum* Combines Low Virulence with Cytosolic Immune Signaling and Improved TB Protection. *Cell Reports* 18:2752–2765. doi:10.1016/j.celrep.2017.02.057
26. Groshong AM, Grassmann AA, Luthra A, McLain MA, Provatas AA, Radolf JD, Caimano MJ (2021) PlzA is a bifunctional c-di-GMP biosensor that promotes tick and mammalian hostadaptation of *Borrelia burgdorferi*. *PLoS Pathog* 17:1–31. doi:10.1371/journal.ppat.1009725
27. Haake DA, Levett PN (2015) Leptospirosis in Humans. *Curr Top Microbiol Immunol* 387:65–97. doi:10.1007/978-3-662-45059-8
28. Hu W, Ge Y, Ojcius DM, Sun D, Dong H, Yang XF, Yan J (2013) p53 controls apoptosis in *Leptospira*-infected macrophages. *Cell Microbiol* 15:1642–1659. doi:10.1111/cmi.12141
29. Hu Y, Dang W, Sun L (2012) A TonB-dependent outer membrane receptor of *Pseudomonas fluorescens*: virulence and vaccine potential. *Arch Microbiol* 194:795–802. doi:10.1007/s00203-012-0812-3

30. Huang X, Li Y, Fu Y, Ji Y, Lian K, Zheng H, Wei J, Cai X (2013) Cross-Protective Efficacy of Recombinant Transferrin-Binding Protein A of *Haemophilus parasuis* in Guinea Pigs. *Clin Vaccine Immunol* 20:912–919. doi:10.1128/CVI.00621-12
31. Hubert K, Devos N, Mordhorst I, Tans C, Baudoux G, Feron C, Goraj K, Tommassen J, Vogel U, Poolman JT (2013) ZnuD, a Potential Candidate for a Simple and Universal *Neisseria meningitidis* Vaccine. *Infect Immun* 81:1915–1927. doi:10.1128/IAI.01312-12
32. Johnson RC, Muschel LH (1966) Antileptospiral activity of serum. I. Normal and immune serum. *J Bacteriol* 91:1403–1409. doi:10.1128/jb.91.4.1403-1409.1966
33. Kleinnijenhuis J, Quintin J, Preijers F, Joosten LAB, Ifrim DC, Saeed S, Jacobs C, Van Loenhout J, De Jong D, Hendrik S, Xavier RJ, Van Der Meer JWM, Van Crevel R, Netea MG (2012) Bacille Calmette-Guérin induces NOD2-dependent nonspecific protection from reinfection via epigenetic reprogramming of monocytes. *Proc Natl Acad Sci USA* 109:17537–17542. doi:10.1073/pnas.1202870109
34. Larsen JEP, Lund O, Nielsen M (2006) Improved method for predicting linear B-cell epitopes. *Immunome Research* 2:2. doi:10.1186/1745-7580-2-2
35. Lin X, Xiao G, Luo D, Kong L, Chen X, Sun D, Yan J (2016) Chimeric epitope vaccine against *Leptospira interrogans* infection and induced specific immunity in guinea pigs. *BMC Microbiol* 16. doi:10.1186/s12866-016-0852-y
36. Liu J, Ma Q, Yang F, Zhu R, Gu J, Sun C, Feng X, Du C, Langford PR, Han W, Yang J, Lei L (2017) B cell cross-epitope of *Propionibacterium acnes* and *Actinobacillus pleuropneumonia* selected by phage display library can efficiently protect from *Actinobacillus pleuropneumonia* infection. *Vet Microbiol* 205:14–21. doi:10.1016/j.vetmic.2017.04.026
37. Lobo N, Brooks NA, Zlotta AR, Cirillo JD, Boorjian S, Black PC, Meeks JJ, Bivalacqua TJ, Gontero P, Steinberg GD, McConkey D, Babjuk M, Alfred Witjes J, Kamat AM (2021) 100 years of Bacillus Calmette–Guérin immunotherapy: from cattle to COVID-19. *Nature Reviews Urology* 18:611–622. doi:10.1038/s41585-021-00481-1
38. Lomize MA, Pogozheva ID, Joo H, Mosberg HI, Lomize AL (2012) OPM database and PPM web server: Resources for positioning of proteins in membranes. *Nucleic Acids Res* 40:370–376. doi:10.1093/nar/gkr703
39. Louvel H, Bommezzadri S, Zidane N, Boursaux-Eude C, Creno S, Magnier A, Rouy Z, Médigue C, Saint Girons I, Bouchier C, Picardeau M (2006) Comparative and functional genomic analyses of iron transport and regulation in *Leptospira* spp. *J Bacteriol* 188:7893–7904. doi:10.1128/JB.00711-06
40. Nascimento ALTO, Ko AI, Martins EAL, Monteiro-Vitorello CB, Ho PL, Haake DA, Verjovski-Almeida S, Hartskeerl RA, Marques MV, Oliveira MC, Menck CFM, Leite LCC, Carrer H, Coutinho LL, Degraive WM, Dellagostin OA, El-Dorry H, Ferro ES, Ferro MIT, Furlan LR, Gamberini M, Gigliotti EA, Góes-Neto A, Goldman GH, Goldman MHS, Harakava R, Jerônimo SMB, Junqueira-De-Azevedo ILM, Kimura ET, Kuramae EE, Lemos EGM, Lemos MVF, Marino CL, Nunes LR, De Oliveira RC, Pereira GG, Reis MS, Schriefer A, Siqueira WJ, Sommer P, Tsai SM, Simpson AJG, Ferro JA, Camargo LEA, Kitajima JP, Setubal JC, Van Sluys MA (2004) Comparative Genomics of Two *Leptospira interrogans* Serovars Reveals Novel Insights into Physiology and Pathogenesis. *J Bacteriol* 186:2164–2172. doi:10.1128/JB.186.7.2164-2172.2004
41. Netea MG, Quintin J, Van Der Meer JWM (2011) Trained immunity: A memory for innate host defense. *Cell Host and Microbe* 9:355–361. doi:10.1016/j.chom.2011.04.006
42. Nielsen M, Lund O (2009) NN-align. An artificial neural network-based alignment algorithm for MHC class II peptide binding prediction. *BMC Bioinformatics* 10:296. doi:10.1186/1471-2105-10-296
43. Noinaj N, Guillier M, Barnard TJ, Buchanan SK (2010) TonB-Dependent Transporters: Regulation, Structure, and Function. *Annu Rev Microbiol* 43–60. doi:10.1146/annurev.micro.112408.134247
44. Nosrati M, Hajizade A, Nazarian S, Amani J, Namvar Vansofla A, Tarverdizadeh Y (2019) Designing a multi-epitope vaccine for cross-protection against *Shigella* spp: An immunoinformatics and structural vaccinology study. *Mol Immunol* 116:106–116. doi:10.1016/j.molimm.2019.09.018
45. Oliveira TL, Rizzi C, da Cunha CEP, Dorneles J, Seixas Neto ACP, Amaral MG, Hartwig DD, Dellagostin OA (2019a) Recombinant BCG strains expressing chimeric proteins derived from *Leptospira* protect hamsters against leptospirosis. *Vaccine* 37:776–782. doi:10.1016/j.vaccine.2018.12.050
46. Oliveira TL, Rizzi C, Dellagostin OA (2017) Recombinant BCG vaccines: molecular features and their influence in the expression of foreign genes. *Appl Microbiol Biotechnol* 101:6865–6877. doi:10.1007/s00253-017-8439-6
47. Oliveira TL, Stedman A, Rizzi C, Dorneles J, da Cunha CEP, Junior ASV, Dellagostin OA, McFadden J (2019b) A standardized BioBrick toolbox for the assembly of sequences in mycobacteria. *Tuberculosis* 119:101851. doi:10.1016/j.tube.2019.07.002
48. Oyarzún P, Kobe B (2016) Recombinant and epitope-based vaccines on the road to the market and implications for vaccine design and production. *Human Vaccines & Immunotherapeutics* 12:763–767. doi:10.1080/21645515.2015.1094595
49. Parish T, Stoker NG (1995) Electroporation of mycobacteria. *Methods in molecular biology*. (Clifton, NJ) 47:237–252. doi:10.1385/0-89603-310-4:237
50. Parvizpour S, Pourseif MM, Razmara J, Rafi MA, Omid Y (2020) Epitope-based vaccine design: a comprehensive overview of bioinformatics approaches. *Drug Discovery Today* 25:1034–1042. doi:10.1016/j.drudis.2020.03.006
51. Pettersen EF, Goddard TD, Huang CC, Couch GS, Greenblatt DM, Meng EC, Ferrin TE (2004) UCSF Chimera – A Visualization System for Exploratory Research and Analysis. *J Comput Chem* 1605–1612. doi:10.1002/jcc.20084
52. Prêtre G, Olivera N, Cédola M, Haase S, Alberdi L, Brihuega B, Gómez RM (2011) Role of inducible nitric oxide synthase in the pathogenesis of experimental leptospirosis. *Microb Pathog* 51:203–208. doi:10.1016/j.micpath.2011.03.011
53. Ramos CRR, Abreu PAE, Nascimento ALTO, Ho PL (2004) A high-copy T7 *Escherichia coli* expression vector for the production of recombinant proteins with a minimal N-terminal his-tagged fusion peptide. *Braz J Med Biol Res* 37:1103–1109. doi:10.1590/S0100-879X2004000800001
54. Sanchez-trincado JL, Gomez-perosanz M, Reche PA (2017) Fundamentals and Methods for T- and B-Cell Epitope Prediction. *Journal of Immunology Research* 2017. doi:doi.org/10.1155/2017/2680160

55. Santecchia I, Ferrer F, Vieira ML, Mart R (2020) Phagocyte Escape of *Leptospira*: The Role of TLRs and NLRs. *Front Immunol* 11. doi:10.3389/fimmu.2020.571816
56. Santecchia I, Vernel-Pauillac F, Rasid O, Quintin J, Gomes-Solecki M, Boneca IG, Werts C (2019) Innate immune memory through TLR2 and NOD2 contributes to the control of *Leptospira interrogans* infection. *PLoS Pathog* 15:1–26. doi:10.1371/journal.ppat.1007811
57. Seixas F, Drawanz D, Maia G, Amaral M, Quevedo M (2007) Recombinant *Mycobacterium bovis* BCG expressing the LipL32 antigen of *Leptospira interrogans* protects hamsters from challenge. *Vaccine* 26:88–95. doi:10.1016/j.vaccine.2007.10.052
58. Shelton AN, Seth EC, Mok KC, Han AW, Jackson SN, Haft DR, Taga ME (2019) Uneven distribution of cobamide biosynthesis and dependence in bacteria predicted by comparative genomics. *ISME J* 13:789–804. doi:doi.org/10.1038/s41396-018-0304-9
59. Soto JA, Gálvez NMS, Rivera CA, Palavecino CE, Céspedes PF, Rey-Jurado E, Bueno SM, Kalergis AM (2018) Recombinant BCG Vaccines Reduce Pneumovirus-Caused Airway Pathology by Inducing Protective Humoral Immunity. *Front Immunol* 9:2875. doi:10.3389/fimmu.2018.02875
60. Toma C, Murray GL, Nohara T, Mizuyama M, Koizumi N, Adler B, Suzuki T (2014) Leptospiral outer membrane protein LMB216 is involved in enhancement of phagocytic uptake by macrophages. *Cell Microbiol* 16:1366–1377. doi:10.1111/cmi.12296
61. Trunz BB, Fine P, Dye C (2006) Effect of BCG vaccination on childhood tuberculous meningitis and miliary tuberculosis worldwide: a meta-analysis and assessment of cost-effectiveness. *Lancet* 367:1173–1180. doi:10.1016/S0140-6736(06)68507-3
62. Vincent AT, Schiettekatte O, Goarant C, Neela VK, Bernet E, Thibeaux R, Ismail N, Khalid MKNM, Amran F, Masuzawa T, Nakao R, Korba AA, Bourhy P, Veyrier FJ, Picardeau M (2019) Revisiting the taxonomy and evolution of pathogenicity of the genus *Leptospira* through the prism of genomics. *PLoS Negl Trop Dis* 13. doi:10.1371/journal.pntd.0007270
63. Wang B, Sullivan JA, Sullivan GW, Mandell GL (1984) Role of specific antibody in interaction of leptospires with human monocytes and monocyte-derived macrophages. *Infect Immun* 46:809–813. doi:10.1128/iai.46.3.809-813.1984
64. Xiao J, Liu J, Bao C, Zhu R, Gu J, Sun C, Feng X, Du C, Han W, Li Y, Lei L (2020) Recombinant tandem epitope vaccination provides cross protection against *Actinobacillus pleuropneumoniae* challenge in mice. *AMB Express* 10:1–13. doi:10.1186/s13568-020-01051-1
65. Yan W, Faisal SM, McDonough SP, Divers TJ, Barr SC, Chang C, Pan M, Chang Y (2009) Immunogenicity and protective efficacy of recombinant *Leptospira* immunoglobulin-like protein B (rLigB) in a hamster challenge model. *Microbes Infect* 11:230–237. doi:10.1016/j.micinf.2008.11.008
66. Yang J, Zhang Y (2015) I-TASSER server: new development for protein structure and function predictions. *Nucleic Acids Res* 43:174–181. doi:10.1093/nar/gkv342
67. Zhang L, Zhang C, Ojcius DM, Sun D, Zhao J, Lin X, Li L, Yan J (2012) The mammalian cell entry (Mce) protein of pathogenic *Leptospira* species is responsible for RGD motif-dependent infection of cells and animals. *Mol Microbiol* 83:1006–1023. doi:10.1111/j.1365-2958.2012.07985.x
68. Zhang Y, Skolnick J (2005) TM-align: A protein structure alignment algorithm based on the TM-score. *Nucleic Acids Res* 33:2302–2309. doi:10.1093/nar/gki524
69. Zhao J, Chen H, Ojcius DM, Zhao X, Sun D, Ge Y, Zheng L, Lin X, Li L, Yan J (2013) Identification of *Leptospira interrogans* Phospholipase C as a Novel Virulence Factor Responsible for Intracellular Free Calcium Ion Elevation during Macrophage Death. *PLoS ONE* 8:e75652. doi:10.1371/journal.pone.0075652
70. Zuerner RL (2015) Host response to *Leptospira* infection. *Curr Top Microbiol Immunol* 387:223–250. doi:10.1007/978-3-662-45059-8_9

Figures

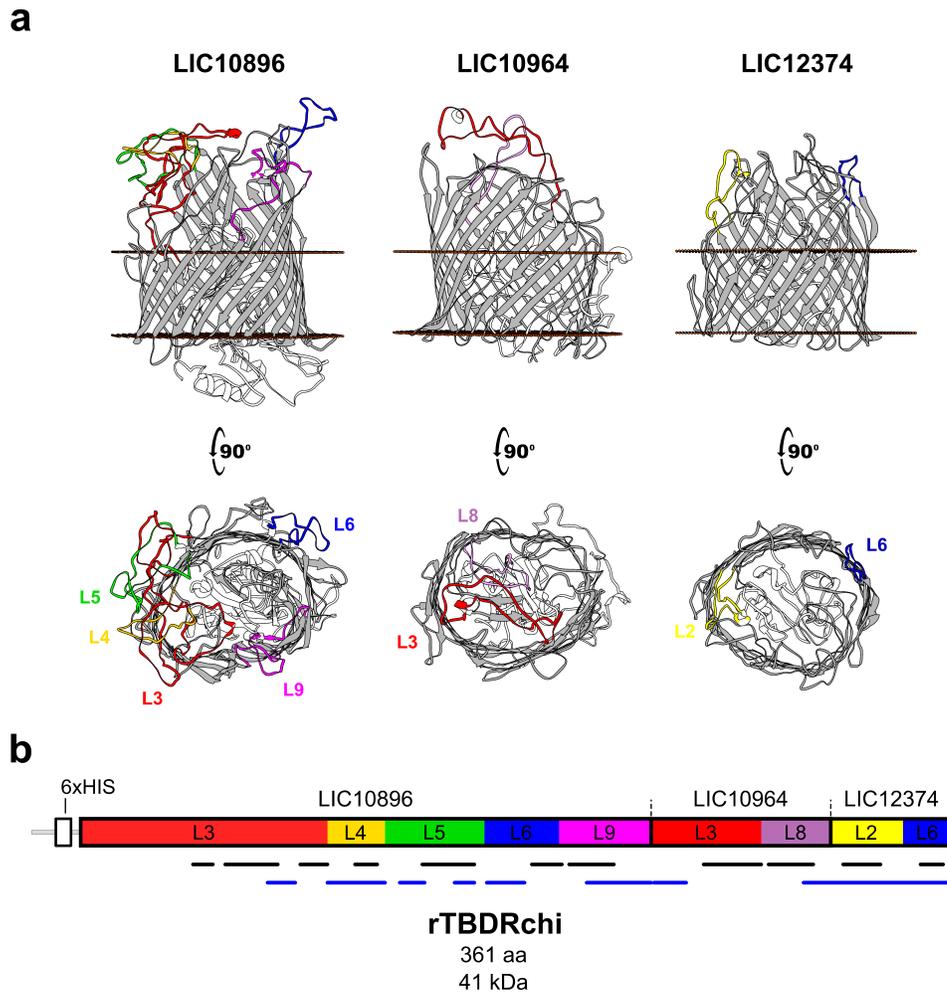


Figure 1

Epitope-mapping of leptospiral TBDRs and in silico construction of rTBDRchi. (a) CD4+ T cell epitopes and linear B cell epitopes were identified by NetMHCII and BepiPred servers, respectively. Epitopes were mapped on 3D structures generated by I-TASSER. Surface-exposed fragments containing identified epitopes (colored) were used to construct the recombinant fusion protein. (b) Nine identified fragments were selected to construct a 41 kDa recombinant fusion protein (rTBDRchi). rTBDRchi contains 11 CD4+ T-cell (black dashes) and 10 linear B-cell (blue dashes) epitopes throughout its sequence.

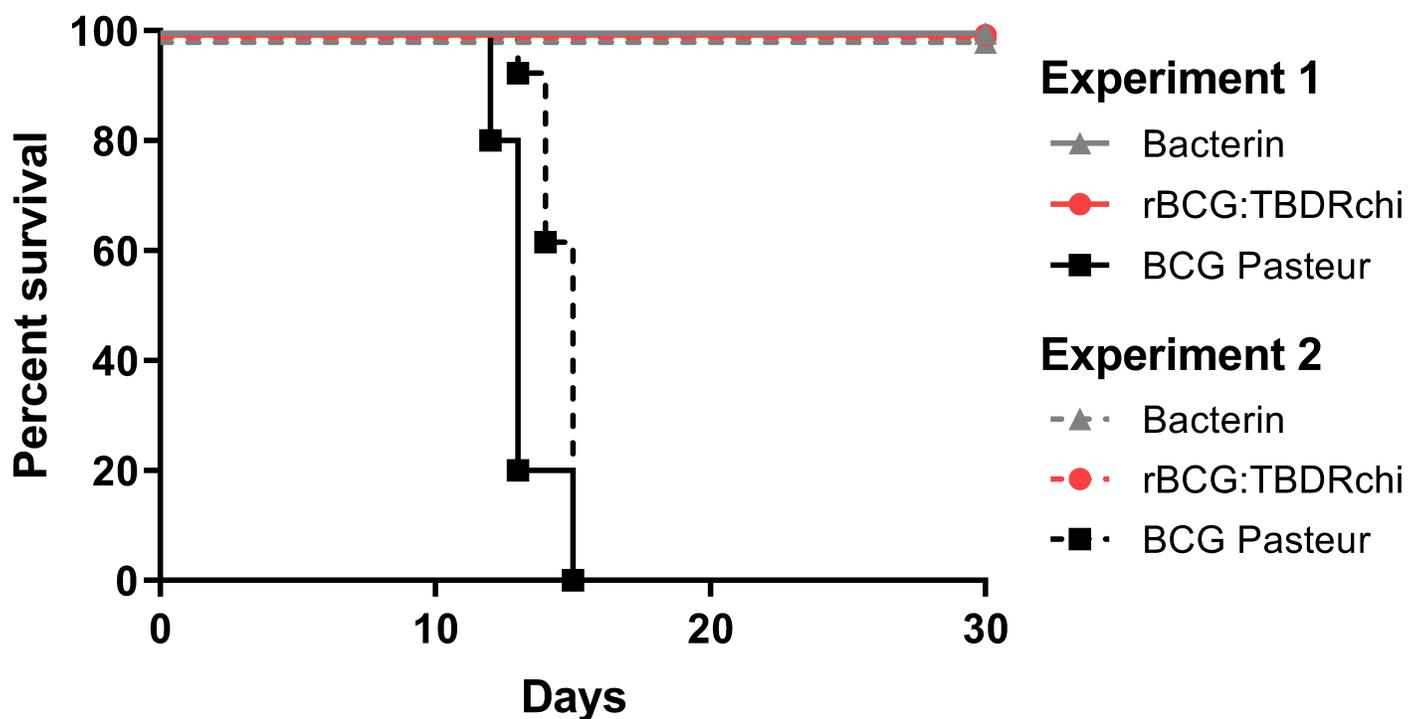
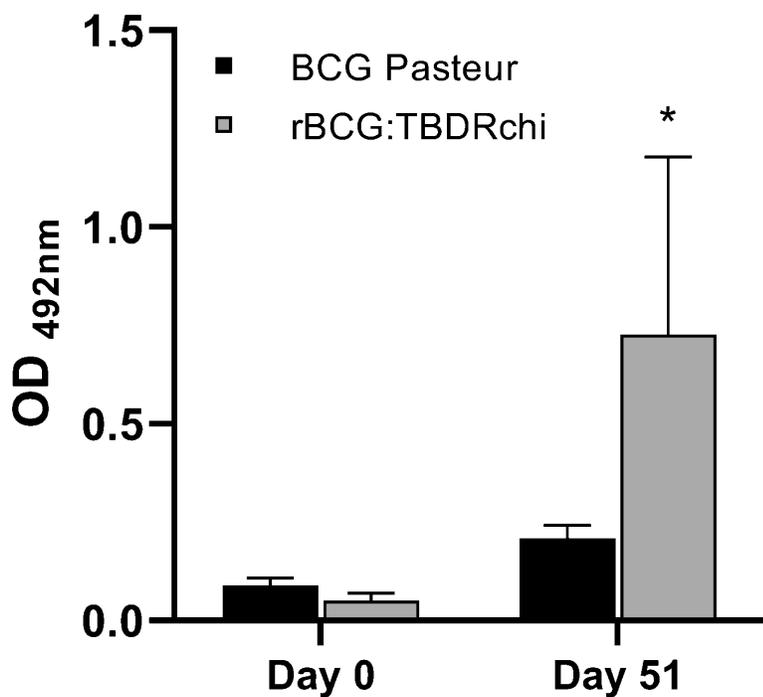


Figure 2
 Protection against lethal leptospirosis elicited by immunization with rBCG:TBDRchi. (a) Survival data from vaccinated hamster after challenge with an endpoint dose of *L. interrogans* sv. Copenhageni st. Fiocruz L1-130 in two independent experiments. All animals that were immunized subcutaneously with 106 recombinant BCG (rBCG:TBDRchi, n = 10), or 107 leptospire (bacterin, n = 4) survived ($p < 0.001$). Animals from the negative control group (BCG Pasteur, n = 13 per experiment) reached endpoint criteria for euthanasia between days 12-15 post-infection.

a



b

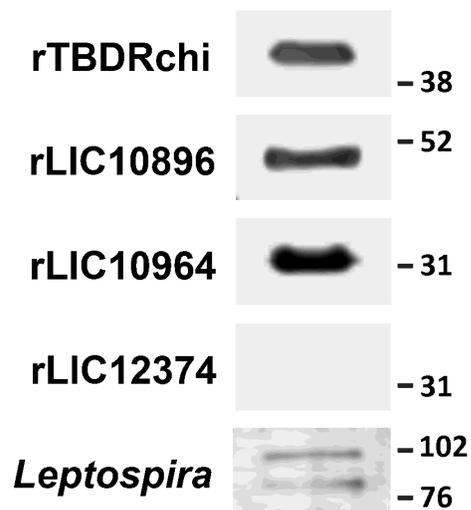


Figure 3

IgG antibody response in hamsters immunized with rBCG:TBDRchi evaluated by ELISA and immunoblot. (a) ELISA plates were coated with 1 µg of rTBDRchi. Sera from hamsters immunized with BCG Pasteur (negative control) and rBCG:TBDRchi, collected at day 0 (pre-immune) and 51 (pre-challenge) were evaluated at 1:50 dilution, in triplicate, with an anti-hamster IgG used as secondary antibody. Results are presented as mean absorbance ± standard deviation. Significant differences were determined by one-way ANOVA, followed by Tukey multiple comparison. Asterisk indicates a significance difference ($P < 0.05$) to other groups. (b) Immunoblot with rTBDRchi, recombinant TBDR proteins and *Leptospira* lysate using sera obtained from hamsters after immunization with rBCG:TBDRchi (1:200). Humoral immune response was primarily direct against two of the leptospiral TBDRs, LIC10896 and LIC10964.

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